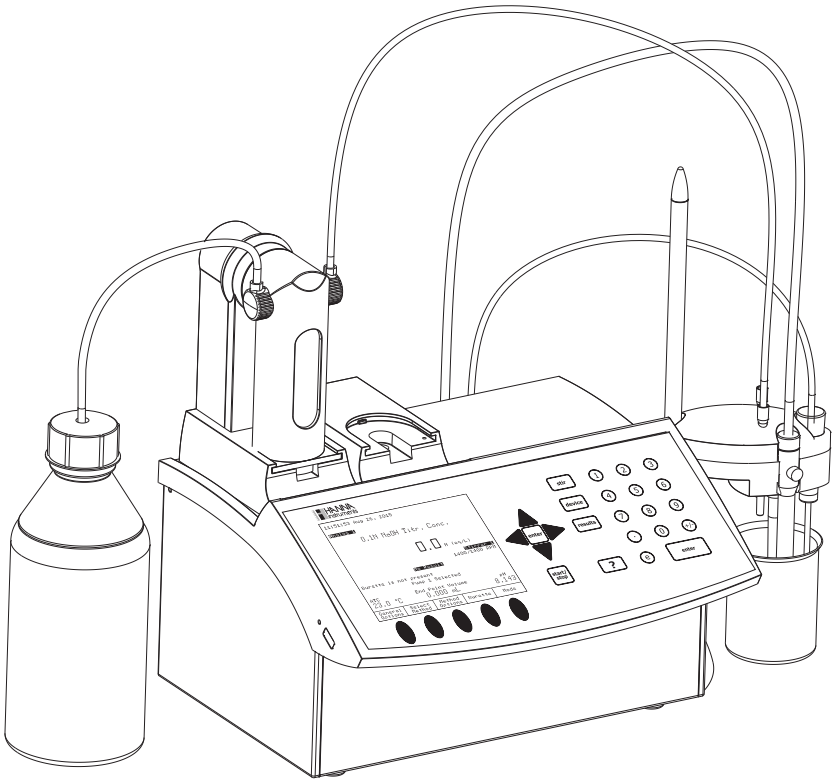
QUICK START GUIDE

HI901 Wine

AUTOMATIC POTENTIOMETRIC TITRATOR

Revision 1.00



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QUICK START GUIDE

Dear customer,

Congratulations on choosing a Hanna Instruments product.

This guide has been written for **HI901W** Winetitrators with color display, USB interface, and software version **1.00** and higher.

Please read this Quick Start Guide carefully before using the instrument. This guide will provide you with the necessary information for the correct use of the instrument.

The purpose of this guide is to present a quick overview of setting up and using the instrument.

For detailed information illustrating the extensive capabilities of your Titrator, please refer to the Instruction Manual.

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INTRODUCTION

The **HI901W** automatic Titrator is designed to perform a wide variety of potentiometric titrations with high accuracy, flexibility and reproducibility, allowing the user to obtain both accurate results and high-speed analysis.
The Titrator can perform fixed endpoint or equivalence point titrations and direct measurements by measuring the pH/mV/ISE and temperature of the sample.
Reports and methods can be transferred to a PC via a USB storage device, saved to a USB storage device or printed directly from the Titrator. An external monitor and keyboard can also be attached for added convenience.

How can I find certain information?

- The **Quick Start Guide** will help the user learn how to operate the Titrator within a short period of time.
- The **Instruction Manual** provides a complete description of the operating principles (user interface, general options, methods, titration/direct reading mode, pH, mV and ISE mode, maintenance, etc.).
- The **Titration Theory** outlines the basic concepts of titration.
- The contextual **Help** screens contain detailed explanations of every screen.

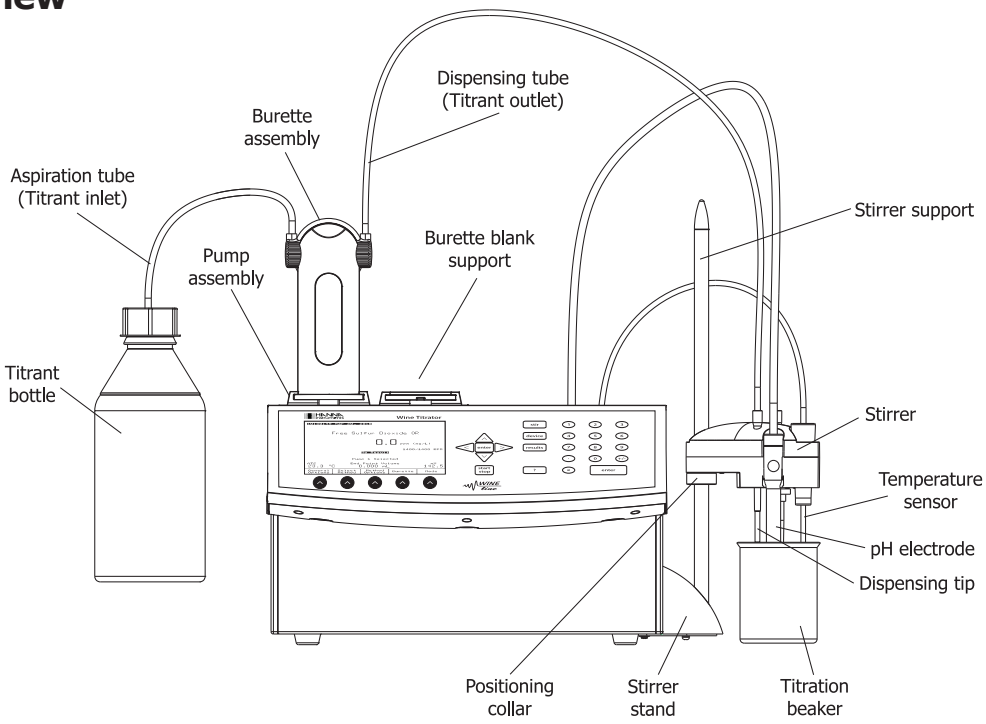
SAFETY MEASURES

The following safety measures must be followed:

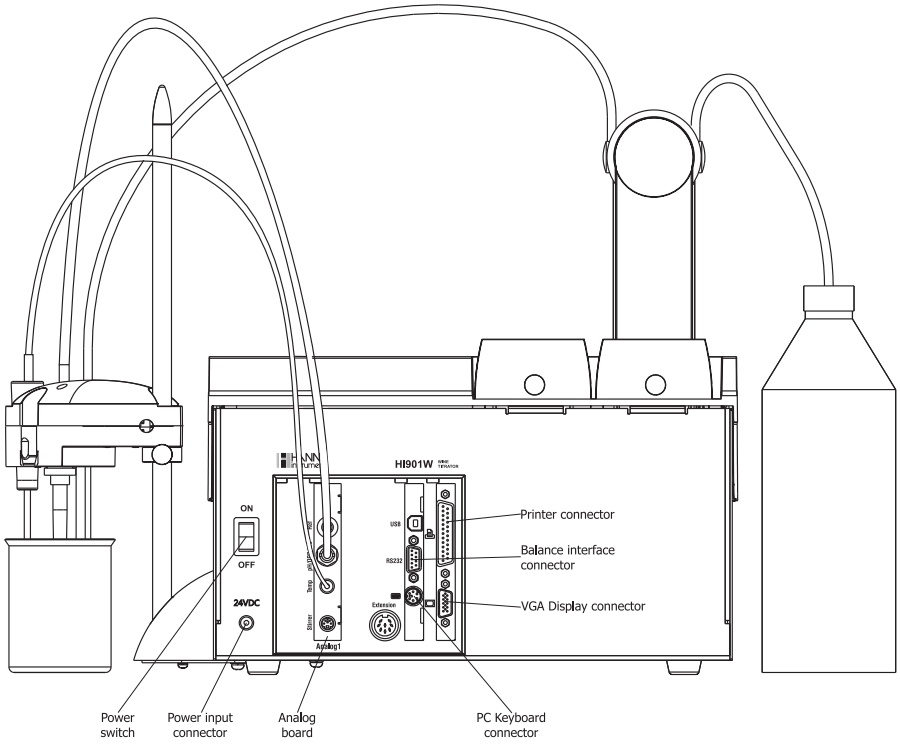
1. Never connect or disconnect the pump assembly or other peripheral with the Titrator turned on.
2. Verify that the burette and the attached tubing are assembled correctly.
3. Always check that the titrant bottle and the titration beaker are placed on a flat, stable surface.
4. Always wipe up spills and splashes immediately.
5. Avoid the following environmental working conditions:
    - Severe vibrations
    - Direct sunlight
    - Atmospheric relative humidity above 95% non-condensing
    - Environment temperatures below 10°C and above 40°C
    - Explosion hazards
6. Have the Titrator serviced by qualified service personnel only.

TITRATOR CONNECTIONS

Front View



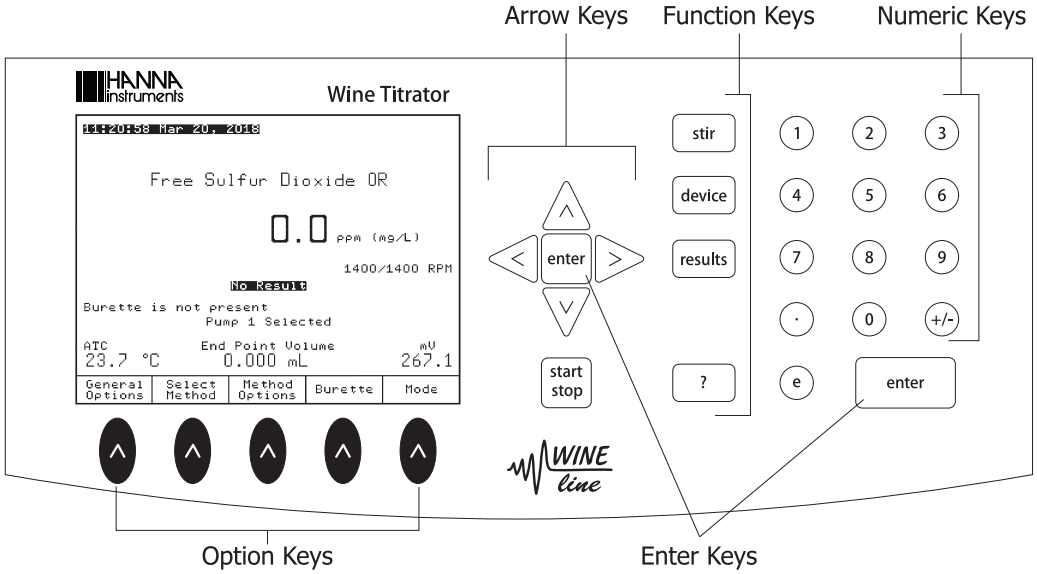
Rear View



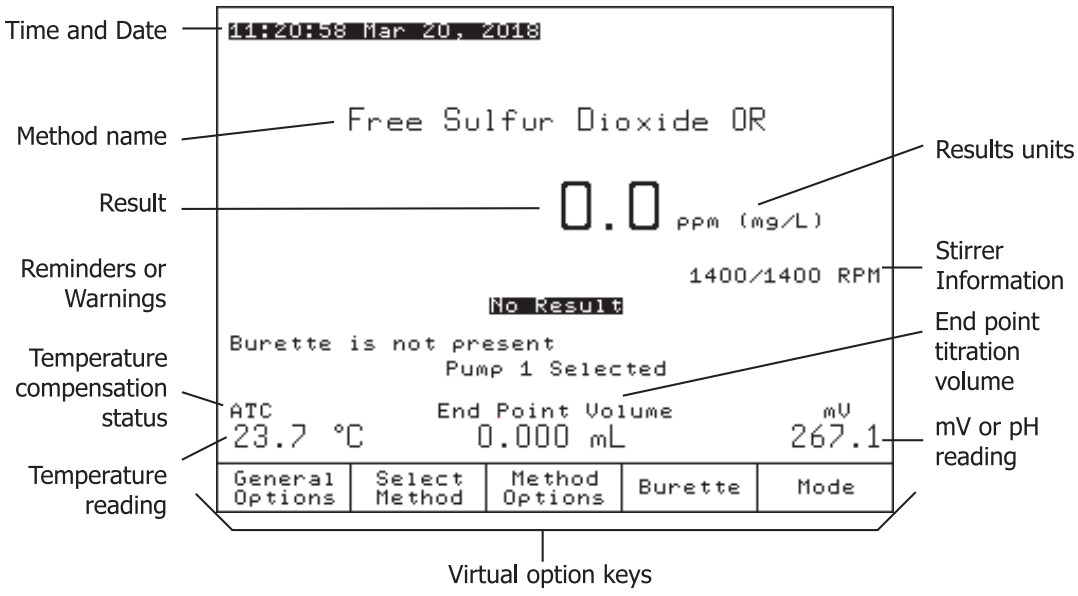
USER INTERFACE

Keypad

The titrator's keypad has 29 keys grouped in five categories, as follows:



Display



The user interface contains several screens. In each screen, many information fields are present at the same time. The information is displayed in an easy-to-read manner.
Virtual option keys describe the function performed when the corresponding option key is pressed.

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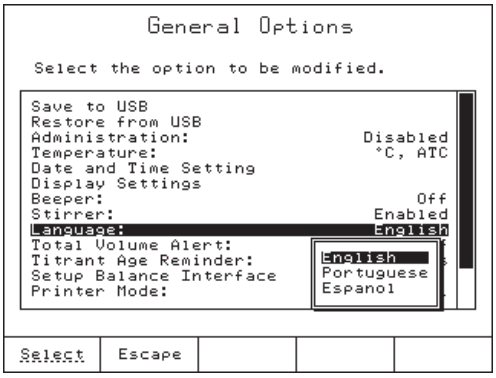
HOW TO SELECT YOUR LANGUAGE

To change the language, press General Options from the main screen. Highlight the *Language* option and then press Select. Using the △ and ▽ keys, select the language from the options listed in the ***Set Language*** screen and press Select.
Restart the Titrator in order to apply the new language setting.



HOW TO USE THE CONTEXTUAL HELP

Information about the Titrator can be easily accessed by pressing ?. The contextual help can be accessed at any time and it provides useful information about the current screen.

METHODS

The **HI901W** Titrator can store up to 100 methods (standard and user).

Standard Methods

Each Titrator is supplied with a package of standard methods for wine analysis.

User-Defined Methods

User defined methods allow the user to create and save their own methods. Each new method is based on an existing method which is altered to suit a specific application.

HOW TO CALIBRATE A pH ELECTRODE

To enter pH calibration mode, press [Mode], then [pH], then [pH Calibr.].

PREPARATION

Pour small quantities pH 4.01, pH 7.01 and pH 10.01 buffer solutions into clean beakers. If possible, use plastic beakers to minimize any EMC interferences.

For accurate calibration and to minimize cross-contamination, use two beakers for each buffer solution: one for rinsing the electrode and one for calibration.

CALIBRATION PROCEDURE

Three buffer entry types are available: Automatic, Semi-automatic and Manual Selection.

The default option is Manual Selection.

- If the instrument has been previously calibrated and calibration was not cleared, the old calibration can be cleared by pressing [Clear Cal].

Note: *It is very important to clear calibration history when a new electrode is used. Most errors and warning messages that appear during calibration depend on calibration history.*

- Use the [Next Buffer] or [Previous Buffer] to select pH 4.01 buffer solution.
- Use the second beaker of pH 4.01 buffer solution to rinse the pH electrode, temperature probe and propeller stirrer.
- Immerse the pH electrode, temperature probe and propeller stirrer in the pH 4.01 buffer solution. The pH electrode's bulb must be completely immersed in the buffer solution and the reference junction needs to be 5-6 mm below the surface. Add additional buffer if necessary.
- Press [stir] to turn on the propeller stirrer.
- Once the reading has stabilized, press [Accept] to update the calibration.
- Repeat this procedure for pH 7.01 and 10.01 buffer solutions.
- Press [Escape] to accept and exit pH calibration mode.

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HOW TO PERFORM A TITRATION

Required Solutions

- Titrant - 500 mL of 0.1 M (mol/L) Sodium Hydroxide (NaOH) in a titrant bottle.
- Sample - 0.1 mol/L Hydrochloric Acid (HCl).
- Distilled or deionized water.

Note: *Analytical grade reagents and water should be used for accurate results.*

Priming the Burette

- Insert the aspiration tube in the titrant bottle and the dispensing tube in a waste beaker.
- From the main screen press [Burette].
- Highlight the *Prime Burette* option and then press [Select].
- Enter the number of burette rinses. At least 3 rinses are recommended.
- Press [Accept] to start.
- The message "Executing..." will be displayed.

Note: *Make sure you have continuous liquid flow inside the burette. For accurate results, the aspiration tube, the dispensing tube and the syringe must be free of air bubbles.*

Method Selection

For this analysis, we will use the **HI1009EN Neutralization w/ NaOH**.
To select this method:

- Press [Select Method]. Use the △ and ▽ keys to highlight **HI1009EN Neutralization w/ NaOH**.
- Press [Select].

Setting Method Parameters

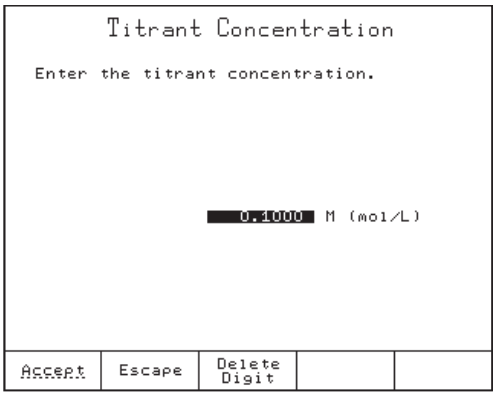
To display the method parameters, press Method Options. The ***View/Modify Method*** screen will be displayed.

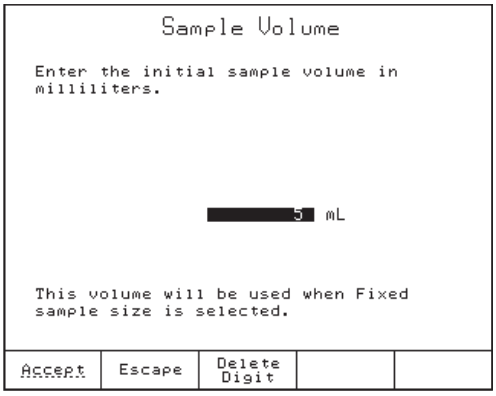
Only certain parameters can be changed.

For this titration, the NaOH titrant concentration and the size of the HCl sample need to be entered.

To accomplish this:

- Highlight *Titrant Conc.* option, then press Select. The ***Titrant Concentration*** screen will be displayed.
- Enter the correct value, then press Accept.
- Highlight *Analyte Size* option, then press Select.
- Enter the volume of the sample (e.g.: 5 mL), then press Accept.
- Press Escape, highlight *Save Method* option and then press Select.





Setup Titration Report

Users can select the information that is stored for each titration.
To setup the titration report, follow the procedure below:

- From the main screen, press results. The ***Data Parameters*** screen will be displayed.
- Highlight *Setup Titration Report* and press Select.
- Mark the fields to be included in the titration report with the "*" symbol. Use the △ and ▽ keys to highlight a field and Select / Unselect to toggle the field.
- Press Save Report to save the customized report .

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Preparing the Sample

- Add 50 to 65 mL of distilled / deionized water to the titration beaker.
- Use a pipette or burette to add 5.0 mL of the sample (0.1M Hydrochloric Acid (HCl)) into the same beaker.
- Slide the stirrer assembly up.
- Place the beaker under the stirrer assembly.
- Lower the stirrer assembly until it rests on the positioning collar.
- Adjust the height of the stirrer assembly so it is as close as possible to the bottom of the beaker.
- Adjust the level of the sample solution with distilled / deionized water so that the pH electrode bulb is completely immersed in the sample solution and the reference junction of the electrode is 5-6 mm below the surface.

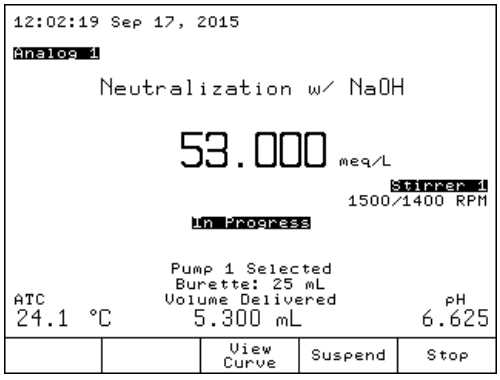
Note: *Make sure that the pH electrode, temperature probe and propeller do not touch each other or the beaker.*

Performing a Titration

- From the main screen, press start/stop. You will be prompted to enter the analyte size. Enter 5 mL and press enter. The Titrator will start the analysis.
- At the end of the titration, the message "Titration Completed" will appear on the display with the final concentration of the analyte in the sample and the equivalence endpoint volume.

Understanding the Displayed Information

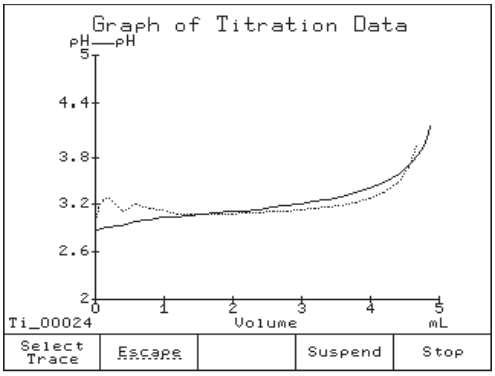
During a titration the following screen is displayed:

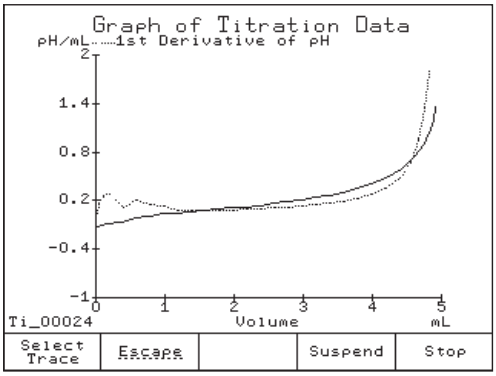


Viewing Graph During Titration

After a few doses are dispensed, [View Curve] will become active. Press [View Curve] to display the real-time titration graph.
The curves displayed are plots of the pH and the 1st derivative versus Titrant Volume (for details, see the Instruction Manual).

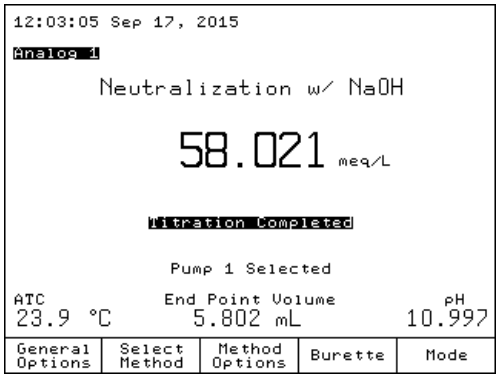
The two graphs are scaled to fit in the same screen window. Press [Select Trace] to change the y-axis scale to either the pH values or the 1st derivative values.

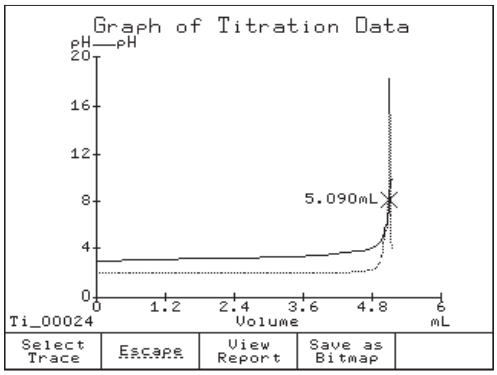




Titration Termination

The titration is normally terminated when the first equivalence endpoint is detected according to the selected algorithm. To ensure the correct detection and interpolation of the equivalence endpoint, the Titrator will dispense a few additional doses after the endpoint was reached.
The titration result can be displayed either in the main screen or in the ***Graph of Titration Data*** screen:





When the titration has ended, the Titrator will display the equivalence endpoint volume and the final concentration of the analyte together with the **Titration Completed** message.

To view the titration graph and/or results, press [results].

When the titration ends, an "x" will mark the endpoint on the pH versus titrant volume curve in the ***Graph of Titration Data*** screen. The value of the endpoint volume is also displayed next to the endpoint.

Results

The results obtained from a titration are stored in a report file that can be viewed, transferred to a USB Storage Device or PC and printed.

Viewing the last titration data

- From the main screen, press [results]. The ***Data Parameters*** screen will be displayed.
- From the ***Data Parameters*** screen highlight the *Review Last Analysis Report* option and press [Select]. The ***Review Result*** screen will be displayed.
- Use the [Page Up] and [Page Down] keys to display information related to the last titration performed. See *Titration Report* on next page.

Printing the titration report

Connect a DOS / Windows-compatible parallel printer directly to the DB 25-pin connector located on the back of the Titrator.

Note: *When connecting the printer, please turn off the Titrator and the printer.*

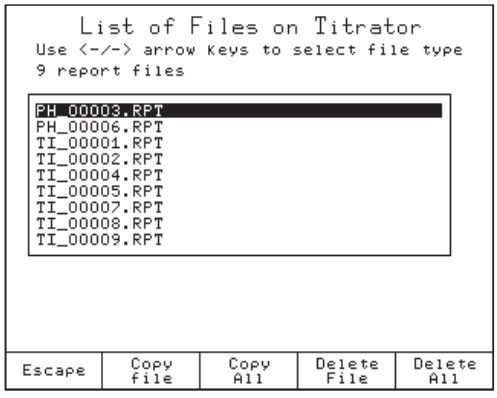
Printing out the report:

- From the ***Review Report*** screen, press [Print Report].
- During the information transfer to the printer, the message "Printing" will be displayed on the screen.
- Press [Escape] to return to the ***Data Parameters*** screen.
- Press [Escape] again to return to the main screen.

Saving data to USB Storage Device

This feature allows saving the results of titrations or pH / mV / ISE logging sessions on a USB storage device.

- From the main screen, press [General Options], the ***General Options*** screen will be displayed.
- Highlight the *Save Files to USB Storage Device* option using the △ and ▽ keys.
- Insert the USB storage device into the USB socket.
- Press [Select], the ***List of Files on Titrator*** screen will be displayed.
- Use the ◁ or ▷ keys to select the file type: "report files".
- Press [Copy All] to transfer all available reports to the USB storage device, or highlight the name of the report file to be transferred and press [Copy File].



- Transferring a report file will automatically transfer the corresponding log file and titration graph (*.BMP file if available).
- Press Escape to return to the ***General Options*** screen.
- Press Escape again to return to the main screen.

Titration report

While scrolling with the Page Up and Page Down keys, the fields below can be seen on the Titrator display or printed. The same information is available on the saved report file (Ti_00007.rpt in this example).

```
            HI901W - Titration Report

Method Name:                Neutralization w/NaOH
Time & Date:                 12:02:58 Mar 22, 2018
Report ID:                                Ti_00007

              Standardization Data
Buffer  Potential  Efficiency  Temperature
                  Time and Date
4.006pH   169.9mV      100.7%     22.0°C  A
              10:20 Mar 22, 2018
7.020pH    -7.8mV       96.5%     22.0°C  A
              10:23 Mar 22, 2018
10.040pH -178.6mV       96.5%     21.9°C  A
              10:25 Mar 22, 2018

          GLP & Instrumentation Data
Sample Name:                          Sample HCl-1
Company Name:                  Hanna Instruments
Operator Name:
Electrode Name:                   HI 1131  NO -2
Field 1:                               Any  text
Field 2:                               Any  text
Field 3:                               Any  text
Titrator Software Version                 v3.00
Base Board Software Version:              v2.00
Pump  1  Software  Version:                 v1.4
Base Board Serial Number:                01040409
Analog Board Serial Number:              30040409
Pump 1 Serial Number:                    70040207
Factory Calibration Date:             Jan 28, 2018

               Method Parameters
Name:                       Neutralization w/NaOH
Method Revision:                              1.0
Analog Board:                            Analog1
```

```
Stirrer Configuration:
        Stirrer:                        Stirrer 1
        Stirring Speed:                  1400 RPM
Pump Configuration:
   Titrant Pump :                       Pump   1
Dosing Type:                            Dynamic
   Min Vol:                              0.050 mL
   Max Vol:                              0.500 mL
   delta E:                             20.000 mV
End Point Mode:              pH 1EQ point, 1st Der
Recognition Options:
   Threshold:                           50 mV/mL
   Range:                                     NO
   Filtered Derivatives:                      NO
Pre-Titration Volume:                   0.000 mL
Pre-Titration Stir Time:                  15 Sec
Measurement Mode:                Signal Stability
   delta E:                               1.0 mV
   delta t:                                2 Sec
   Min wait:                               2 Sec
   Max wait:                              15 Sec
Electrode Type:                              pH
Calculations:               Sample Calc. by Volume
Dilution  Option:                       Disabled
Titrant Name:                                NaOH
Titrant Conc.:                  0.1000 M (mol/L)
Analyte Size:                           5.000 mL
Analyte Entry:                            Manual
Maximum Titrant Volume:                20.000 mL
Stirring Speed:                         1400 RPM
Potential Range:             -2000.0 to 2000.0 mV
Volume/Flow Rate:             25 mL / 50.0 mL/min
Signal Averaging:                      1 Reading
Significant  Figures:                      XXXXX
M (mol/L) —> M (mol/L)

V mol mol
-*—*——
   L  mol
——————

mL L
—*———
  1000mL

V = volume dispensed in liters
0.100 mol/L -> titrant conc.
1.000 mol/mol -> (sample/titrant)
5.000 mL -> sample volume

Nr Volume[ml]  mV        pH         Graphic  Temp[°C]     Time
0    0.000        235.2   2.857      0.0    19.1     A    00:00:00
1    0.050        234.6   2.866    -10.2    19.0     A    00:00:21
2    0.100        233.9   2.880    -15.8    19.1     A    00:00:27
3    0.200        232.2   2.908    -16.7    19.1     A    00:00:39
4    0.390        231.1   2.928     -6.0    19.1     A    00:00:45
```

5	0.590	228.6	2.970	-12.3	19.1	A	00:01:04
6	0.790	226.9	3.000	-8.7	19.1	A	00:01:20
7	0.990	225.5	3.024	-6.9	19.1	A	00:01:37
8	1.190	224.7	3.038	-4.0	19.1	A	00:01:43
9	1.390	223.9	3.051	-4.0	19.1	A	00:01:49
10	1.590	223.0	3.066	-4.3	19.1	A	00:01:55
11	1.790	222.1	3.082	-4.6	19.1	A	00:02:01
12	1.990	221.2	3.098	-4.6	19.1	A	00:02:06
13	2.190	220.1	3.115	-5.1	19.1	A	00:02:11
14	2.390	219.0	3.134	-5.6	19.1	A	00:02:17
15	2.590	217.8	3.155	-6.0	19.1	A	00:02:23
16	2.790	216.5	3.177	-6.6	19.1	A	00:02:29
17	2.990	215.1	3.202	-7.3	19.1	A	00:02:34
18	3.190	213.4	3.231	-8.4	19.1	A	00:02:40
19	3.390	211.5	3.263	-9.3	19.1	A	00:02:46
20	3.590	209.2	3.302	-11.4	19.1	A	00:02:51
21	3.790	206.6	3.348	-13.4	19.1	A	00:02:57
22	3.990	203.2	3.406	-16.8	19.1	A	00:03:02
23	4.190	198.9	3.479	-21.4	19.1	A	00:03:08
24	4.390	193.1	3.578	-29.0	19.1	A	00:03:14
25	4.556	186.2	3.697	-41.7	19.1	A	00:03:20
26	4.670	179.6	3.810	-57.8	19.1	A	00:03:25
27	4.753	172.9	3.925	-81.2	19.1	A	00:03:31
28	4.812	166.4	4.036	-110.0	19.2	A	00:03:37
29	4.856	160.1	4.144	-143.5	19.2	A	00:03:43
30	4.889	153.7	4.253	-189.9	19.2	A	00:03:54
31	4.915	147.1	4.367	-259.9	19.2	A	00:04:00
32	4.934	141.0	4.471	-322.7	19.2	A	00:04:11
33	4.949	135.2	4.571	-388.0	19.2	A	00:04:17
34	4.964	127.5	4.702	-512.0	19.2	A	00:04:23
35	4.979	117.3	4.877	-680.0	19.2	A	00:04:29
36	4.994	104.2	5.102	-875.3	19.2	A	00:04:35
37	5.009	87.9	5.381	-1088.0	19.2	A	00:04:41
38	5.024	69.6	5.695	-1221.3	19.2	A	00:04:50
39	5.039	51.2	6.010	-1226.0	19.2	A	00:05:08
40	5.054	31.6	6.344	-1301.3	19.2	A	00:05:36
41	5.069	7.3	6.762	-1625.3	19.2	A	00:06:07
42	5.084	-37.9	7.557	-3010.0	19.2	A	00:06:38
43	5.099	-120.0	9.024	-5476.0	19.2	A	00:06:48
44	5.114	-144.7	9.464	-1642.7	19.2	A	00:06:54
45	5.129	-158.2	9.705	-900.7	19.2	A	00:07:01
46	5.144	-168.1	9.883	-664.0	19.2	A	00:07:08

Titration Results

Method Name:	Neutralization w/NaOH
Time & Date:	12:02:58 Mar 22, 2018
Analyte Size:	5.000 mL
End Point Volume:	5.090 mL
pH Equivalence Point:	8.131
Results:	0.10 meq/L
Initial & Final pH:	2.857 to 9.884
Titration Duration:	7:09 [mm:ss]
Operator Name:	

Analyst Signature: _____

INSTRUCTION MANUAL

HI901W

AUTOMATIC TITRATOR FOR WINE ANALYSIS

Revision 1.00

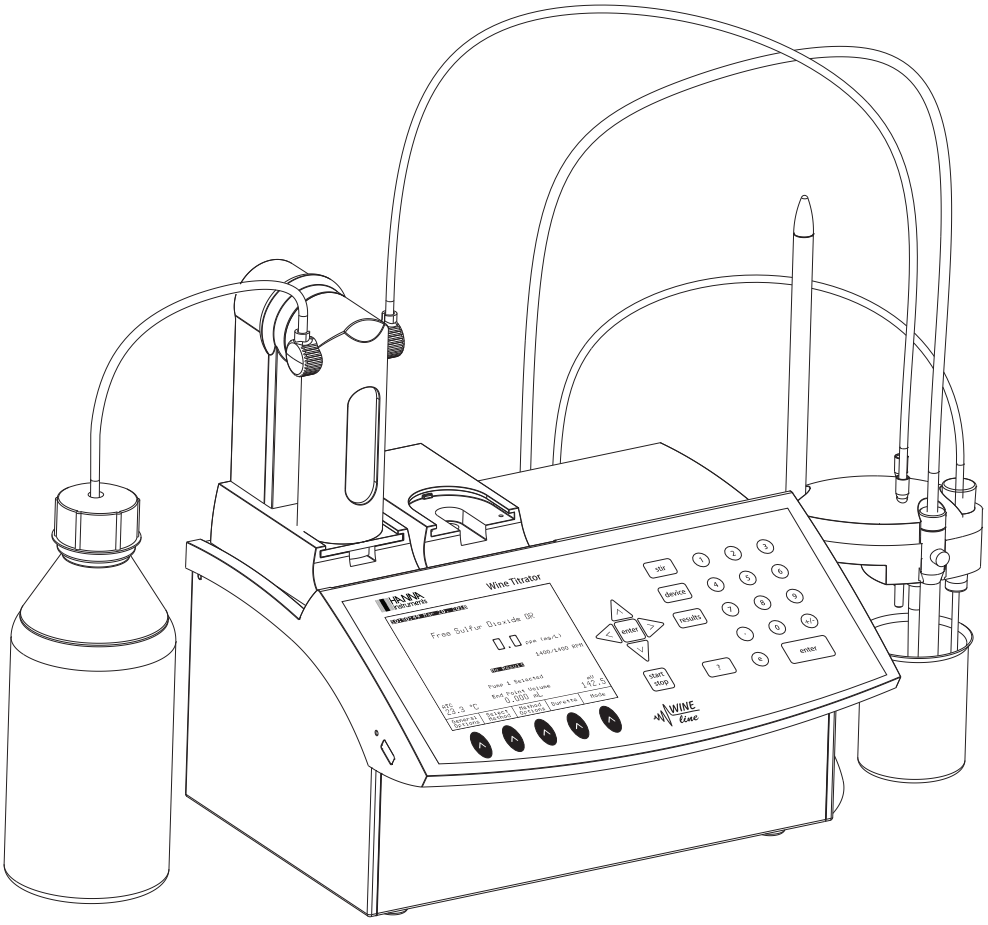


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TABLE OF CONTENTS

Dear customer,

Thank you for choosing a Hanna Instruments Product.
This instruction manual has been written for the **HI901W** Titrator product.
Please read this instruction manual carefully before using the instrument. This manual will provide you with the necessary information for the correct use of the instrument.

1 INTRODUCTION

HI901W is an automatic titrator for wine analysis with high accuracy, great flexibility and repeatability.

The Titrator is designed to perform a variety of wine titrations, allowing the user to obtain both good results and high speed analysis.

The main attributes of this Titrator is:

Flexibility	Support up to 100 titration methods (standard and user defined) Preloaded Wine Analysis Method Pack User-customizable titration / analysis methods (equivalence point, fixed pH/mV end point)
High accuracy	Precise dosing system (under 0.1% accuracy) Precise mV and pH measurements (± 0.1 mV and ± 0.001 pH accuracy) Interpolated end point volume Titrant age and standardization reminders
Repeatability	Powerful built-in algorithms for equivalence point detection (first derivative and second derivative detection algorithms, filtered derivatives option, settable range for equivalence point detection) Fixed end point mV or pH
Quick results	Pre-defined titration methods Pre-titration dosing feature Dynamic / Linear dosing feature
Complete report	The results are displayed directly in the selected units Titration graph can be displayed on the screen and saved as a bitmap User customized reports can be printed, saved or transferred to PC
Direct measurements	The Titrator can also be used for precise mV, pH, ISE and temperature measurements Report of data logging is available for direct measurements
Research grade meter	pH/ mV/ ISE and Temperature meter with Cal Check Up to five calibration points Data logging (log-on-demand or lot logging)
Graphical display	5.7” (320 x 240 pixels) color display with easy-to-view text and graphs Integrated help screens Clearly displayed warning and error messages Self-diagnosis features for peripheral devices including the pump, burette and stirrer

This manual provides information regarding installation and functionality of the Titrator and refined operation suggestions.

Before using the Titrator, it is recommended you become familiar with its various features and functionality.

Chapter 2. Contents

2 SETUP

2.1 Unpacking

The Titrator and the accessories are shipped in a single box containing:

	ITEM	QUANTITY
1	Titrator	1 pc.
2	Pump Assembly	1 pc.
3	Burette Assembly • Burette (with 25-mL syringe) • Aspiration Tube with Fitting and Protection Tube • Dispensing Tube with Normal Dispensing Tip, Fitting, Protection Tube and Tube Guide • Tube Locks • Tool for Burette Cap Removal • Light Spectrum Protection Screen	1 pc.
4	Stirrer Assembly • Overhead Stirrer • Propeller (3 pcs.) • Stirrer Stand • Stirrer support with positioning collar and positioning screw	1 pc.
5	Burette Blank Support	1 pc.
6	Pump and Burette Locking Screws with Plastic Head	2 pcs.
7	Temperature Sensor	1 pc.
8	Shorting Cap	1 pc.
9	Power adapter	1 pc.
10	Instruction Manual Binder	1 pc.
11	USB Memory Stick	1 pc.
12	Quality Certificate	1 pc.

See **Appendix 2**, *Titrator components* section for pictures.

If any of the items are missing or damaged, please contact your sales representative.

Note: *Save all packing materials until you are sure that the instrument functions correctly. Any damaged or defective items must be returned in their original packing materials together with the supplied accessories.*

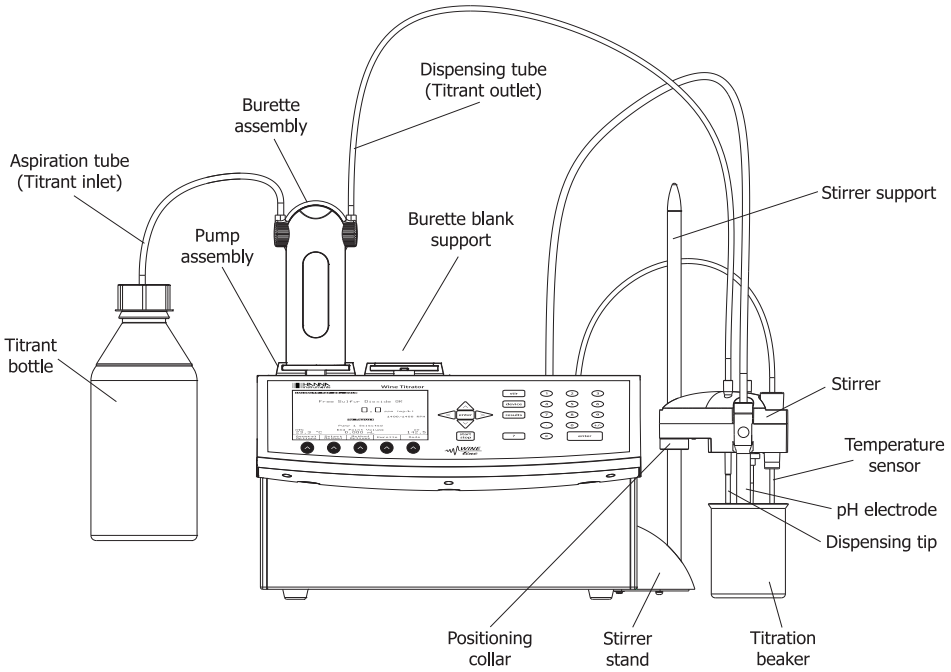
2.2 Safety Measures

The following safety measures must be followed:

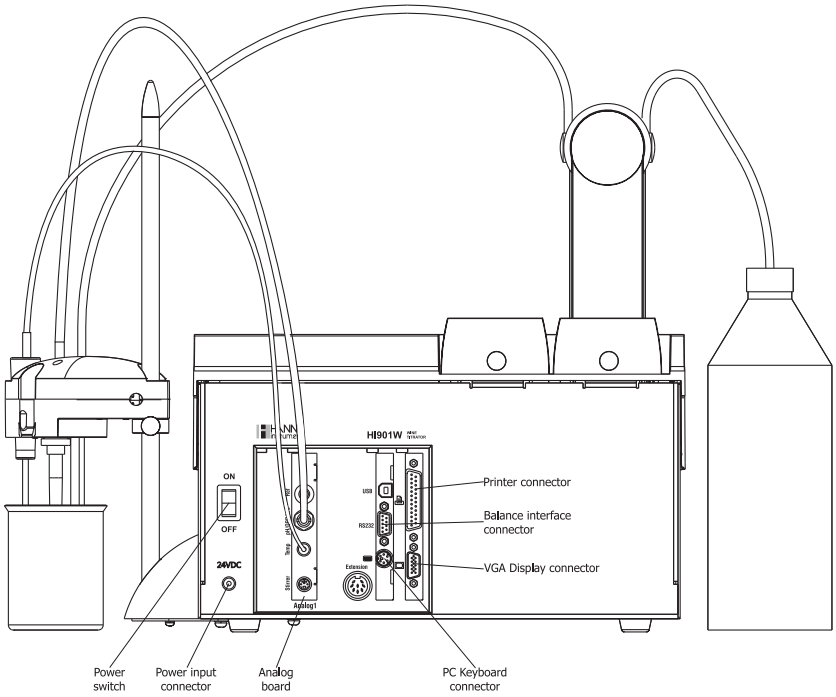
1. Never connect or disconnect the pump assembly with the Titrator turned on.
2. Verify that the burette and the attached tubing are assembled correctly (see **Maintenance, Peripherals**, *Burette Maintenance* section for more details).
3. Always check that the titrant bottle and the titration beaker are on a flat surface.
4. Always wipe up spills and splashes immediately.
5. Avoid the following environmental working conditions:
    - Severe vibrations
    - Direct sunlight
    - Atmospheric relative humidity above 95% non-condensing
    - Environment temperatures below 10°C and above 40°C
    - Explosion hazards
6. Have the Titrator serviced only by qualified service personnel.

2.3 Installation

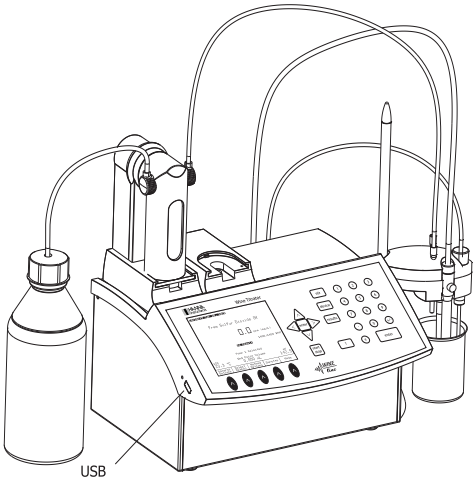
2.3.1 Titrator Front View



2.3.2 Titrator Rear View



2.3.3 Titrator Left-side View



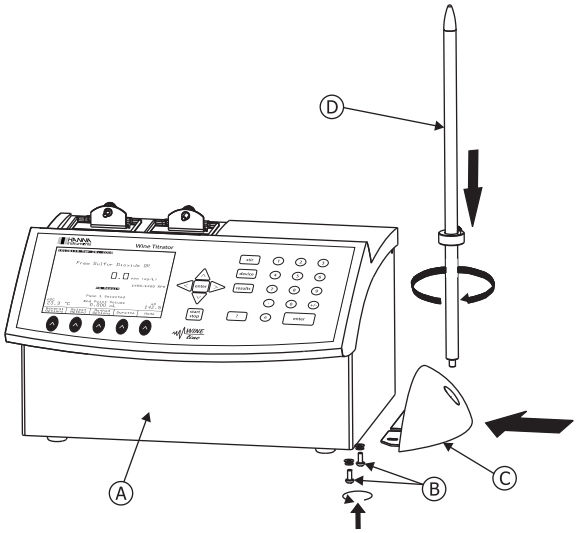
2.3.4 Titrator Assembly

Note: *Assembly operations must be completed before connecting the Titrator to the power supply!*

2.3.4.1 Assembling Stirrer Stand and Support

To assemble the stirrer stand and support:

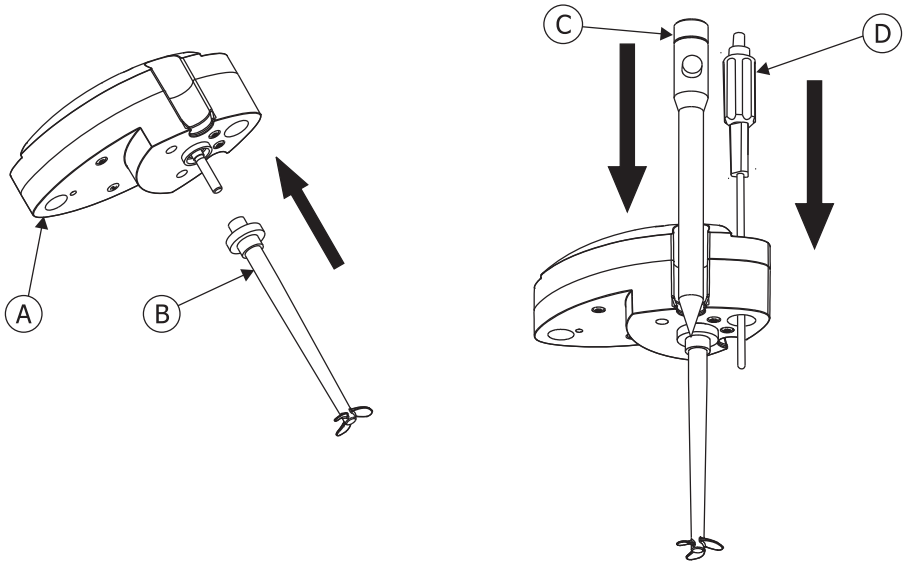
- Remove the screws (B) from the Titrator base (A).
- Attach the stirrer stand (C) to the Titrator.
- Tighten the stirrer stand (C) using the previously removed screws (B).



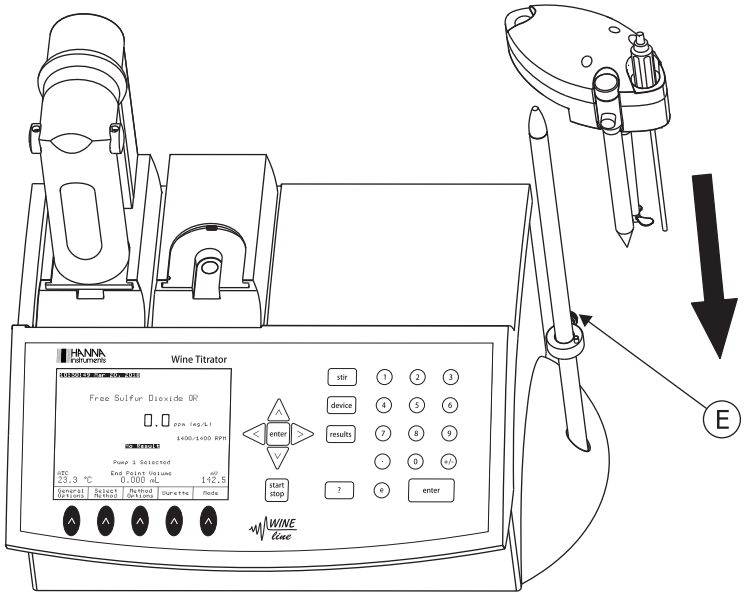
- Screw the stirrer support (D) in the stirrer stand (C).

2.3.4.2 Attaching Stirrer

To attach the stirrer to the Titrator, follow these steps:



- Attach the propeller (B) to the stirrer (A) by pressing it onto the stirrer shaft.
- Insert the electrode (C) and temperature sensor (D) into the dedicated holes on the stirrer. Push them in until they are in a stable position.



- Slide the stirrer on the stirrer support and set the height by tightening the screw located on the positioning collar (E).

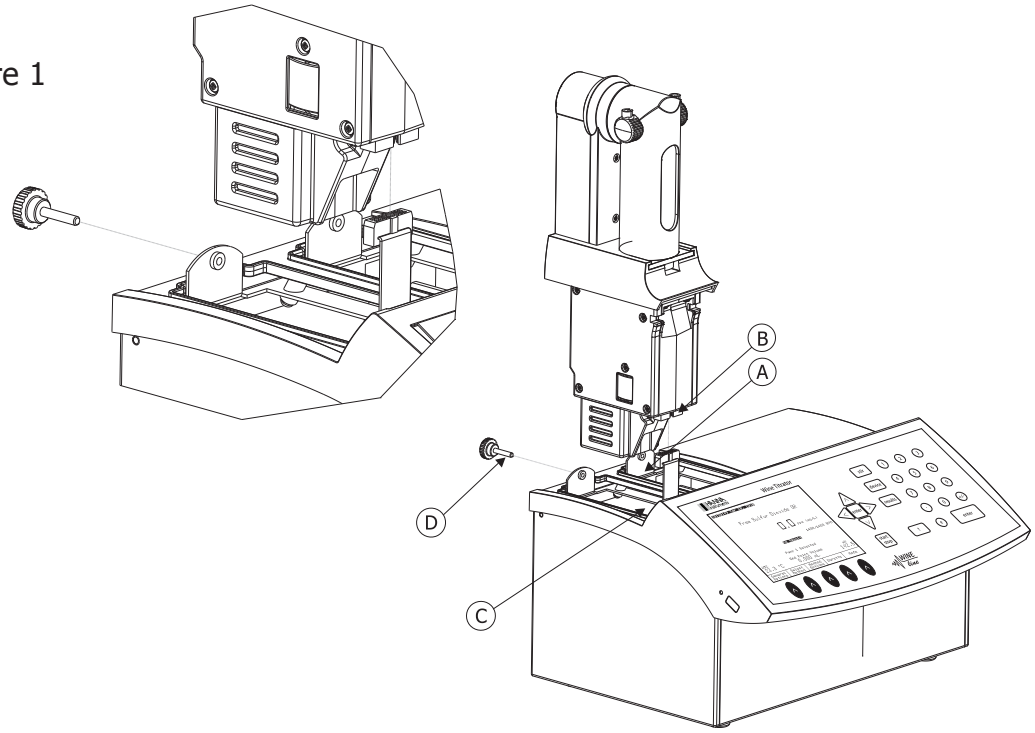
2.3.4.3 Connecting the Pump

To connect the pump, follow these steps:

- Retrieve the pump cable from inside the bay. The pump 1 connector is located in the left bay.
- Connect the cable (A) to the pump as shown below. The pump connector (B) is located in the lower part of the pump, near the motor.
- Lower the pump into the Titrator, then slide it towards the front of the Titrator chassis (C) until it is firmly latched.
- Secure the pump with the locking screw (D).

This procedure can be repeated to connect a second pump.

Figure 1



2.3.4.4 Attaching Burette Blank Support

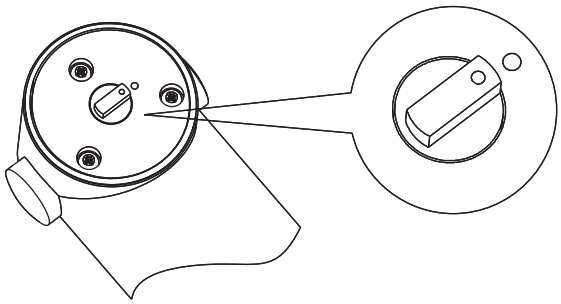
To attach the burette blank support, follow these steps:

- Insert burette blank support into the bay. Lower the burette blank support into the Titrator, then slide it towards the front of the Titrator chassis until it is firmly latched.
- Secure the burette blank support with the locking screw.

2.3.4.5 Attaching Burette

Make sure that the mark from the valve actuating cap and from the burette body are aligned as shown in Figure 2.

Figure 2



While ensuring the correct coupling between the syringe plunger (1) and the pump piston (2) (Figure 3), slide the burette into the support on the burette pump (Figure 4).

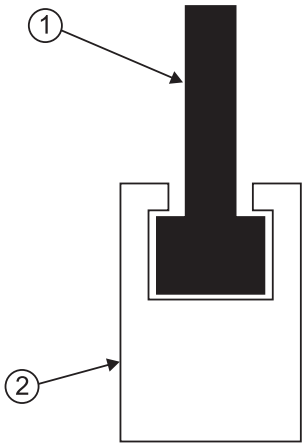


Figure 3

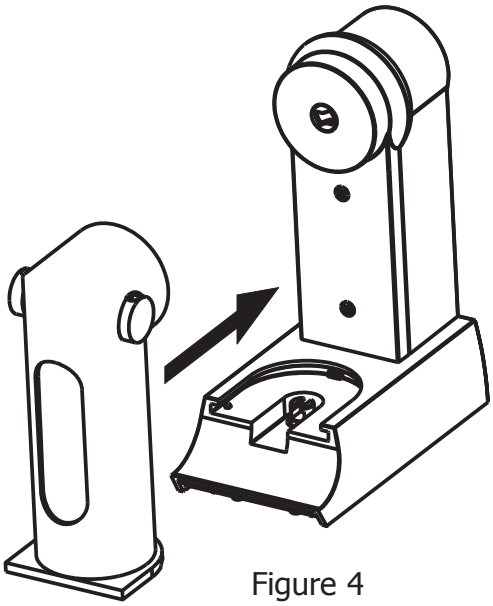
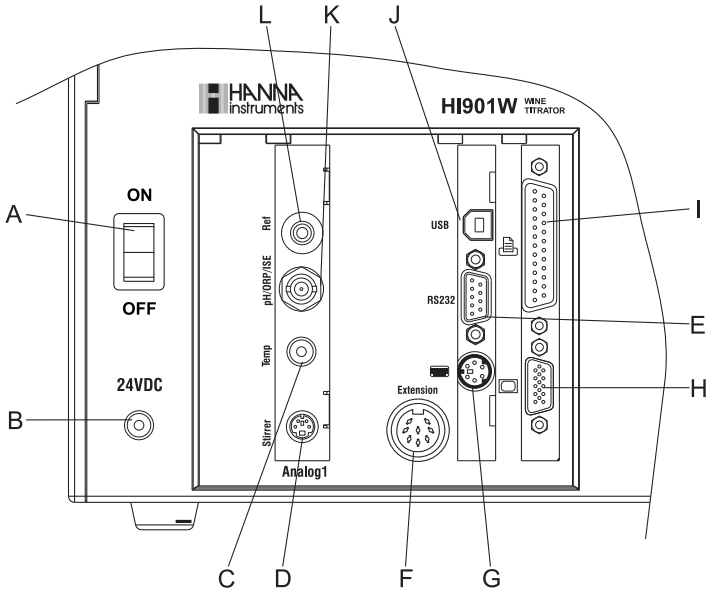


Figure 4

2.3.4.6 Electrical Connections

- Connect the electrode to the BNC connector (K).
- Connect the temperature sensor to the RCA connector (C).
- Connect the stirrer to the MINI-DIN connector (D).
- Connect the power adapter cable to the power input connector (B).



Nr	Function	Type of Connector
A	Power switch	
B	Power input connector (24VDC)	DC Power jack connector
C	Temperature sensor	RCA Socket
D	Stirrer	4-pin Mini DIN
E	RS232 interface (Balance Interface)	Standard DB 9 Pin Socket
F	Connector for expansion device (Reserved)	8-pin DIN
G	External PC keyboard	6-pin Mini DIN (Standard PS2)
H	External display	Standard VGA Display 15-pin Socket
I	Printer	DB 25–pin Socket
J	USB interface (Reserved)	USB Standard B
K	Connection for pH, ORP and ISE half-cell or combination electrodes	BNC Socket
L	Reference electrode	Ø 4 mm Banana Socket

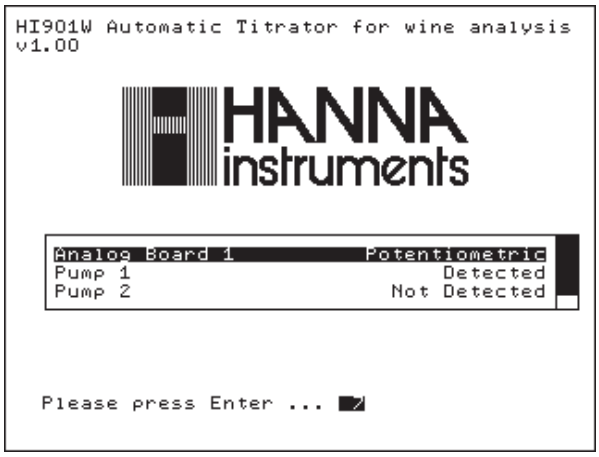
Chapter 3. Contents

3 USER INTERFACE

3.1 Start Up

Once the instrument is assembled and installed, follow the steps below to start the Titrator:

- Connect the instrument to a power outlet with the supplied power adapter.
- Turn on the Titrator from the power switch located on the back of the instrument.
- Wait until the Titrator finishes the initialization process.
- Press [enter] when prompted or wait a few seconds for Titrator to start.



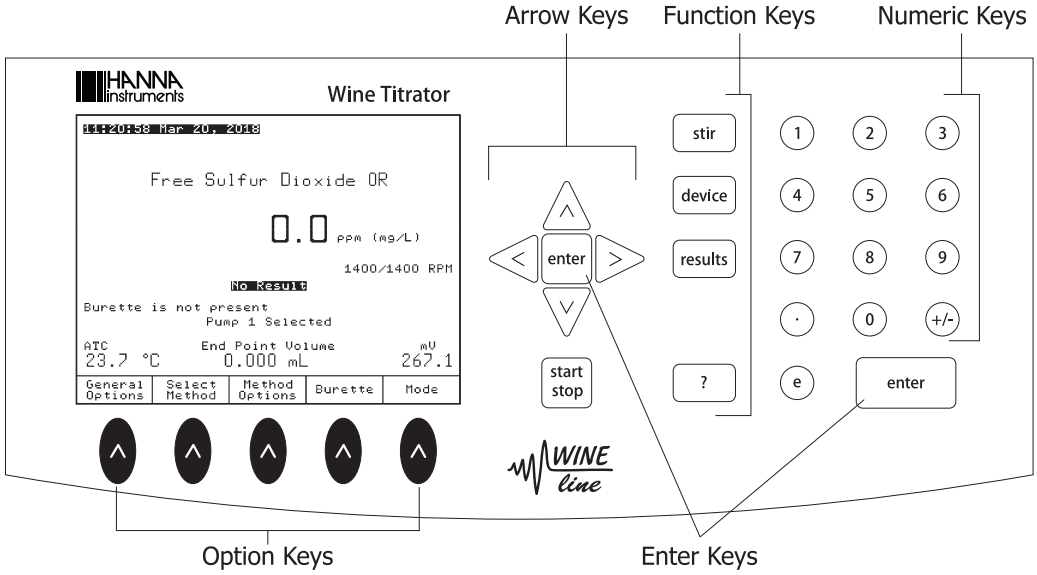
Note: *All the performed initialization processes must be successfully completed. If one of them is terminated by a* "Failed" *message, restart the Titrator from the power switch. If the problem persists, contact your dealer.*

3.2 Description

This chapter describes the basic principles of navigating through the user interface, selecting fields and entering values from the keypad.

3.2.1 Keypad

The titrator's keypad is grouped into five categories, as follows:



3.2.1.1 Function Keys

If one of these keys is pressed, the associated function is immediately performed. Some of the keys are active only in specific screens:

start/stop	Starts or stops a titration
stir	Turns the selected stirrer ON and OFF
device	Reserved
results	Access the results menu
?	Displays contextual Help

3.2.1.2 Option Keys

These keys are assigned to the virtual keys on the display. Their functions are listed in the boxes above the buttons and vary depending on the displayed screen.

An underlined virtual key can also be activated by pressing enter.

3.2.1.3 Arrow Keys

These keys have the following functions:

- Move the on-screen cursor.
- Increase and decrease the stirrer speed and other settings.
- In the alphanumeric screen, to select a character.
- Navigate through menu options.

3.2.1.4 Numeric Keys

Keys	(0) to (9)	Used for numeric entries.
	(+/-)	Toggles between positive and negative values.
	(·)	Decimal point.
	(e)	Initiates entry of exponent for scientific notation.

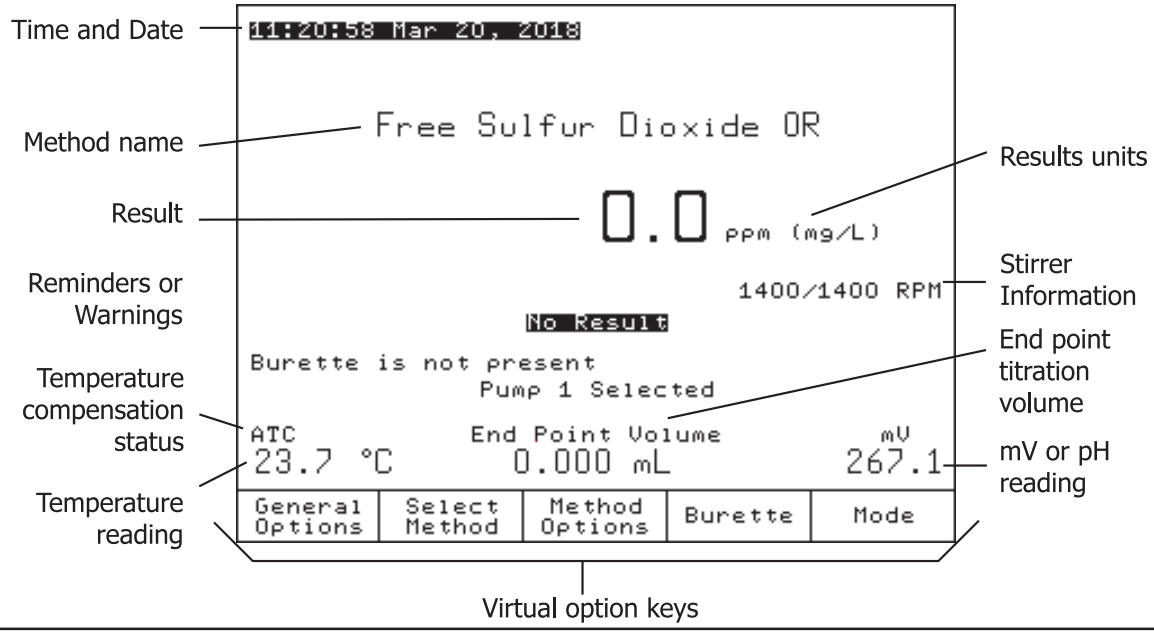
3.2.1.5 Enter Key

Both [enter], [enter] keys perform the same functions:

- Accept alphanumeric data entry.
- Executes the default (underlined) virtual option key.

3.2.2 Display

The Titrator has a large color graphical display. The main screen is shown below with short explanations of the screen segments.



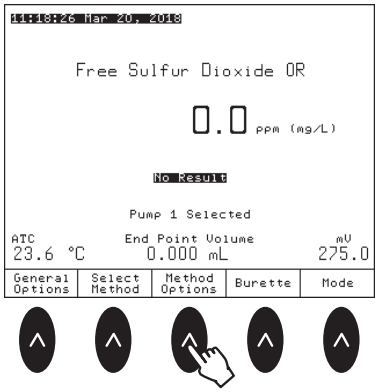
The user interface contains several screens. For each titrator function, one or more screens are used.

3.2.3 The Main Screen

After start up and initialization, the first screen displayed is the main screen.
Main screen fields:

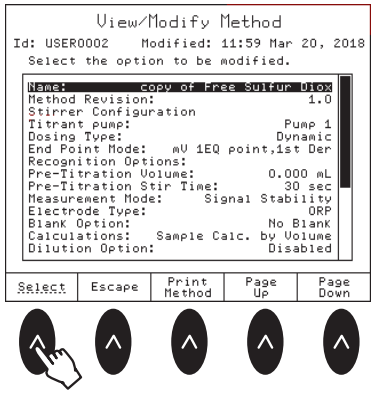
Method name:	Displays the name of the selected method.
Time and date:	Displays the current date and time.
Temperature reading:	Displays the measured temperature.
ATC:	Automatic temperature compensation
Manual:	Manual temperature compensation
Manual:	Temperature probe is not connected, manual temperature compensation
Stirrer information:	Actual / Set stirrer speed is displayed in RPM. When stirrer is off, the stirrer information is not displayed.
End point volume:	Displays the volume delivered to reach the titration end point. When no titration has been performed, the displayed volume is "0.000 mL".
Result:	Displays the titration result or the direct reading measurement.
mV or pH reading:	Displays the current readings. The reading will be in mV or pH.
mV:	Indicates actual potential reading.
rel mV:	Indicates relative potential reading.
pH:	Indicates actual pH value.
Titration status:	Displays the status of the selected titration. **No results** is displayed when a titration has not been performed.
Reminders:	Indicates when a task needs to be performed and displays error or warning messages.
Pump 1 Selected:	Displays the active pump.

3.3 Menu navigation



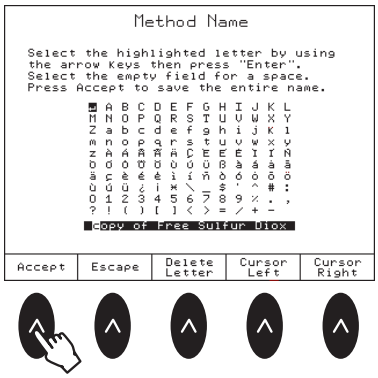
3.3.1 Selecting an Option

To select an option, simply press the option key below the virtual key. For example, to access the ***Method Options*** screen press the option key below it.



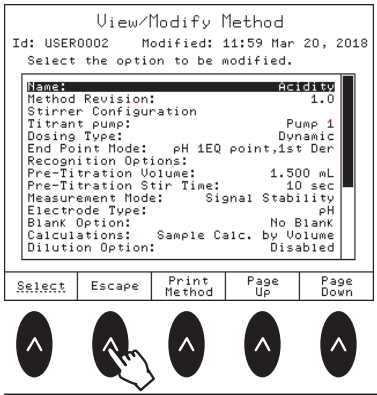
3.3.2 Selecting a Menu Item

To select an item from the menu screen, use the arrow keys △ and ▽ to move the cursor.
When the menu is larger than the display, a scroll bar is active on the right side. The Page Up and Page Down keys can be used to scroll through the pages.
To activate the selected menu item, press enter or Select.

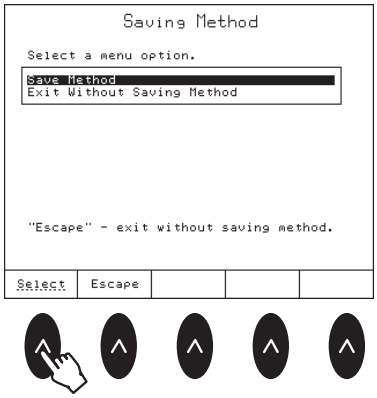


3.3.3 Entering Text

To enter text in an alphanumeric input box, first erase the previous text by using Delete Letter.
To enter a letter, highlight it using the arrow keys then press enter. Use the same procedure to enter the whole name.
For editing, use the Cursor Left and Cursor Right keys.
When editing is complete, press Accept.

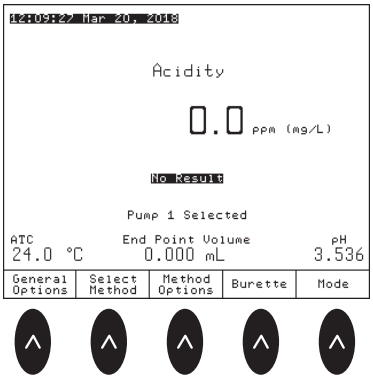


The method name will be updated and displayed in the name field of the ***View/Modify Method*** screen.
When all the desired parameters have been set, press Escape.



3.3.4 Saving Modifications

The ***Saving Method*** screen allows the user to save the modifications. To exit from ***Saving Method*** screen without saving, press Escape or highlight the *Exit Without Saving Method* option and then press Select. To save the modifications highlight the *Save Method* option and then press Select.



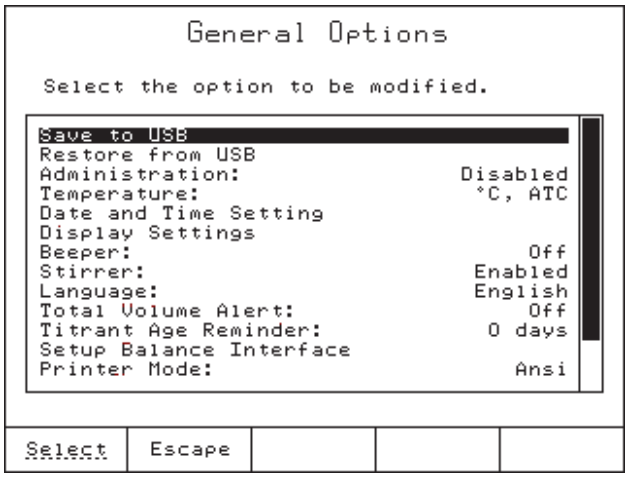
After the method name is changed, it appears in the method name field.

Note: *To access the contextual help menu, press ? at any time. Help is related to the displayed screen. Press Escape or press ? again to return to the previous screen.*

Chapter 4. Contents

4 GENERAL OPTIONS

The ***General Options*** screen gives access to options that are not directly related to the titration process or pH / mV / ISE measurement. To access this screen, press General Options from the main screen.
The available menus are described below:

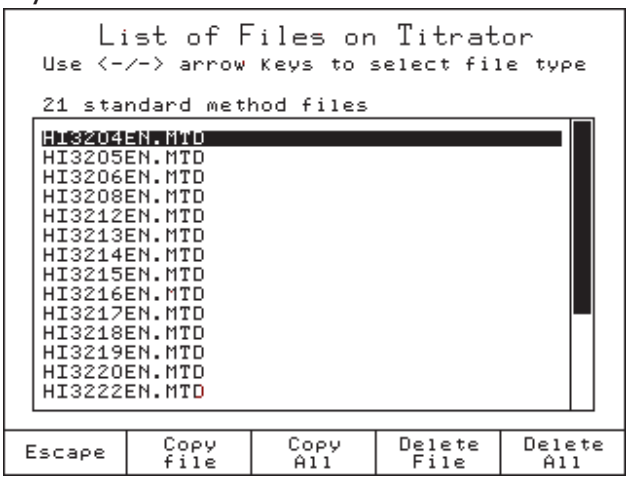


4.1 Save Files to USB Storage Device

This option allows the user to save files from the Titrator to a USB storage device.
On the Titrator, the available file types are:

Standard Method Files	**- HI3204EN.MTD** or **HI3205EN.MTD**
User Method Files	**- USERXXXX.MTD** (e.g.: USER0001.MTD)
Report Files	**- Ti_XXXXX.RPT, mV_XXXXX.RPT, pH_XXXXX.RPT, ISEXXXXX.RPT, mVrXXXXX.RPT** (e.g.: Ti_00001.RPT, mV_00001.RPT, pH_00001.RPT, ISE00001.RPT, mVr00001.RPT)

Use the ◁ and ▷ keys to select the file type. The number of files and each file name on the Titrator will be displayed.



The option keys allow the following operations:

Delete File	Deletes the highlighted file.
Delete All	Deletes all currently displayed files.
Copy File	Copies the highlighted file from Titrator to a USB storage device.
Copy All	Copies all currently displayed files from Titrator to a USB storage device.
Escape	Returns to the ***General Options*** screen.

The status of the transfer ("Successful" / "Unsuccessful") and the file name of the currently processed file are displayed during copying or deleting.

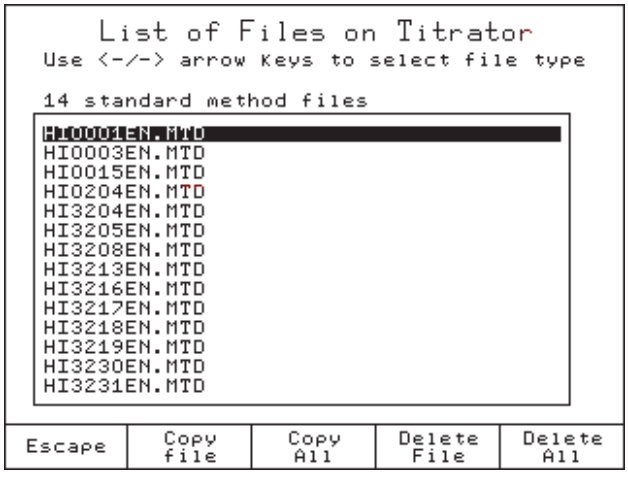
Note: The saved files will be stored on the USB key in the **HI901W** folder, as follows:
- Methods: **USB Drive: *HI901W\Methods*.mtd***
- Reports: **USB Drive: *HI901W\Reports*.rpt***

4.2 Restore Files from USB Storage Device

This screen allows the user to transfer files from the USB storage device to the Titrator.
The file types that can be transferred are:

Standard Method Files	**- HI3204EN.MTD** or **HI3205EN.MTD**
User Method Files	**- USERXXXX.MTD** (e.g.: USER0001.MTD)
Report Files	**- Ti_XXXXX.RPT, mV_XXXXX.RPT, pH_XXXXX.RPT, ISEXXXXX.RPT, mVrXXXXX.RPT** (e.g.: Ti_00001.RPT, mV_00001.RPT, pH_00001.RPT, ISE00001.RPT, mVr00001.RPT)

Use the ◁ and ▷ keys to select the file type.
The number of files and the name of each file found on the USB storage device is displayed on the screen.



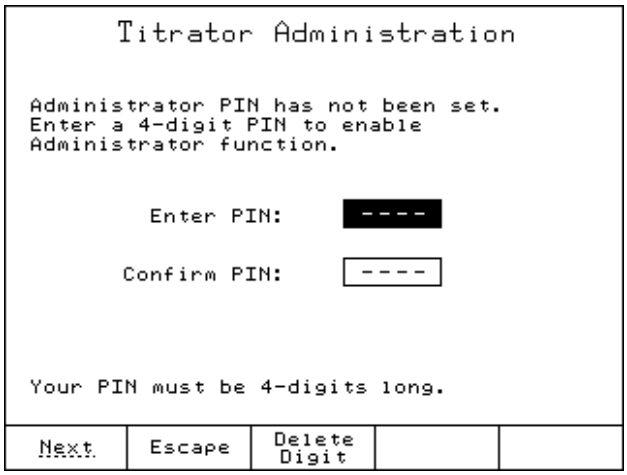
The option keys allow the following operations:

Delete File	Deletes the highlighted file from the USB storage device.
Delete All	Deletes all currently displayed files from the USB storage device.
Copy File	Copies the highlighted file from the USB storage device to the Titrator.
Copy All	Copies all currently displayed files from the USB storage device to the Titrator.
Escape	Returns to the ***General Options*** screen.

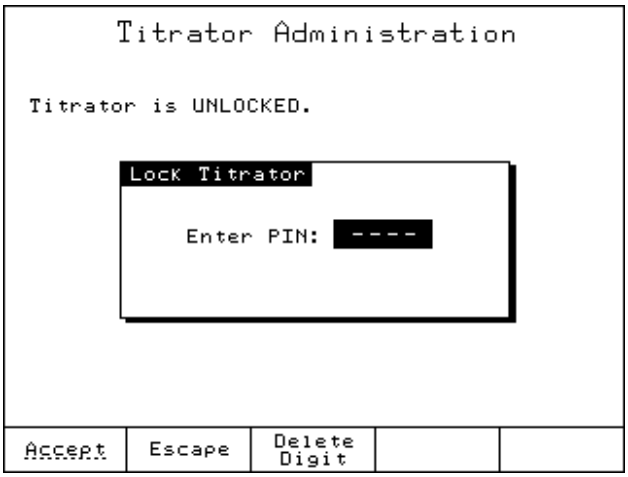
Note: In order to restore files from a USB key, please ensure that the methods and / or reports you wish to transfer to the Titrator are in the correct folder:
- Methods: **USB Drive: *HI901W\Methods*.mtd***
- Reports: **USB Drive: *HI901W\Reports*.rpt***

4.3 Administration

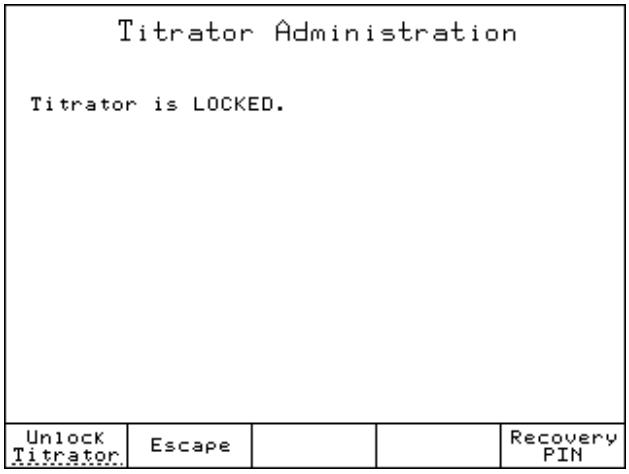
A 4-digit numeric PIN can be set to prevent unauthorized changes from being made.
When the user enters administration and a pin has not been set, the user will be prompted to enter a new PIN.



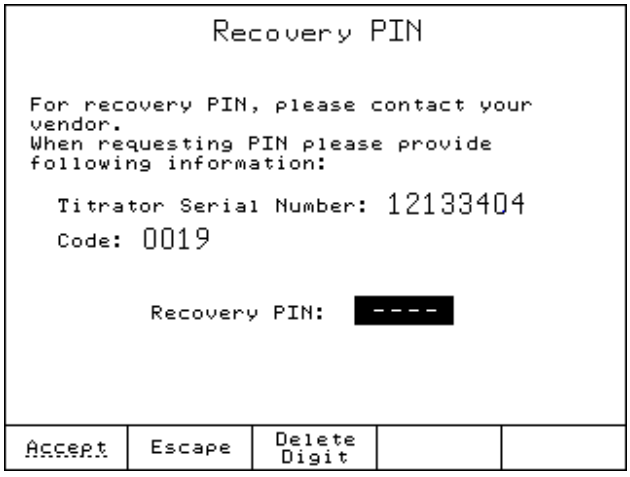
Once a PIN has been set, the Titrator can be locked. When a Titrator is locked, the users cannot modify methods or delete reports. Basic functions are still available (review reports, save to USB, etc.).



To return to administrator mode, the Titrator can be unlocked by entering the PIN.

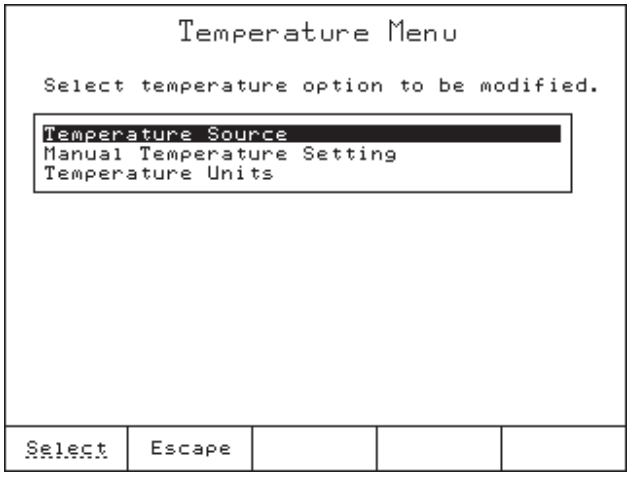


If the PIN is lost or forgotten, press recovery pin and contact technical support to supply the required information.



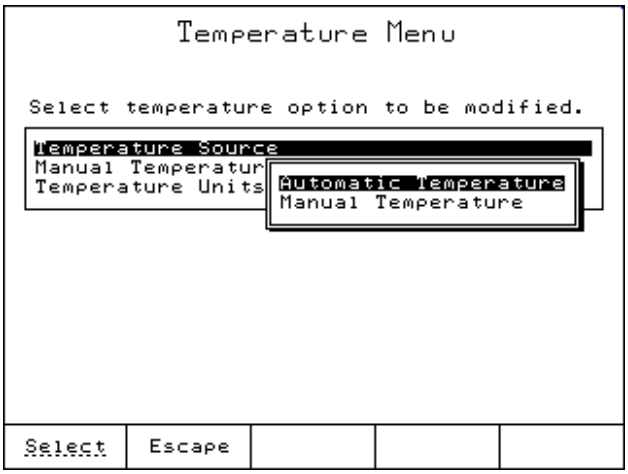
4.4 Temperature

The ***Temperature Menu*** allows access to all of the settings related to temperature.



4.4.1 Temperature Source

Select the temperature source used for temperature compensation.



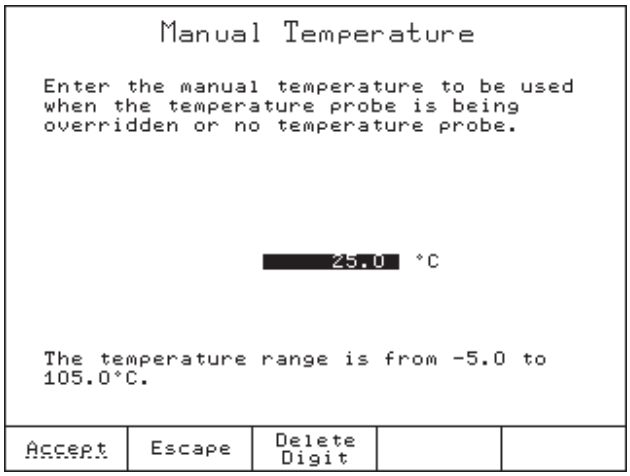
When *Automatic Temperature Compensation* is selected, "ATC" is displayed on the main screen and the temperature is read by the temperature probe.

When *Manual Temperature* is selected, "Manual" is displayed on the main screen and a preset temperature value is used for temperature compensation.

Note: *The selected temperature source will be indicated in the report files: A for Automatic and M for Manual.*

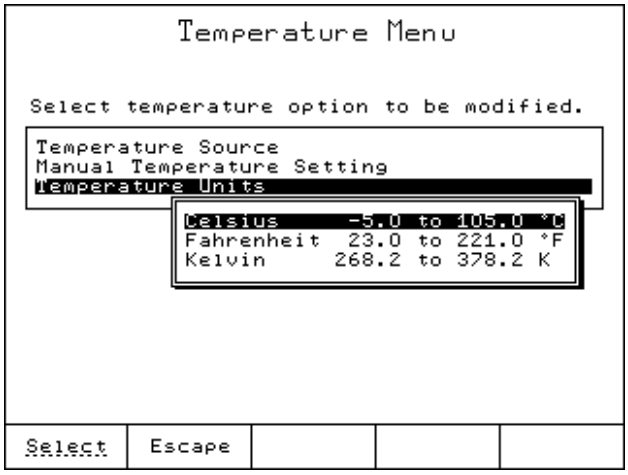
4.4.2 Manual Temperature Setting

If the temperature probe is not connected, the user can manually set the temperature used by the Titrator for compensation. This can be done when the *Manual Temperature* option is selected.
The temperature value can be set between –5 and 105 °C.



4.4.3 Temperature Units

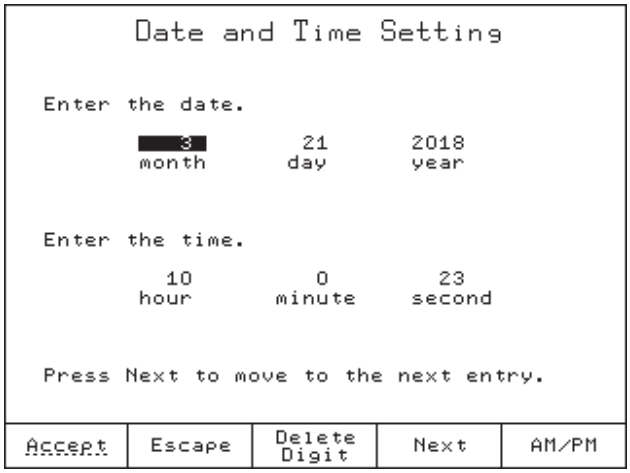
The following temperature units can be selected.



The temperature ranges are as displayed in the ***Temperature Units*** screen.

4.5 Date and Time Setting

This screen allows the user to set the date and time.



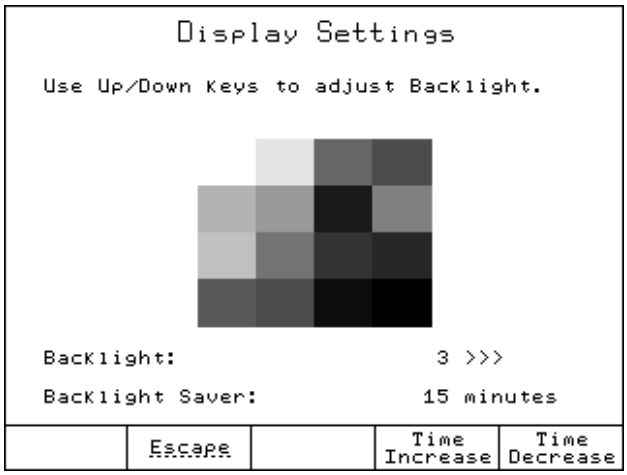
Use the △ and ▽ keys or the numeric keys to modify the date and time.

Press [Next] to move the cursor to the next field.

Press [AM / PM] or [24-hour] to change the time format.

4.6 Display Settings

This screen allows the user to customize the display settings.



Option Keys:

Time Increase	Increases the backlight saver time interval
Time Decrease	Decreases the backlight saver time interval

The backlight intensity can be adjusted using △ and ▽ keys.

There are 8 levels of backlight intensity, ranging from 0 to 7.

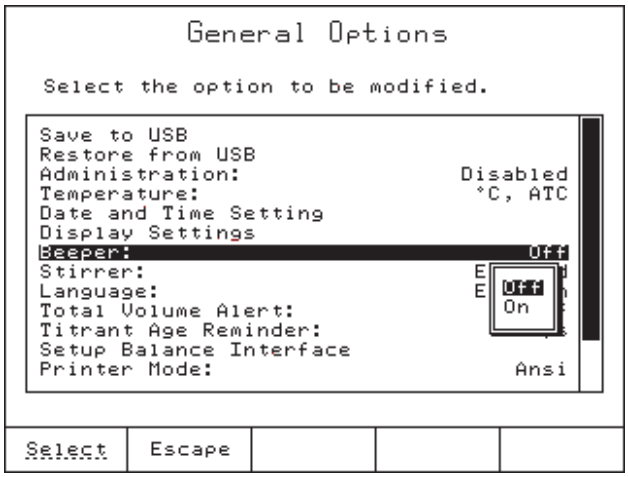
A color palette is displayed in the center of the screen allowing an easy selection of the appropriate backlight intensity.

The backlight saver option protects the display during standby periods when no keys have been pressed for a set amount of time.
If the display backlight is off, any keystroke will activate the backlight without performing any action.

The range for the backlight saver timer is 1 to 60 minutes. To disable the backlight saver, increase the time to the maximum allowed. The “Off” indication will appear.

4.7 Beeper

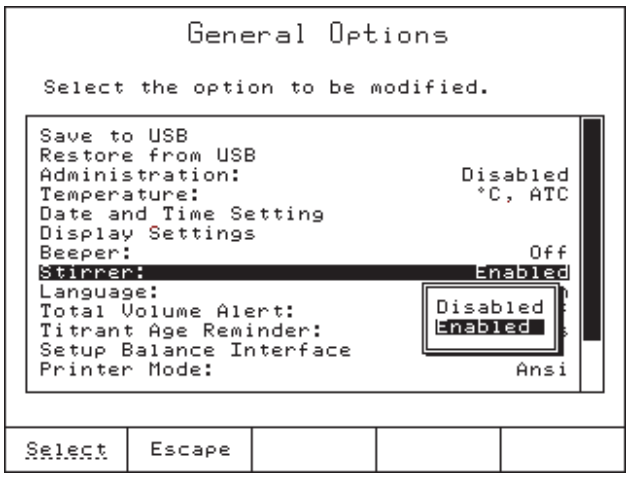
This screen allows the user to be turn the Beeper On (enabled) or Off (disabled).



The beeper will sound after a titration is completed, when an invalid key is pressed or when a critical error occurs during titration.

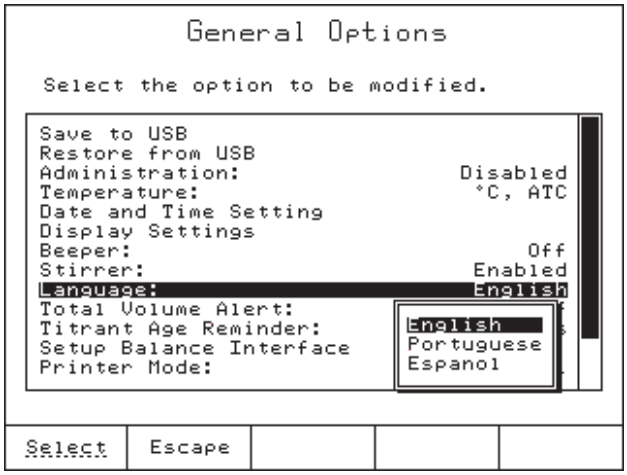
4.8 Stirrer

This screen allows the stirrer to be enabled or disabled.



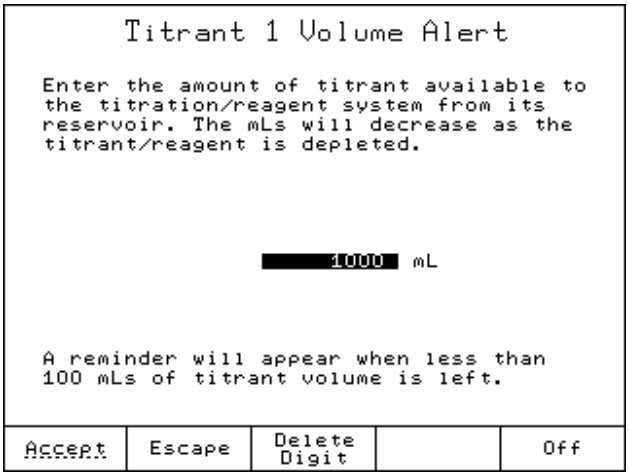
4.9 Language

Select an available language.



4.10 Total Volume Alert

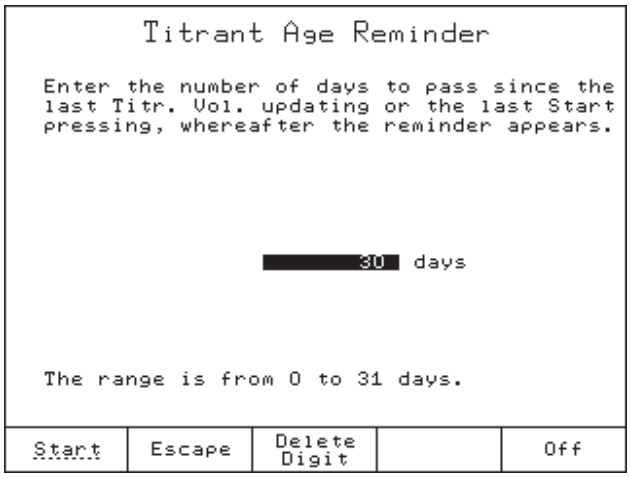
This screen allows a programmable reminder to appear when the titrant reservoir is below 100 mL. The titrant volume will decrease as the titrant is used.



The "Low Titrant Volume" reminder message will appear when the available titrant volume is under 100 mL.
After the new titrant volume has been set on the Titrator (in the ***Total Volume Alert*** screen), a warning message appears reminding the user to perform titrant re-standardization. The volume of titrant can be set from 0 to 10,000 mL.

4.11 Titrant Age Reminder

A programmable reminder will appear when it is time to verify the titrant concentration or to change the titrant.

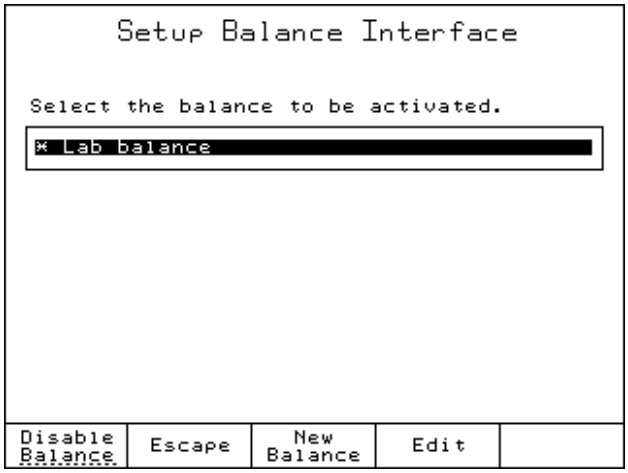


The "Check Titrant Concentration" reminder will appear when the set number of days has passed since the total volume alert was set or since the timer was started. The reminder can be disabled by pressing [Off].

The range is from 0 to 31 days.

4.12 Setup Balance Interface

This screen allows the users to connect an analytical balance for automatic acquisition of sample mass prior to titration or standardization.



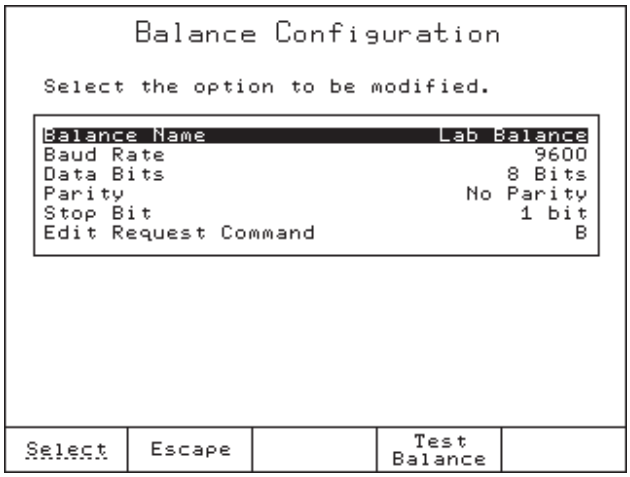
The balance is connected to the Titrator via RS 232 interface.

Press [New Balance] to add a new balance to the list.

Press [Enable Balance] to enable the balance interface feature.

Press [Disable Balance] to disable the balance feature (automatic weight acquisition will be not available).

Press [Edit] to customize the serial communication parameters by accessing the ***Balance Configuration*** screen.

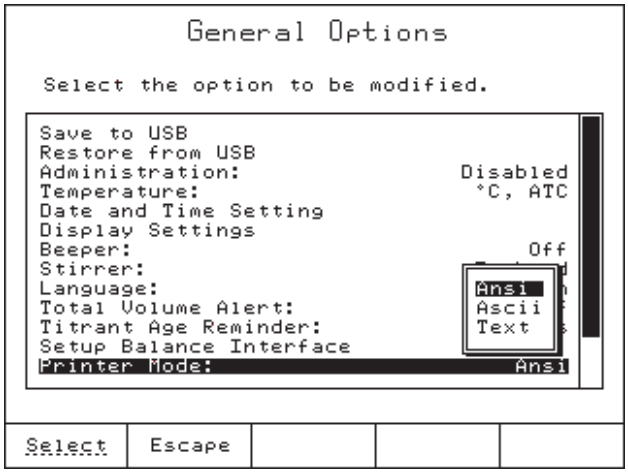


Be sure that the settings on the Titrator *Balance Configuration* menu match the settings for your particular balance (baud rate, data bits, parity, stop bits number, request command syntax). It may be necessary to change settings on your balance. Users should consult their balance instruction manual.

Before leaving this screen, be sure the connection with the balance is working properly by pressing the [Test Balance] key.

4.13 Printer Mode

This screen allows the users to select the printing mode: Ansi (default), Ascii and Text mode.



Ansi mode:

Use this mode when your printer is set as Ansi. In this case all the accented characters / symbols available in Titrator will be printed on your printer.

Ascii mode:

Use this mode when your printer is set as Ascii. In this case only some of the accented characters / symbols available in Titrator will be printed on your printer.

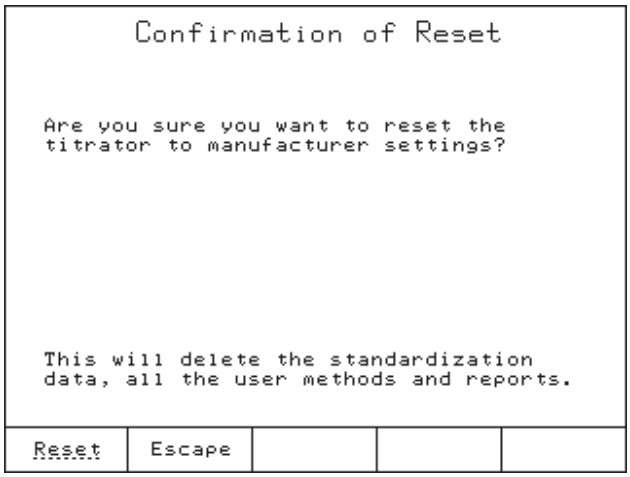
Text mode:

Use this mode when you don't need to print the accented characters.

4.14 Reset to Default Settings

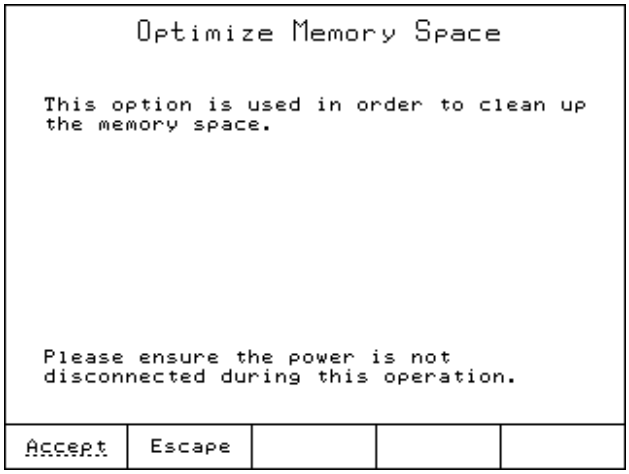
This option restores the manufacturer settings.

Note: *This will also delete all the user - created methods and restore all manufacturer settings such as titrator configuration, standard method parameters, etc.*



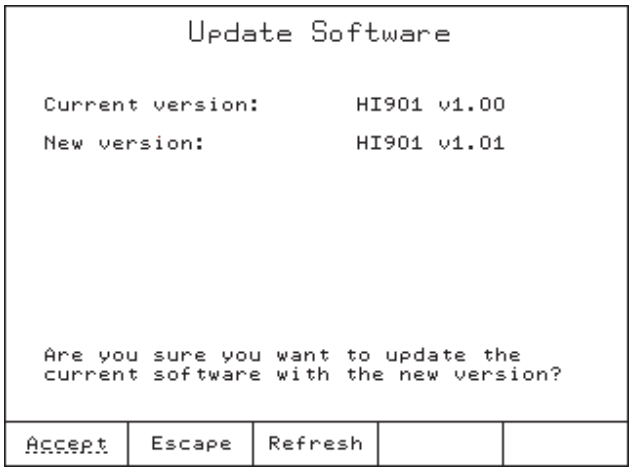
4.15 Optimize Memory Space

This screen allows the user to optimize the memory.



4.16 Update Software

This screen allows the user to update the titrator software from a USB storage device containing a software setup kit.



To update the software:

- Copy the "SET901W" folder to a USB storage device.
- Insert the USB storage device into the Titrator.
- Go to "General Options", then "Update Software". The Titrator should display the current and new software versions.
- Press Accept. When prompted, remove the USB storage and restart the Titrator.

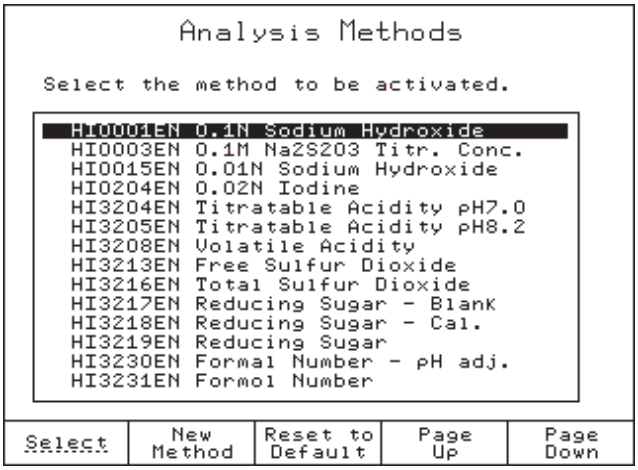
Chapter 5. Contents

5 TITRATION METHODS

All of the parameters required to complete an analysis are grouped into a method.
The Titrator is supplied with a pack of standard methods.
Standard and user methods can be upgraded, saved or deleted using a USB storage device.

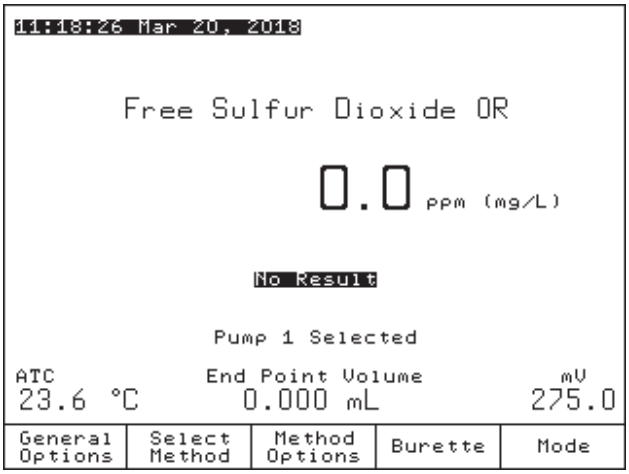
5.1 Selecting Methods

To select a method, press Select Method from the main screen. A list of available methods will be displayed.



In the ***Analysis Methods*** screen, you can view the list of all available methods (standard and user methods).

To select a method, highlight the method then press Select. The name of the selected method will be displayed on the main screen.



5.2 Standard Methods

The standard methods are developed for the most common types of analysis.
Only specific method parameters can be modified by the user (see *Method Options* section).
Also, standard methods can be used as models to create new user methods.

5.2.1 Upgrading Standard Methods

To upgrade the Titrator with new standard methods, follow the steps below:

From USB Storage Device:

- Insert the USB storage device into the USB port, located on the left side of the Titrator.
- Press General Options from the main screen.
- Using △ and ▽ keys, highlight the *Restore Files from USB Storage Device* option and choose Select.
- Using ◁ and ▷ keys, navigate through file types to find "standard method files". The list with available standard methods will be displayed.
- Press the Copy File or Copy All key to upgrade the Titrator with the standard methods.
- Press Escape to return to ***General Options*** screen.

5.2.2 Deleting Standard Methods

Unnecessary standard methods can be removed from the Titrator by following the procedure below:

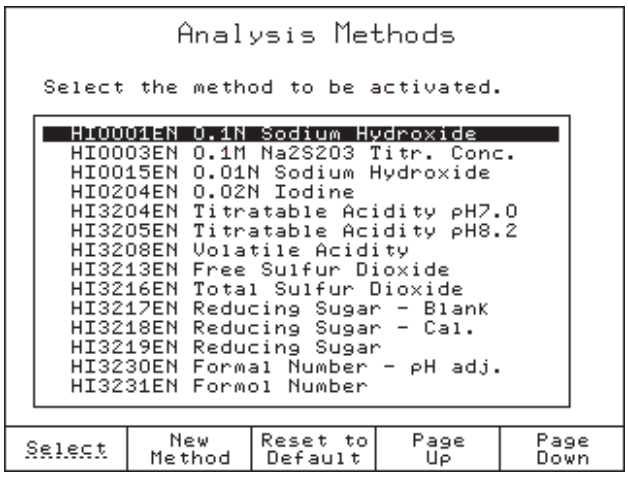
From General Options Screen:

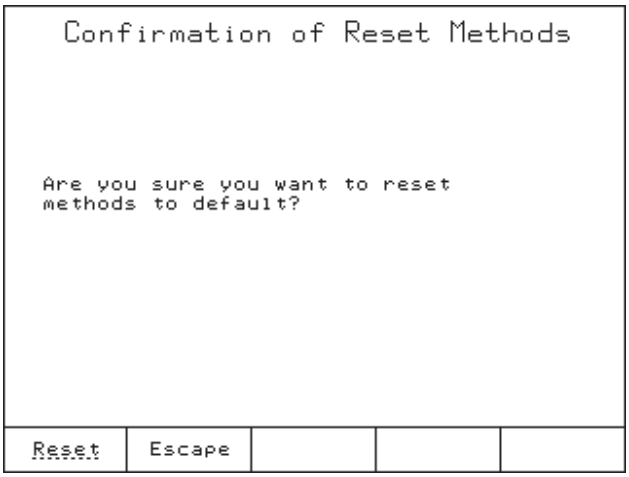
- Using the △ and ▽ keys, highlight the *Save Files to USB Storage Device* option and press Select.
- Using the ◁ and ▷ keys, navigate through the file types menu to find "standard method files". The available standard methods will be displayed.
- Press the Delete or Delete All keys to remove unnecessary standard methods.
- Press Escape to return to the ***General Options*** screen.

Note: *Only a limited number of user methods can be generated. The Titrator can hold 100 methods (standard and user). When it is reached, a warning message will be displayed.*

5.2.3 Restore the Standard Methods to the Manufacturer Settings

You can restore the standard methods to the manufacturer setting by highlighting a standard method and pressing [Reset to Default].





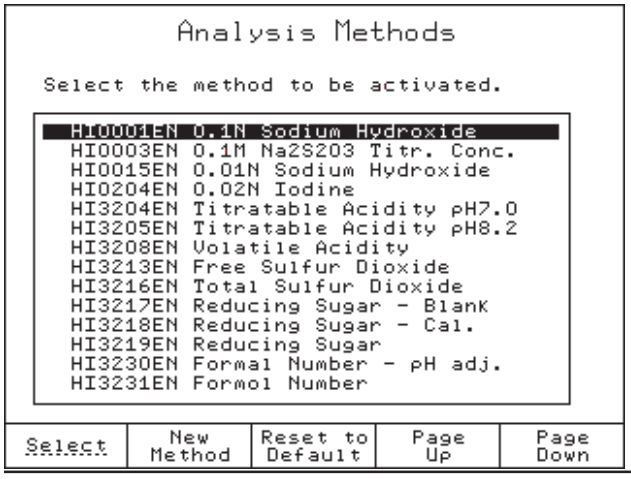
5.3 User Methods

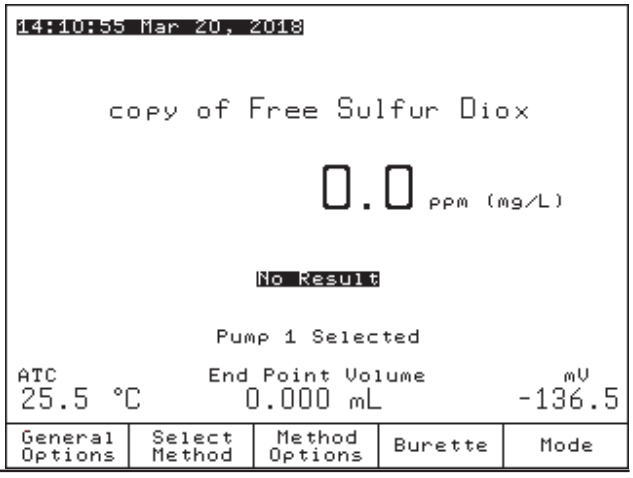
These methods are defined by the user (usually by modifying a standard method).
The user methods can be developed in accordance with the requirements of the user. All method parameters can be modified by the user.

5.3.1 Creating User Methods

To create a new user method, start from a standard or user method and follow these steps:

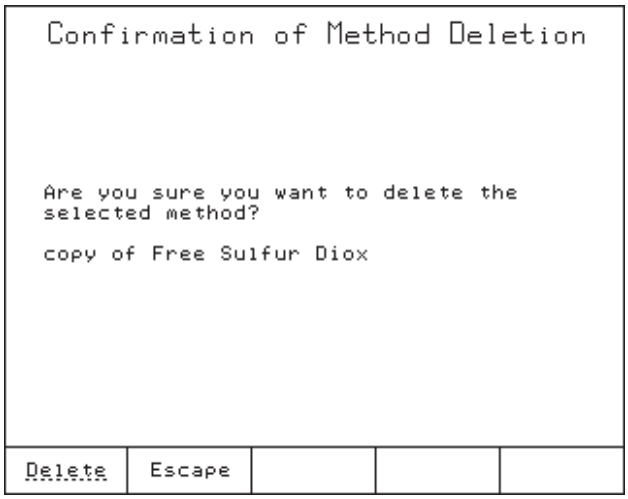
- Press [Select Method] from the main screen.
- Using the △ and ▽ keys, highlight an existing method from the method list.
- Press [New Method]. A new user method will be generated.
- Press [Select] to activate the new user method.





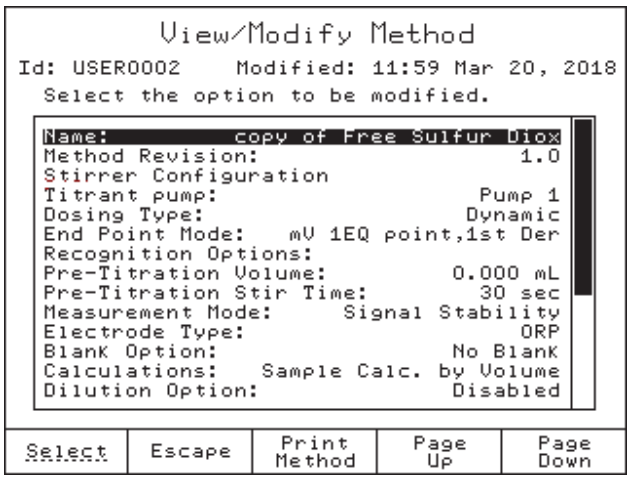
5.3.2 Deleting User Methods

To remove a user method, press [Select Method] from the main screen. Highlight the user method that you want to delete and press [Delete]. A screen will appear in order to confirm the deletion. Press [Delete] again to confirm, or press [Escape] to cancel the operation.

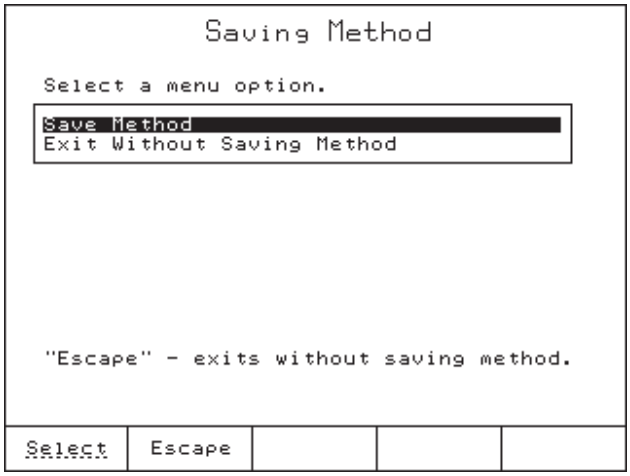


5.4 View / Modify Method

To modify the method parameters, press [Method Options] from the main screen. A list of all the parameters for the selected method will be displayed. Using the △ and ▽ keys, highlight the option that you want to modify and choose [Select].



To exit the ***View / Modify Method*** screen, press Escape.
You can choose to save the modifications or to discard them.

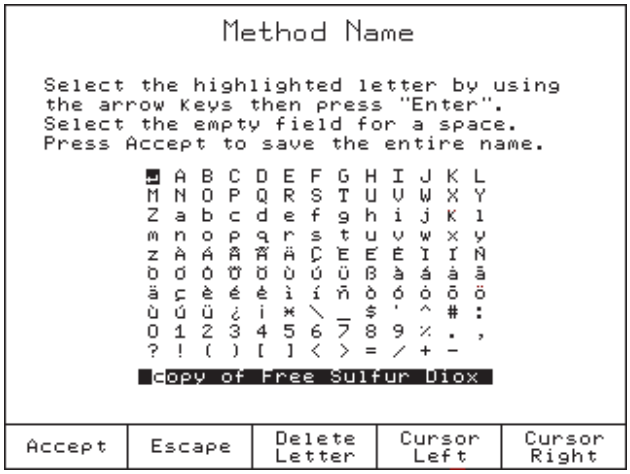


5.5 Method Options

***Note**: Only certain method options can be changed for Standard methods.*

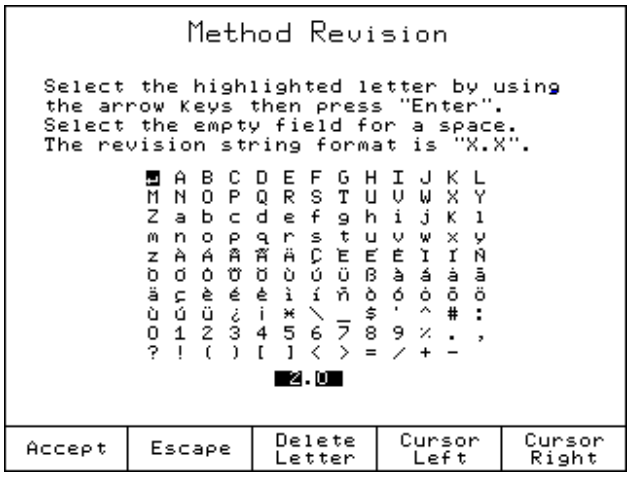
5.5.1 Name

Enter a name for the new method (up to 24 characters). Use the arrow keys to navigate through the character table. Press enter to add the highlighted character to the name.



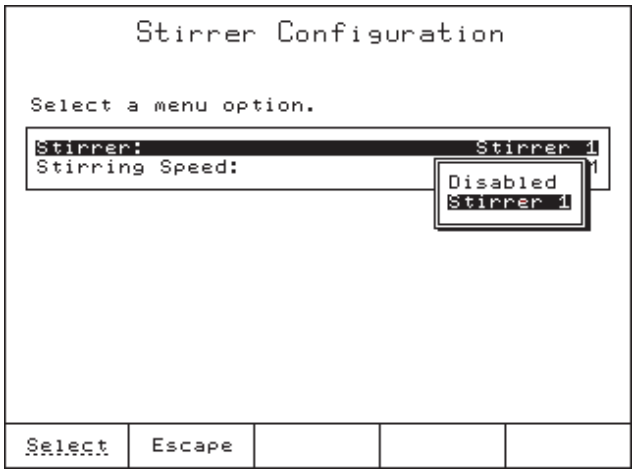
5.5.2 Method Revision

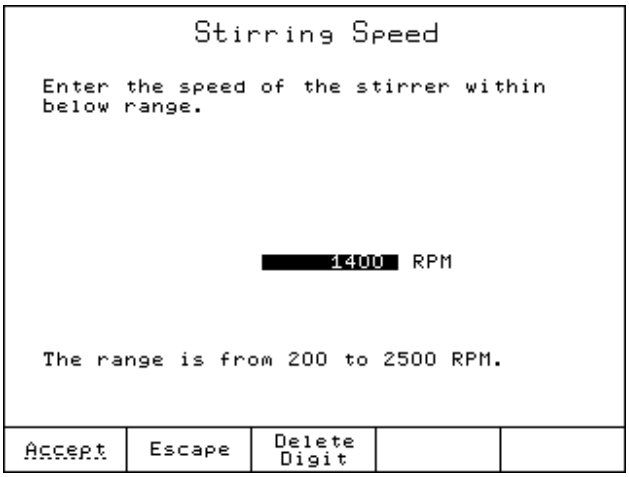
A string representing the current method revision can be entered. The revision string format should be “X.Y”, where X and Y are numerical digits.



5.5.3 Stirrer Configuration

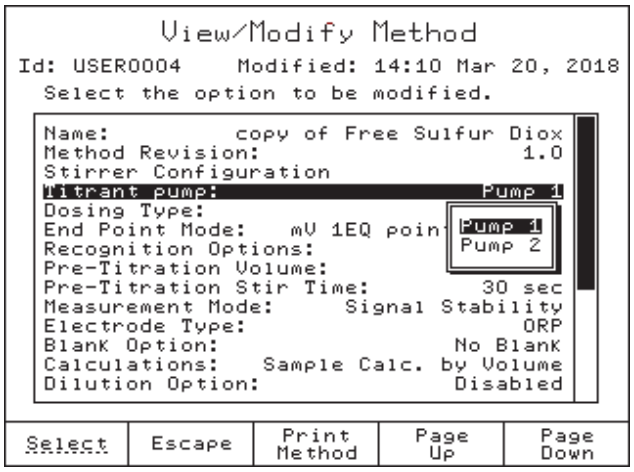
Select the stirrer to be used for the titration/analysis and set the stirrer speed.





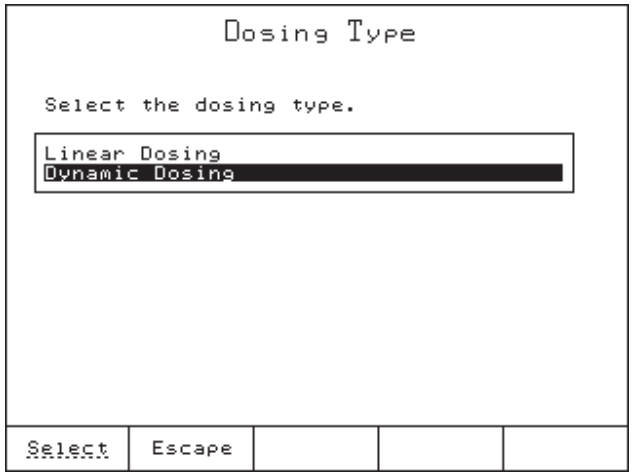
5.5.4 Pump Configuration

Choose the pump that will be used for the titration.



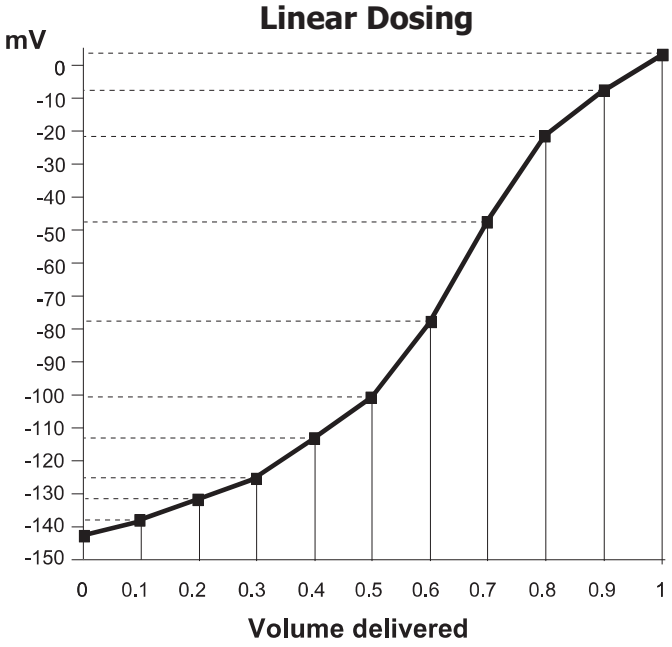
5.5.5 Dosing Type

The Titrator has two dosing types: *Linear Dosing* and *Dynamic Dosing*.



5.5.5.1 Linear Dosing

Linear dosing dispenses a pre-defined volume of titrant with every addition.



The *Linear Dosing* option is recommended for titrations with a slower reaction rate, difficult nonaqueous titrations, and specific applications.

Note: *For steep and normal titration curves, smaller volume increments are recommended, to obtain many points around the equivalence point.*
For flat titration curves, larger volume increments are recommended for equivalence point detection.

To set the dosing volume, select *Linear Dosing* and enter the optimum dose.

Dosing volume ranges are:

5 mL burette	0.001	to	4.750	mL
10 mL burette	0.001	to	9.500	mL
25 mL burette	0.005	to	23.750	mL
50 mL burette	0.005	to	47.500	mL

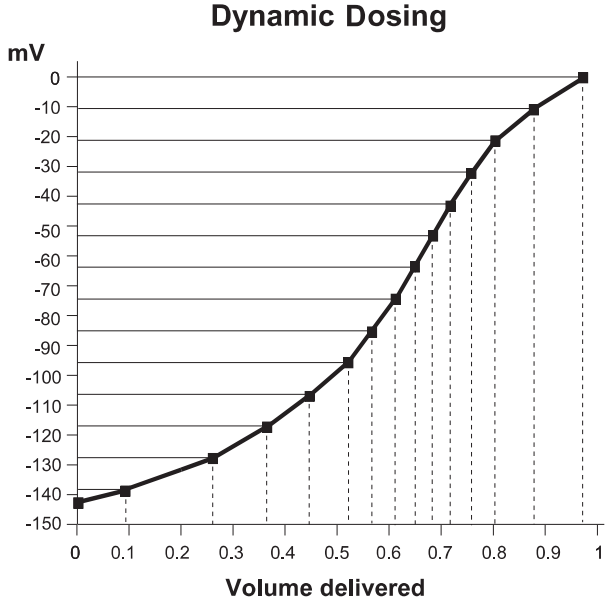
5.5.5.2 Dynamic Dosing

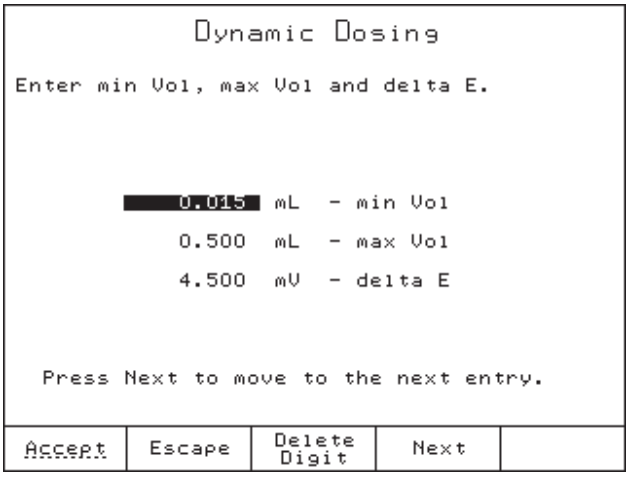
The Titrator determines the titrant dose by trying to maintain a certain potential change *(delta E)* with each addition.

After a titrant dose, if the potential change is lower than the set *delta E*, the next dose will be progressively increased until *max Vol* is attained. If the potential change is still lower than the set value, the titration will continue with *max Vol* doses.

After a titrant dose, if the potential change is higher than the set *delta E*, the next dose will be progressively decreased until *min Vol* is attained. If the potential change is still higher than the set value, the titration will continue with *min Vol* doses.

The titrant is added in volumes that depend on the proximity of the end point as shown in the graph below.





Dynamic dosing allows for larger doses far from the end point, reducing the total titration time. Closer to the end point, smaller doses are made, providing more data and improved accuracy.

The following parameters must be set:

min Vol: The smallest dose to be dispensed during a titration.
The *min Vol* must be greater than or equal to:
0.001 mL for a 5 mL burette
0.001 mL for a 10 mL burette
0.005 mL for a 25 mL burette
0.005 mL for a 50 mL burette

max Vol: The largest dose to be dispensed during a titration.
The *max Vol* must be less than or equal to 4.000 mL.

delta E: Sets the fixed potential jump that has to be achieved after each titrant dose.
The allowed range is between 0.1 and 99.999 mV.

Recommendations for dosing parameters:

For steep and normal titration curves the recommended settings are:

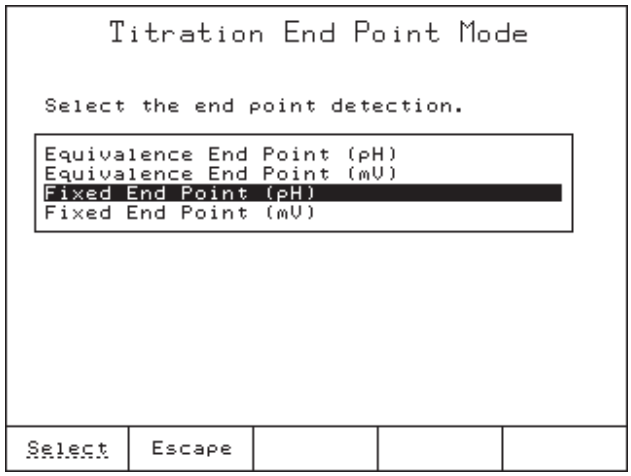
delta E	3.5	to	9 mV
min Vol	0.010	to	0.025 mL (for a 25 mL burette)
max Vol	0.075	to	0.250 mL (for a 25 mL burette)

For flat titration curves the recommended settings are:

delta E	10	to	15 mV
min Vol	0.050	to	0.150 mL (for a 25 mL burette)
max Vol	0.400	to	0.600 mL (for a 25 mL burette)

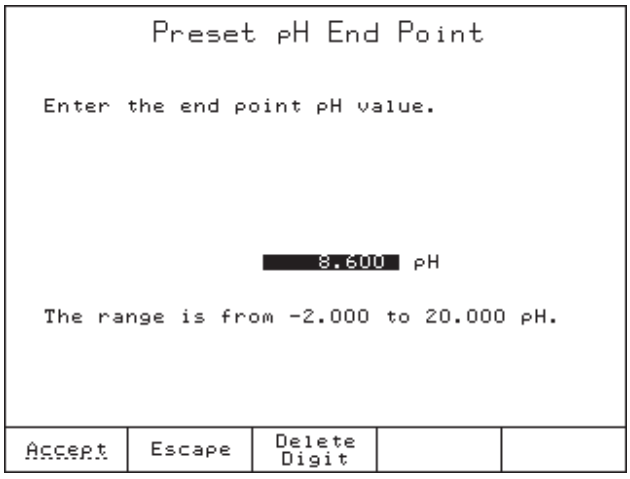
To achieve the highest levels of accuracy and reproducibility, it is recommended that 20-80% of the nominal burette volume used for each titration is consumed. If lower volumes of titrant are required, a smaller burette can be used.

5.5.6 End Point Mode

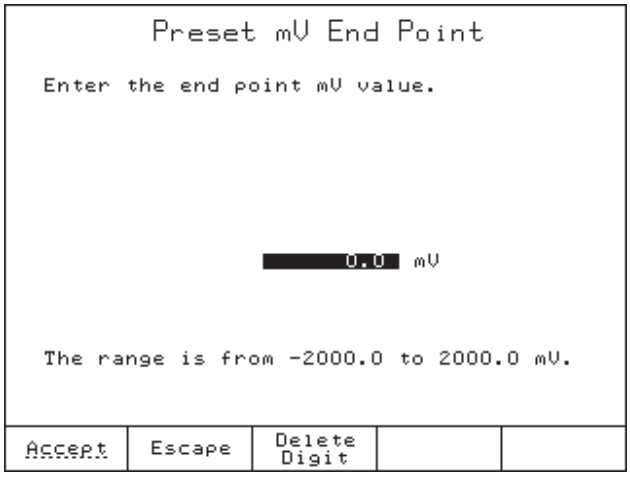


5.5.6.1 Fixed End Point (pH or mV)

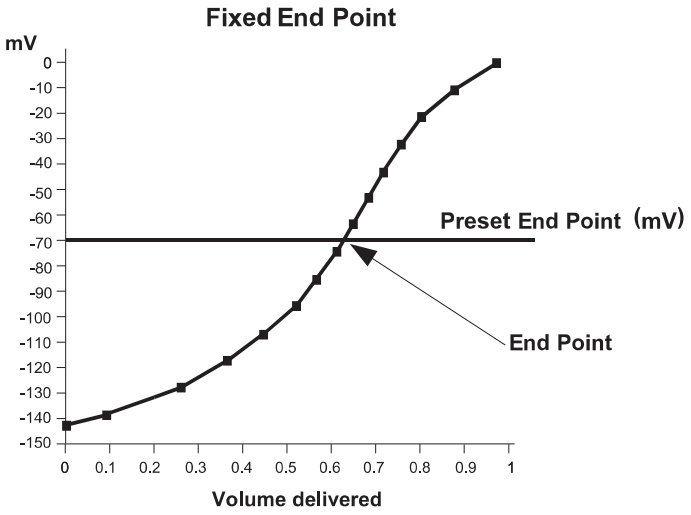
Fixed End Point (pH):
The titration is terminated when the preset pH value has been exceeded. The end point volume is a calculated value based on the dispensed volume when pH is under the preset value and the dispensed volume when pH exceeded the preset value.



The range is from - 2.000 to 20.000 pH.
Fixed End Point (mV):
The end point detection algorithm is the same as for pH, but the threshold value is expressed in mV.



The range is from - 2000.0 to 2000.0 mV.

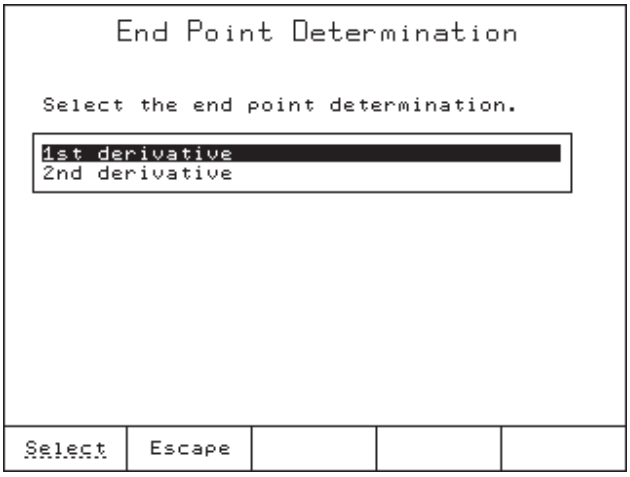


5.5.6.2 Equivalence End Point (pH or mV)

The titration is normally terminated when the equivalence point is detected (the point where the added quantity of titrant equals the quantity of analyte present in the sample).

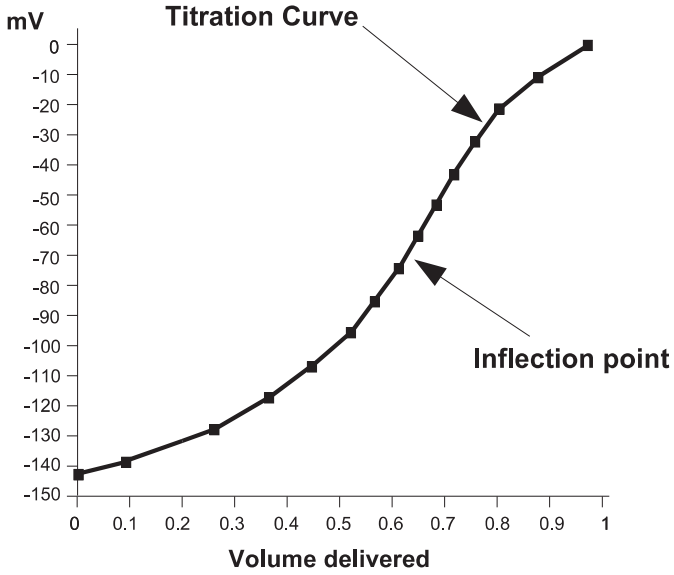
End Point Determination

The first and the second derivative of the titration curve can be used to detect the equivalence point.



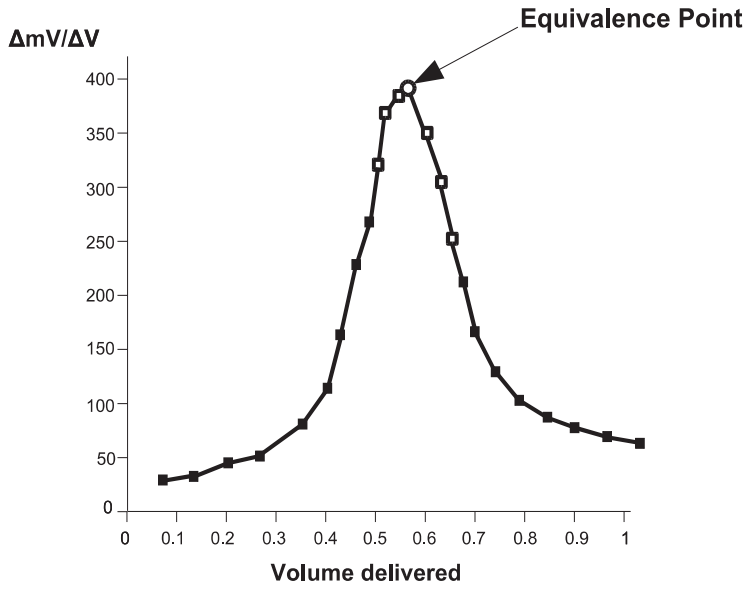
The equivalence point detection algorithm requires three additional titrant doses to be dispensed after the equivalence point is reached.
The reported end point volume is a calculated value based on a number of points around the equivalence point.
The potentiometric titration curve is the response in mV potential or pH between the indication of the electrode versus the volume of titrant added.
The inflection point of the titration curve is assumed to be the equivalence point of the

chemical reaction.
For non-symmetric titration curves, the theoretical error can be reduced by using the dynamic dosing.



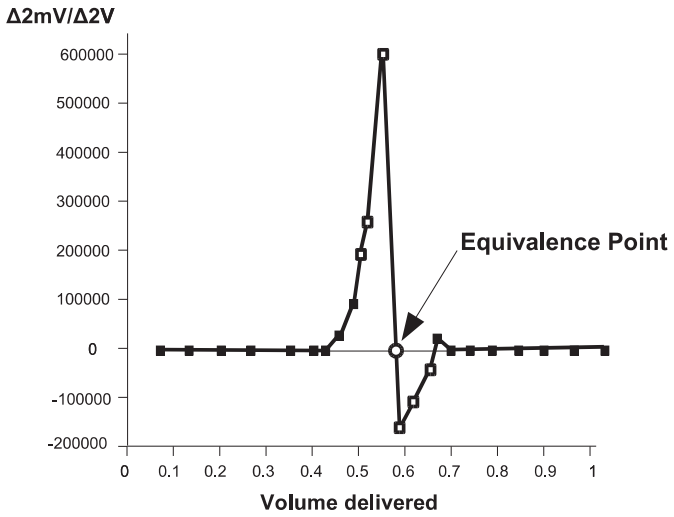
1st Derivative:

When first derivative is used to recognize the equivalence point, the titration curve inflection point (EQP) is the point where the first derivative reaches its maximum value.



The detection algorithm looks for the maximum value of the first derivative. The first derivative must be greater than the threshold value at the maximum point (see ***Recognition Options*** section).

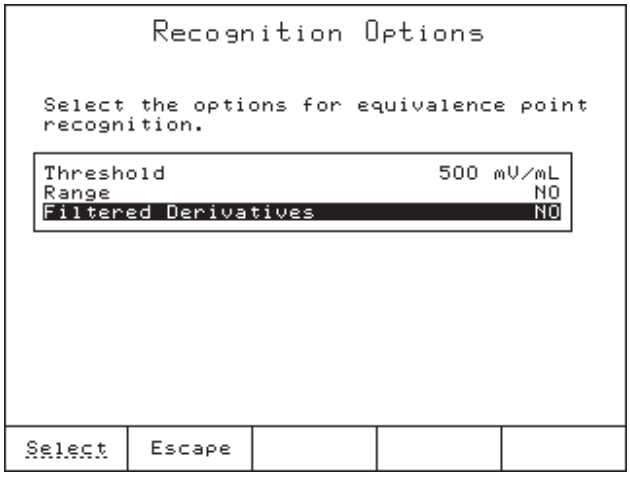
2nd Derivative:
When second derivative is used to recognize the equivalence point, the titration curve inflection point (EQP) is the point where the second derivative crosses zero.



The detection algorithm looks for the point where the second derivative changes sign.
The checked point, or first derivative, must be greater than the threshold value (see ***Recognition Options*** section).

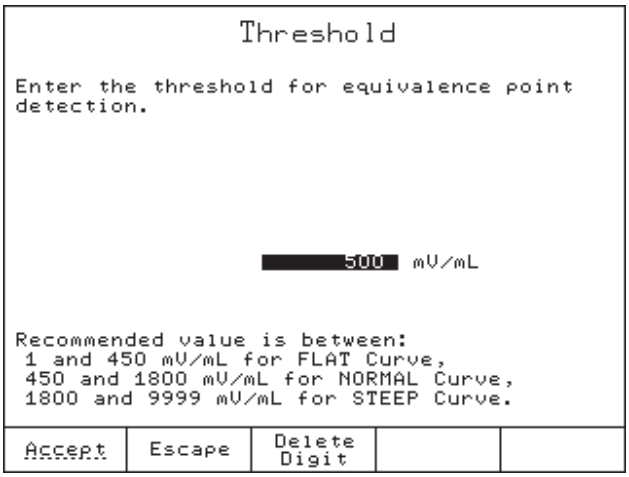
5.5.7 Recognition Options (Equivalence End Point only)

The ***Recognition Options*** screen is a set of parameters used to avoid false detection of the equivalence point due to the chemical system (titrant / sample species and concentrations) and / or electrode response.
The ***Recognition Options*** screen is available only when *Equivalence End Point (pH or mV)* option is selected.

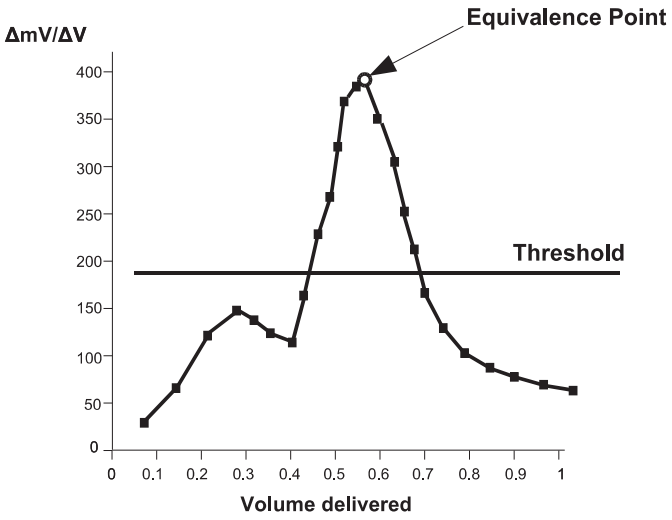


5.5.7.1 Threshold

This parameter must be set by the user according to the analysis.
The threshold represents the absolute value of the first derivative, expressed in mV/mL, below which the detection algorithm does not search for the equivalence point.



Range is between 1 and 9999 mV/mL.
The recommended value is 40% of the absolute value of the first derivative.

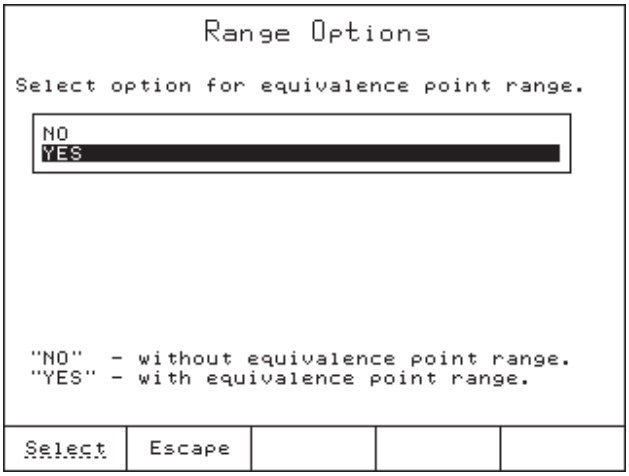


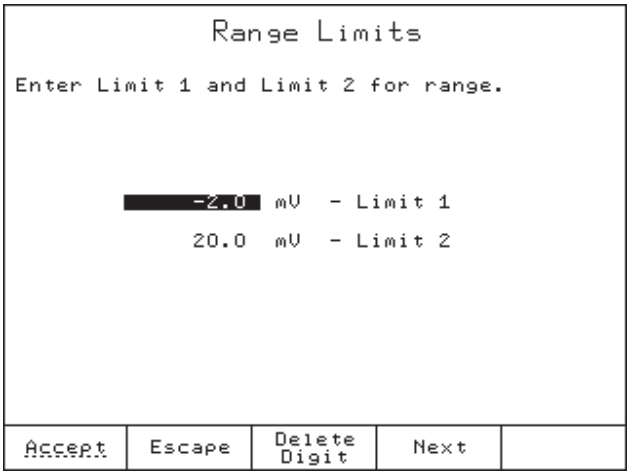
Depending on the titration curve profile, the following guide can be used:

TITRATION CURVE PROFILE	THRESHOLD (mV/mL)
Flat	1 to 450
Normal	50 to 1800
Steep	1800 to 9999

5.5.7.2 Range

Range is an optional feature for equivalence point recognition. The Titrator will only look for an equivalence point between the set values.
The *Range* option can be enabled by selecting *YES* in the ***Range Options*** screen.

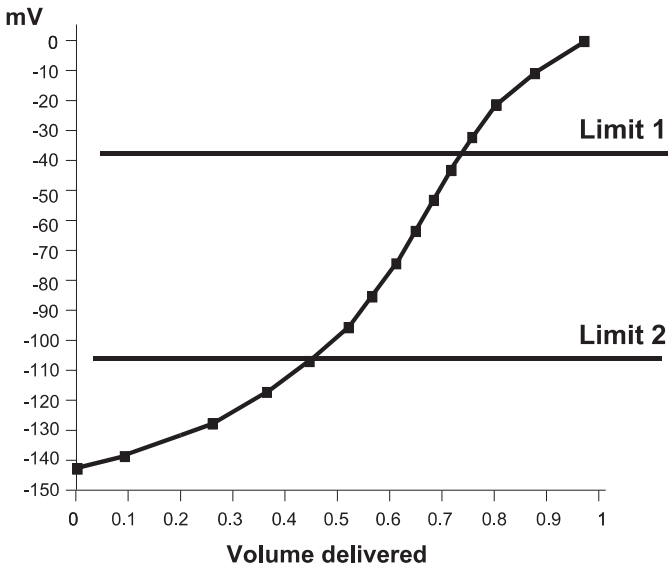




pH Range -2.000 to 20.000 pH

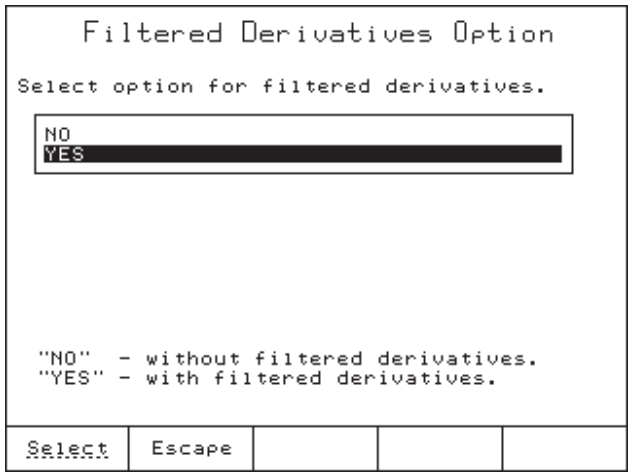
mV Range -2000.0 to 2000.0 mV

The Limit 2 value must not be equal to the Limit 1 value.



5.5.7.3 Filtered Derivatives

This option adds a filtering procedure in the 1st and 2nd derivative computation algorithm that reduces the influence of pH or mV noise.
The *Filtered Derivatives* option can be enabled by selecting *YES* in the ***Filtered Derivatives Option*** screen.



Noise can be due to:

- Chemical system properties (sample, titrant, solvent), such as slow chemical reactions or unbuffered samples such as wastewater, tap water, wine
- Electrode response
- Incorrect method parameters settings such as *Signal Stability, Stirring Speed*, etc.
- Insufficient titrant additions

Note*: A shift in the end point volume by 1 or 2 doses may be seen due to filtering.*

5.5.8 Pre-Titration Volume

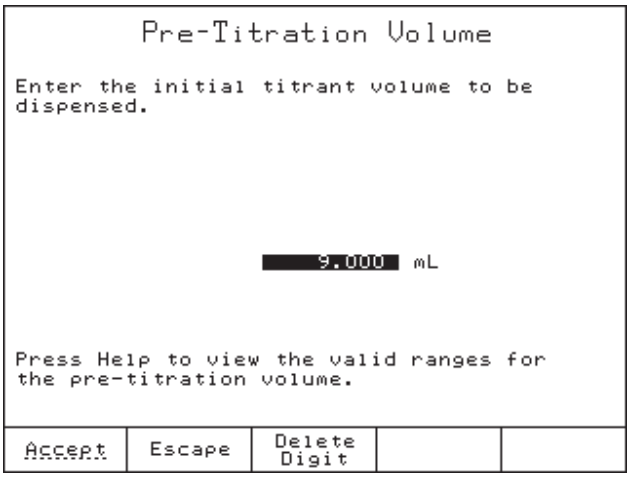
During a titration, the equivalence point is reached after many titrant doses. These doses take up extra time while having no relevance for equivalence point detection.

Pre-titration volume adds a large initial dose to jump directly to the proximity of the equivalence point.

This first dose occurs after the pre-titration stir time is completed.

The ranges for pre-titration volumes are shown below:

0.001 to 4.750 mL for a 5 mL burette
0.001 to 9.500 mL for a 10 mL burette
0.005 to 23.750 mL for a 25 mL burette
0.005 to 47.500 mL for a 50 mL burette

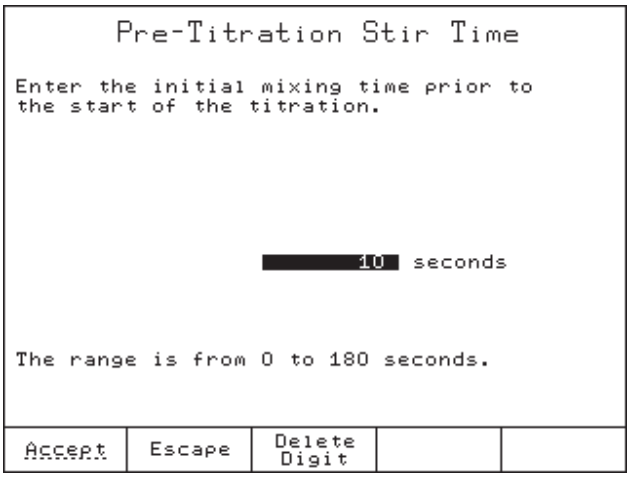


To disable a pre-titration volume, enter 0.000 mL.

Note: *A pre-titration volume is highly recommended whenever possible. Fewer doses will considerably shorten the overall titration duration.*

5.5.9 Pre-Titration Stir Time

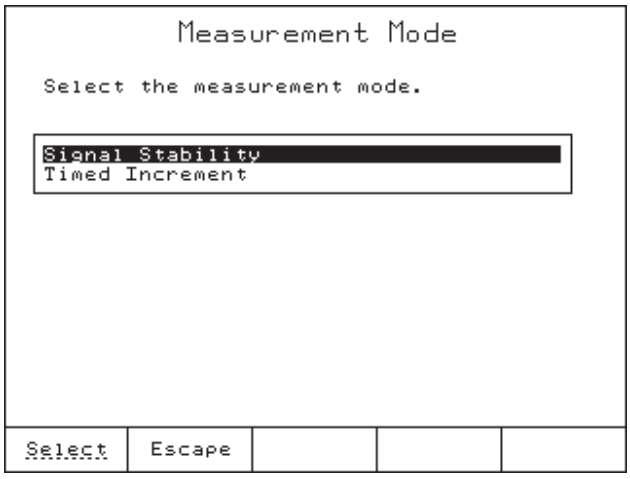
When enabled, the sample is mixed for a set period of time before any titrant is added. This allows the sample to become homogeneous.
The range is from 0 to 180 seconds.



The *Pre-Titration Stir Time* option is disabled if 0 seconds is entered.

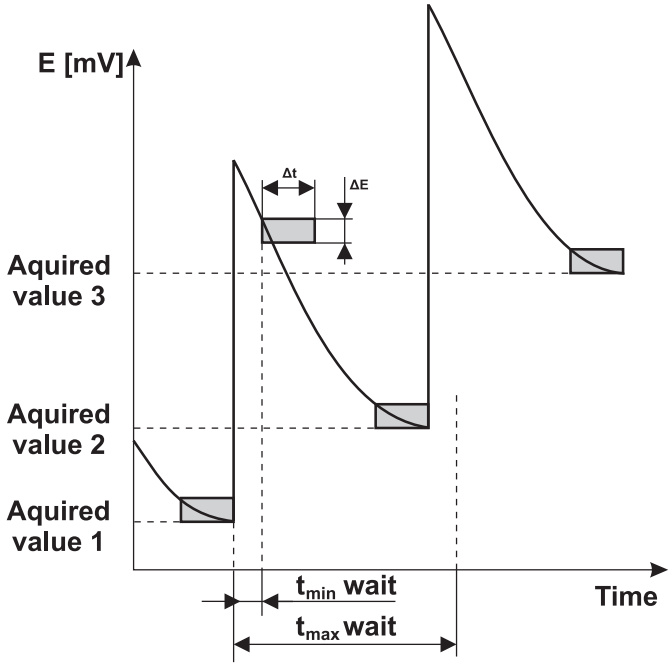
5.5.10 Measurement Mode

During titration, the acquisition of the potential (mV) value of the solution can be done in two ways: by using either *Signal Stability* or *Timed Increment* option.



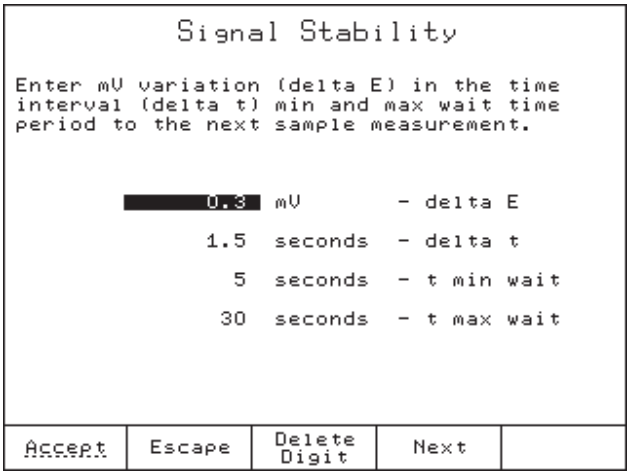
5.5.10.1 Signal Stability

When *Signal Stability* is selected, the Titrator acquires the potential (mV) only when stable conditions are reached.
The principles of signal stability are plotted below:



The signal stability window (condition) represents the time interval (Δt) during which the potential measured in solution (mV) is confined inside the potential interval (ΔE).
The new signal value is acquired if the stability condition is reached after the minimum (t min) wait time.

If the stability condition is not reached and the maximum (t max) wait time has elapsed, the potential is acquired.



delta E - maximum change in potential during delta t

The range is from 0.1 to 99.9 mV.

delta t - the time interval during which the potential is measured.

The range is from 0.5 to 10.0 seconds.

t min wait - the minimum elapsed time before a stability check. This is also the minimum elapsed time between two doses.

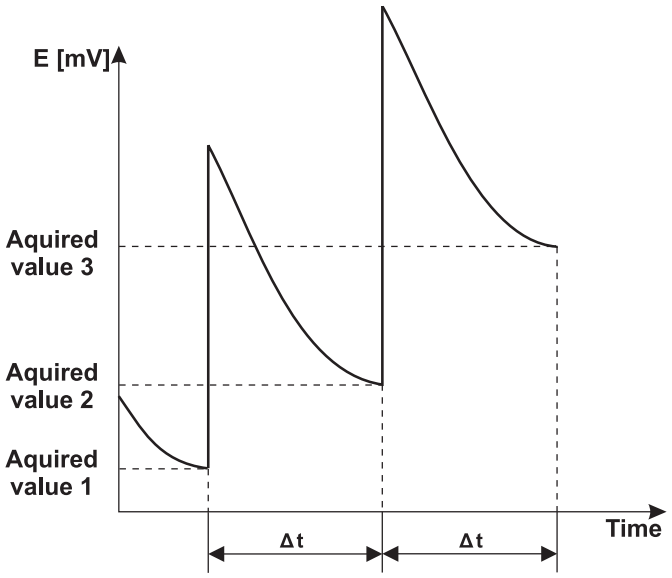
The range is from 2 seconds to *t max wait* time.

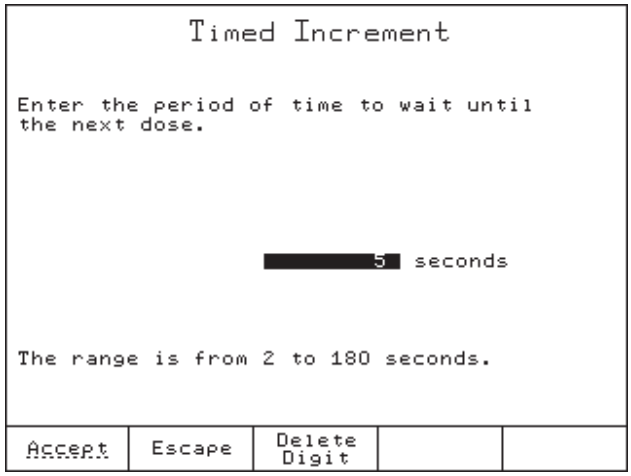
t max wait - the maximum elapsed time between two successive doses. If the *t max wait* has elapsed, a new dose is added even if the signal stability condition is not reached.

The range is from *t min wait* time to 180 seconds.

5.5.10.2 Timed Increment

When *Timed Increment* is selected, the Titrator acquires the potential (mV) at a fixed time interval (no signal stability check).
The time period between two acquisitions must be set according to the reaction and the response time of the electrode.

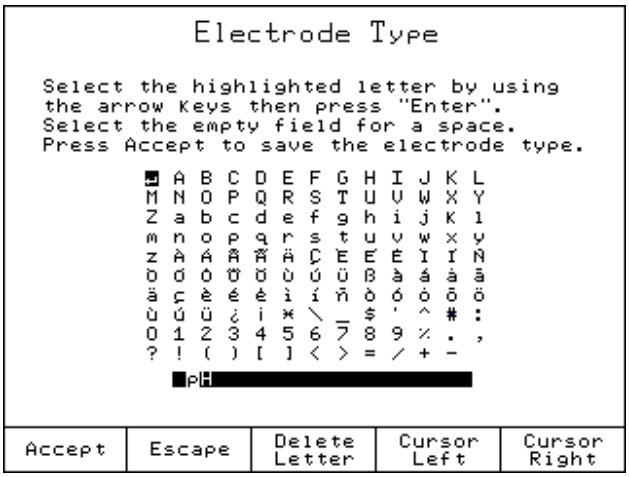




The range is from 2 to 180 seconds.

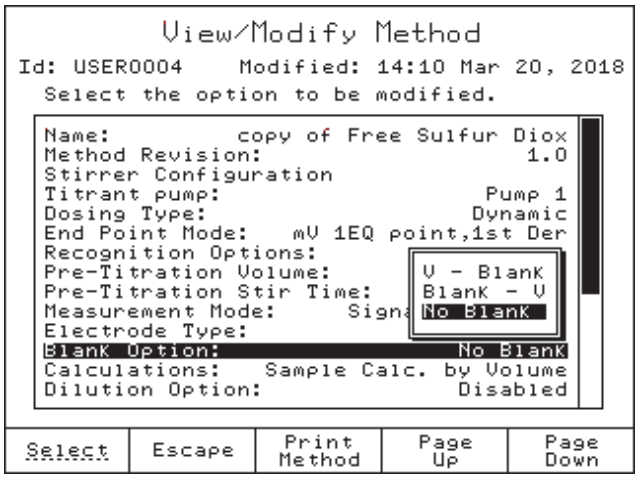
5.5.11 Electrode Type

Enter the type of the electrode, up to 20 characters.

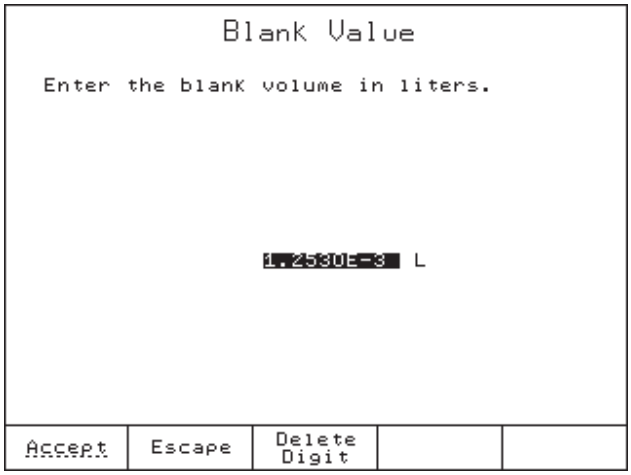


5.5.12 Blank Option

This feature allows the user to select the procedure for the blank calculations (where *V* is the volume of titrant dispensed during the titration and *Blank* is the volume of titrant consumed by the blank sample).

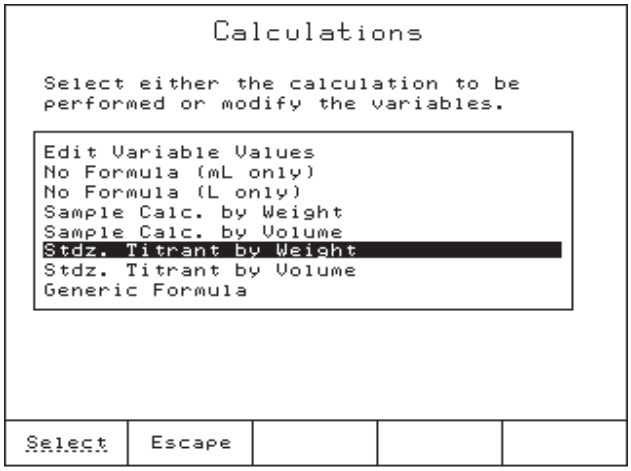


If one of the options (*V-Blank* or *Blank-V*) is selected in the ***View / Modify Method*** screen, the *Blank Value* will be active on the View/Modify Method screen and the value of the blank can be set (in liters).



5.5.13 Calculations

The final result is calculated using the end point volume (titrant volume at the equivalence point or at the fixed end point), and a formula selected by the user.



5.5.13.1 Edit Variable Values

Edit the variables in a previously selected calculation.
For each formula, selected variables can be changed.

5.5.13.2 No Formula (mL only)

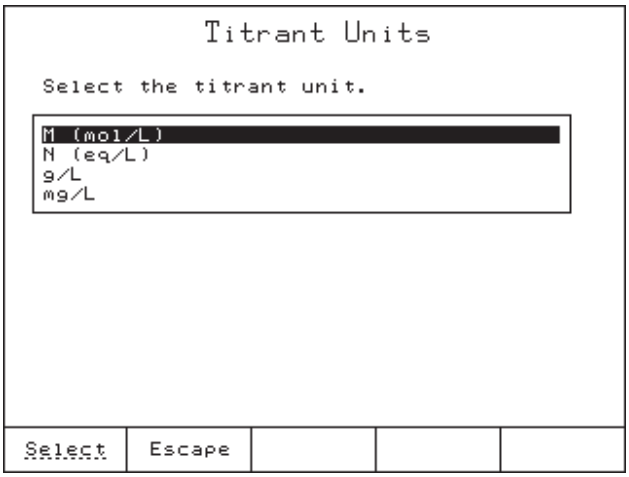
Only the volume of titrant (mL) required to reach the end point will be displayed.

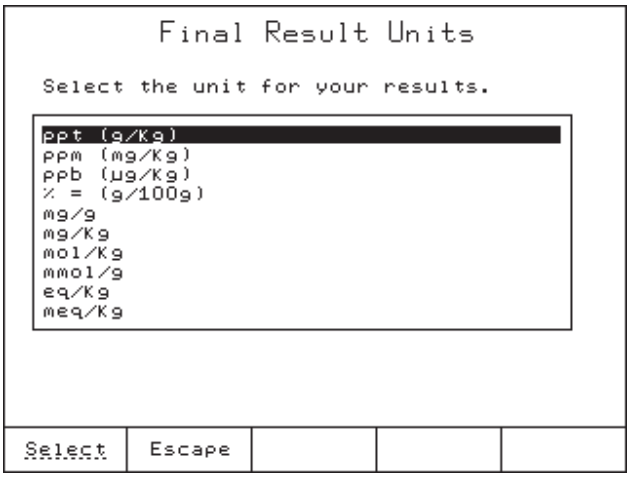
5.5.13.3 No Formula (L only)

Only the volume of titrant (L) required to reach the end point is displayed.

5.5.13.4 Sample Calculations by Weight

This calculation is used when the concentration of an analyte is determined by the weight of the sample. The results are based on the initial sample weight (in grams).
The Titrator will calculate the results based on the selected units.





The Titrator will provide the results based on the titrant and sample units selected.

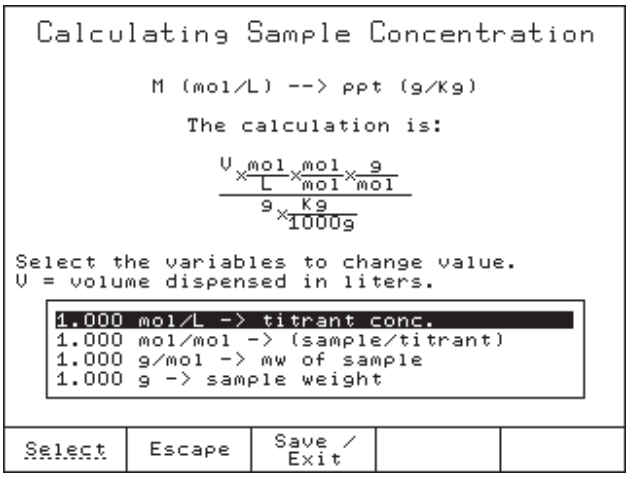
Titrant Units:

M (mol/L)	moles/liter
N (eq/L)	equivalents/liter
g/L	grams/liter
mg/L	milligrams/liter

Final Result Units:

ppt (g/kg)	parts per thousand (grams/kilogram)
ppm (mg/kg)	parts per million (milligrams/kilogram)
ppb (µg/kg)	parts per billion (micrograms/kilogram)
% (g/100 g)	percentage in weight (grams/100 grams)
mg/g	milligrams/gram
mg/kg	milligrams/kilogram
mol/kg	moles/kilogram
mmol/g	millimoles/gram
eq/kg	equivalents/kilogram
meq/kg	milliequivalents/kilogram

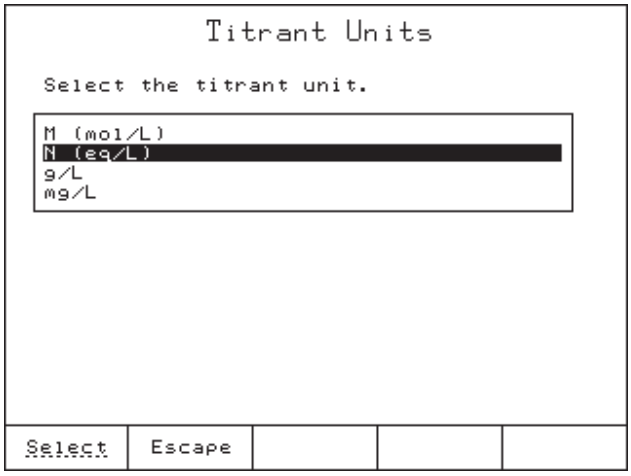
A formula example is shown below using M (mol/L) as the titrant unit and ppt (g/kg) as the final result unit:

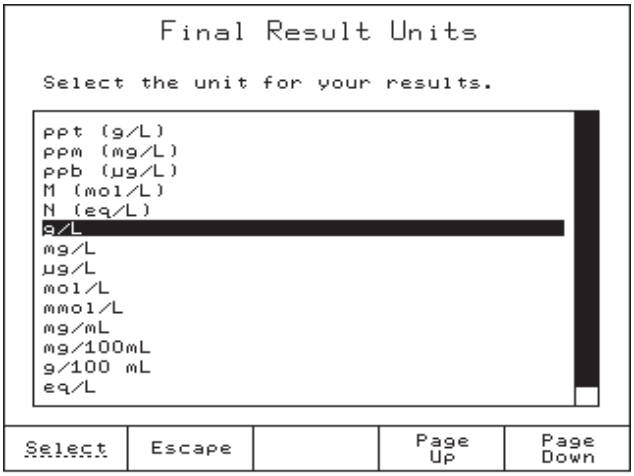


Variables can be set according to the amount of sample and titrant used.

5.5.13.5 Sample Calculations by Volume

This calculation is used when the concentration of an analyte is determined in terms of the volume of sample. The results are based on the initial sample volume (in milliliters).
The Titrator will calculate the results based on the selected units.





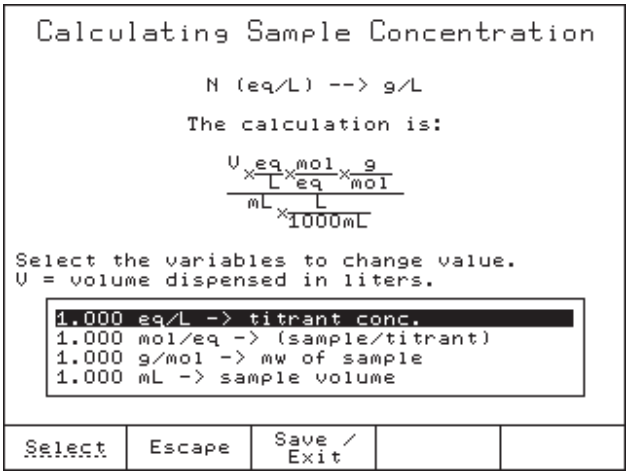
Titrant Units:

M (mol/L)	moles/liter
N (eq/L)	equivalents/liter
g/L	grams/liter
mg/L	milligrams/liter

Final Result Units:

ppt (g/L)	parts per thousand (grams/liter)
ppm (mg/L)	parts per million (milligrams/liter)
ppb (µg/L)	parts per billion (micrograms/liter)
M (mol/L)	Molarity (moles/liter)
N (eq/L)	Normality (equivalents/liter)
mg/L	milligrams/liter
µg/L	micrograms/liter
mmol/L	millimoles/liter
mg/mL	milligrams/milliliter
mg/100 mL	milligrams/100 milliliters
g/100 mL	grams/100 milliliters
eq/L	equivalents/liter
meq/L	milliequivalents/liter

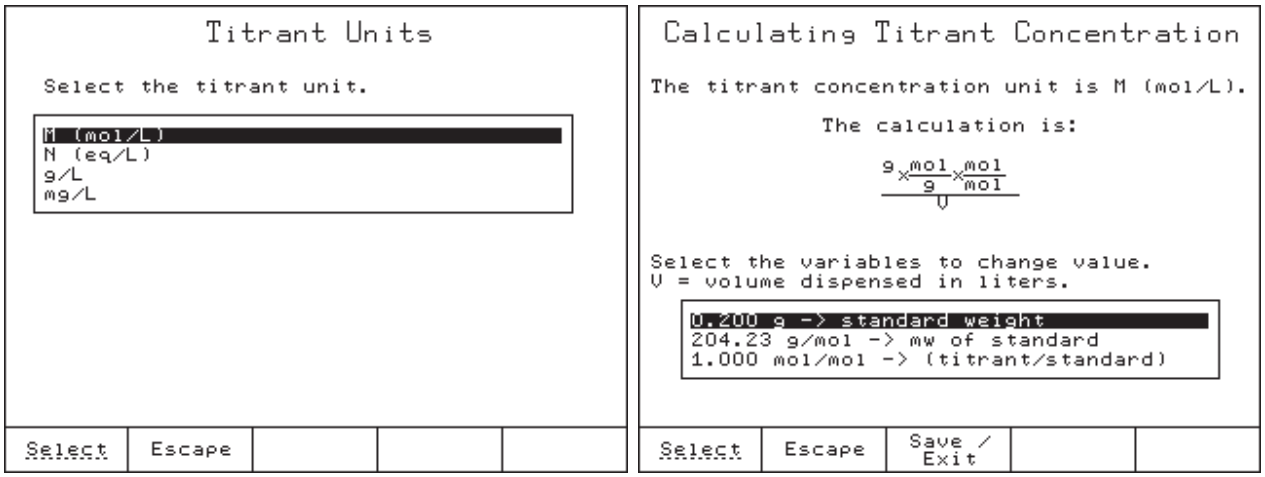
A formula example is shown below using N (eq/L) as the titrant units and g/L as the final result units:



Variables can be set according to the amount of sample and titrant used.

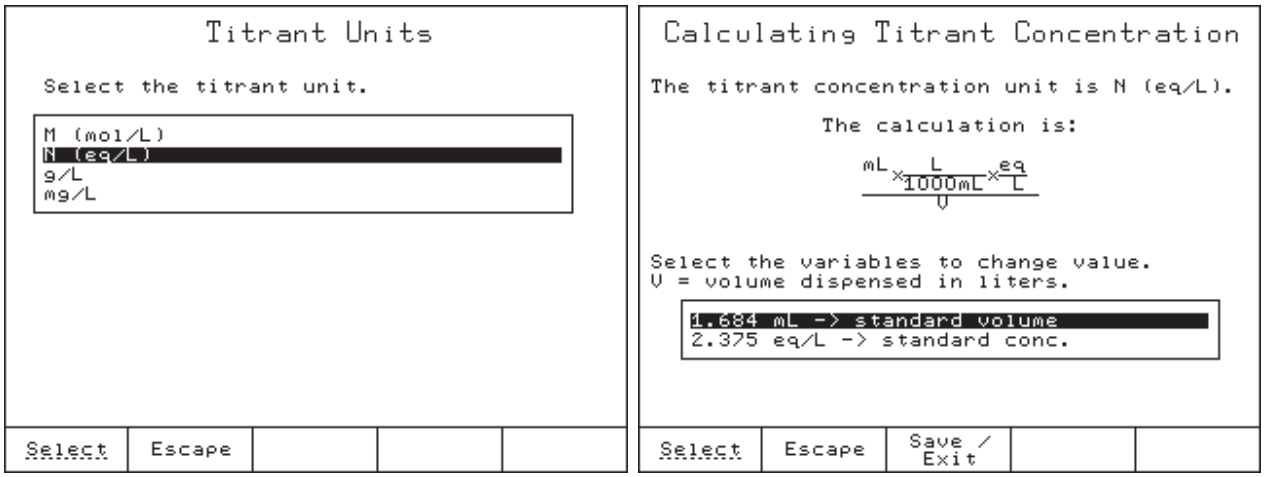
5.5.13.6 Standardize Titrant by Weight

This calculation is used when the concentration of the titrant is determined using a solid standard. Determination of the titrant concentration is based on the primary standard weight (in grams).
The calculation is based on the selected titrant unit. If the titrant unit is M (mol/L), the formula used to calculate the result is displayed below:



5.5.13.7 Standardize Titrant by Volume

This calculation is used when the concentration of the titrant is determined using a primary standard solution. Determination of the titrant concentration is based on the primary standard volume (in milliliters).
The Titrator will perform the calculation based on the titrant unit selected.
The calculation is based on the selected titrant unit. If the titrant unit is N (eq/L), the formula used to calculate the result is displayed below:



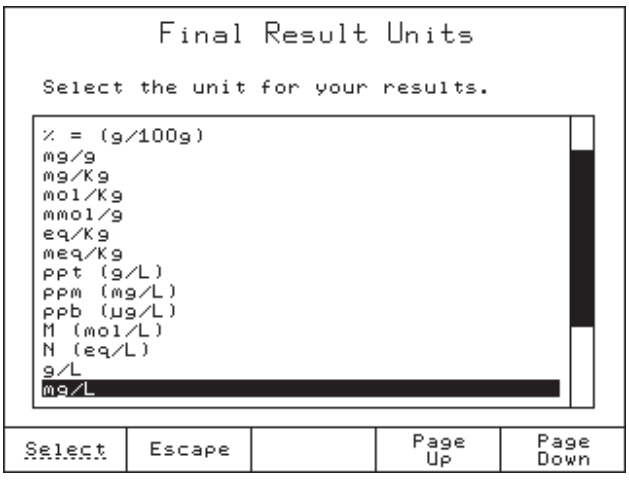
5.5.13.8 Generic Formula

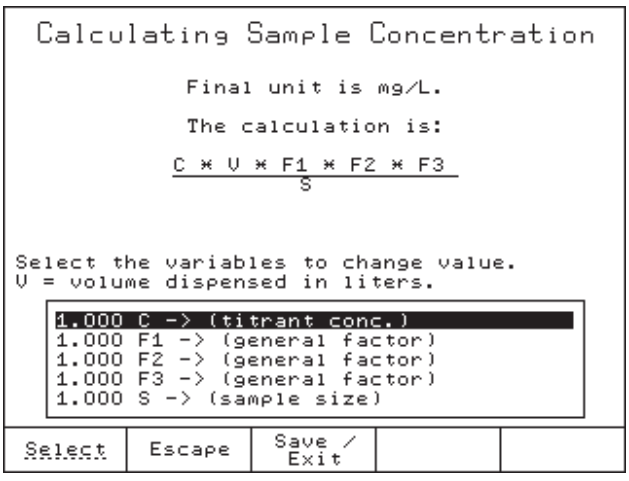
The user can define their own calculation formula based on the final result units in a solid or liquid sample.

Final Result Units:

ppt (g/kg)	parts per thousand (grams/kilogram)
ppt (g/L)	parts per thousand (grams/liter)
ppm (mg/kg)	parts per million (milligrams/kilogram)
ppm (mg/L)	parts per million (milligrams/liter)
ppb (µg/kg)	parts per billion (micrograms/kilogram)
ppb (µg/L)	parts per billion (micrograms/liter)
% (g/100 g)	percentage in weight (grams/100 grams)
M (mol/L)	Molarity (moles/liter)
mg/g	milligrams/gram
N (eq/L)	Normality (equivalents/liter)
g/L	gram/liter
mg/kg	milligrams/kilogram
mg/L	milligrams/liter
mol/kg	moles/kilogram
µg/L	micrograms/liter
mol/L	moles/liter
mmol/g	millimoles/gram
eq/kg	equivalents/kilogram
mmol/L	millimoles/liter
meq/kg	milliequivalents/kilogram
mg/mL	milligrams/milliliter
mg/100 mL	milligrams/100 milliliters
g/100 mL	grams/100 milliliters
eq/L	equivalents/liter
meq/L	milliequivalents/liter
No Unit	No result unit

The Titrator will calculate the results based on the selected unit.
The formula can be either for titrant standardization or sample analysis.
Where:





C = the concentration of the titrant
F1 = general factor
F2 = general factor
F3 = general factor
S = sample size, in grams or milliliters
V = the volume delivered, in liters, to reach the preset or equivalence end point (determined by the Titrator)

General factors:

Weight Conversion:
One of the general factors should be a weight conversion factor.
Examples of concentration units:

mol/L	moles/Liter
eq/L	equivalents/Liter
g/L	grams/Liter
mg/L	milligram/Liter

Reaction Ratio:
The reaction ratio is the ratio between the analyte and titrant or standard and titrant.
Examples of ratios:

mol/mol	moles of sample/moles of titrant
mol/eq	moles of sample/equivalents of titrant
eq/mol	equivalents of sample/moles of titrant
mol/mol	moles of titrant/moles of standard
eq/mol	equivalents of titrant/moles of standard

Example: 2 moles of NaOH react with 1 mole of H_2SO_4

Unit Conversion factor:

Used to convert between various measurement units.

Examples: L/1000 —> mL
g/1000 —> mg

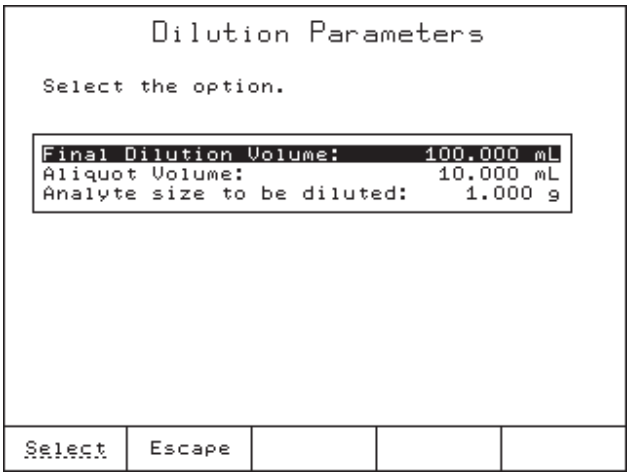
Weight Conversion factor:

Used to convert between weight measurement bases (kg, g, mg, µg or mole, mmole).

Example: g —> mol

5.5.14 Dilution Option

When the initial sample is diluted, a titration is made with an aliquot of the diluted sample, dilution calculations can be used.
The calculations are based on the original sample weight (volume) in order to express the results for the initial sample.



Final Dilution Volume: The volume of the sample after dilution

Aliquot Volume: Sample volume used for the titration

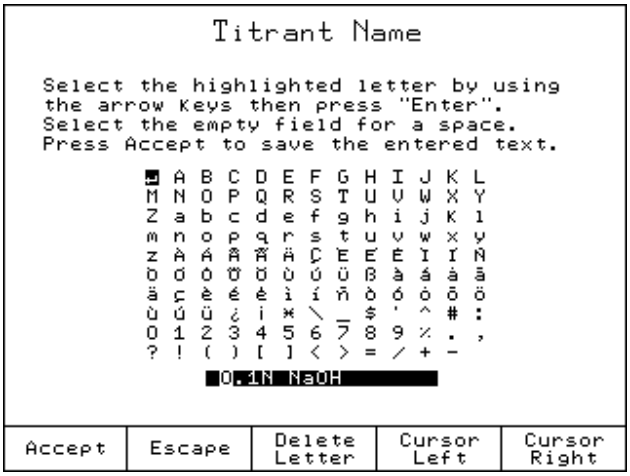
Analyte size to be diluted: The initial sample weight (volume)

The sample size used in the calculations:

$$\frac{\text{Analyte size to be diluted} * \text{Aliquot Volume}}{\text{Final Dilution Volume}}$$

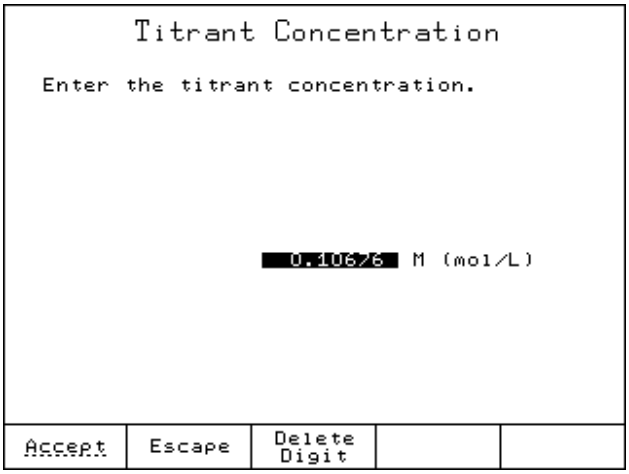
5.5.15 Titrant Name

Enter the name of the titrant (up to 20 characters). This name will appear in the titration report.



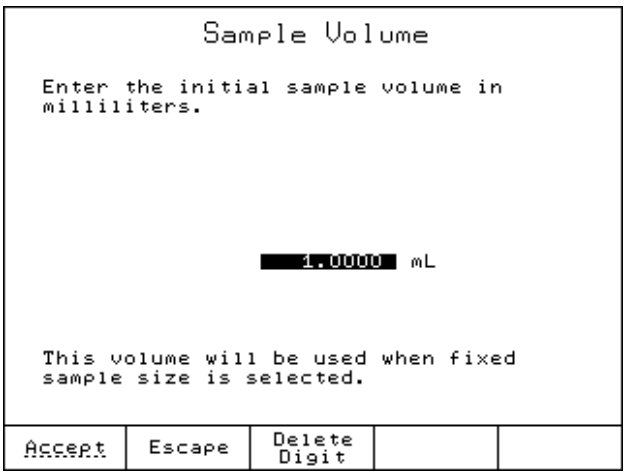
5.5.16 Titrant Concentration

Enter the concentration of the titrant to be used. When determining the titrant concentration, only the concentration unit is displayed. The titrant concentration can not be set.



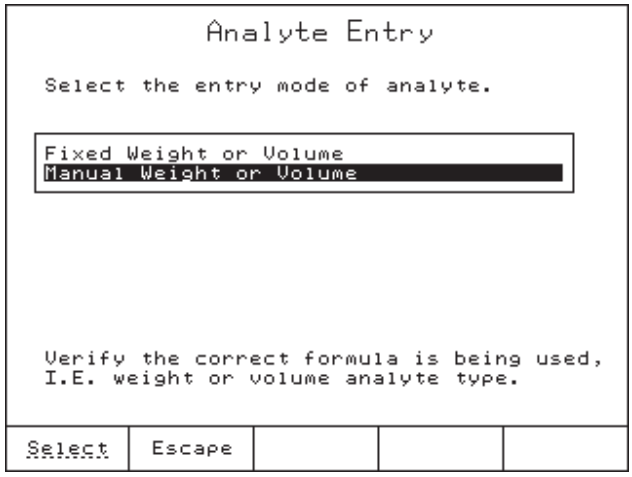
5.5.17 Analyte Size

Enter the size of the sample (for sample concentration determinations) or standard (for titrant concentration determination).



5.5.18 Analyte Entry

Select the analyte entry mode.



5.5.18.1 Fixed Weight or Volume

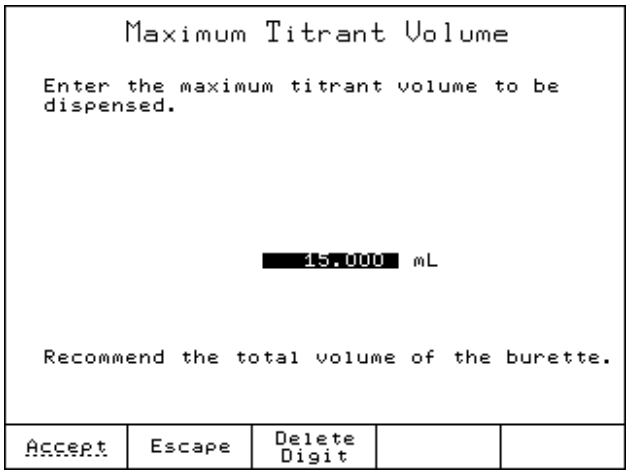
Each titration will use a set weight or volume in the calculations.

5.5.18.2 Manual Weight or Volume

Each titration, the exact weight or volume can be entered. The Titrator will prompt for the analyte weight or volume at the beginning of each titration.

5.5.19 Maximum Titrant Volume

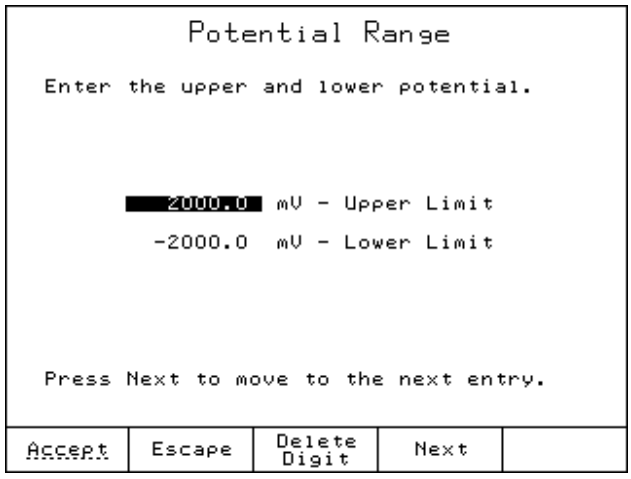
The maximum titrant volume used in the titration must be set according to the analysis. If the titration end point (fixed or equivalence End Point) is not reached, the titration will be terminated after the maximum titrant volume has been dispensed. The error message ("Limits Exceeded") will appear on the display.



Range is from 0.100 to 100.000 mL.

5.5.20 Potential Range

The input potential range can be set by the user. The titration will be terminated and an error message will appear if the potential is outside these limits.
These limits provide protection against a titration that does not generate an end point due to potential over-range.

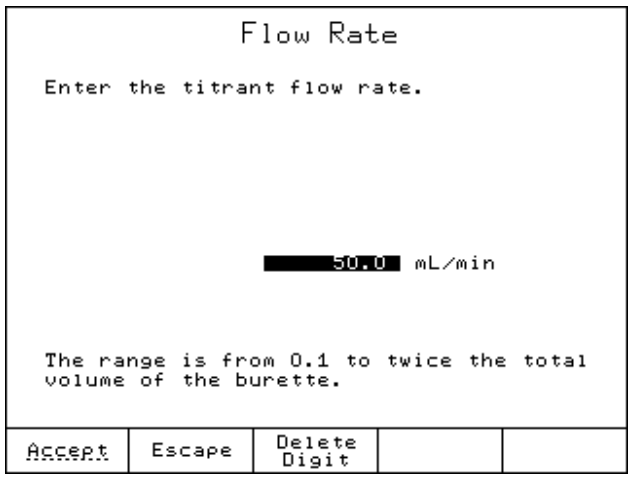


The ranges can be set between -2000.0 to 2000.0 mV.

5.5.21 Volume/Flow Rate

The flow rate for the dosing system can be set by the user in an interval of 0.1 to two times the burette volume:

0.1 to 10 mL/min for a 5 mL burette
0.1 to 20 mL/min for a 10 mL burette
0.1 to 50 mL/min for a 25 mL burette
0.1 to 100 mL/min for a 50 mL burette



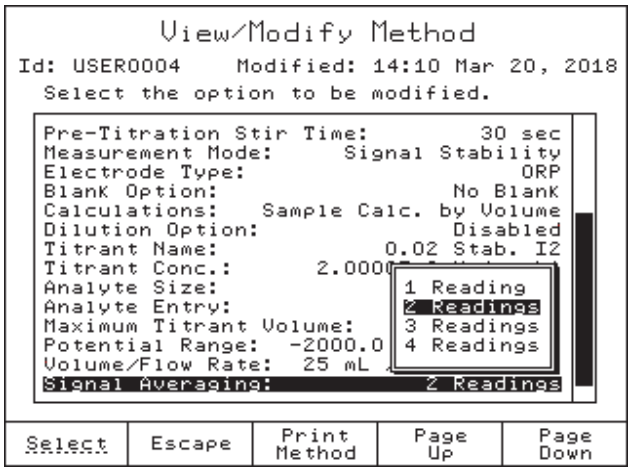
Note: *The Titrator will automatically detect the burette size and display the correct high limit volume.*

The flow rate is set for all burette operations.

5.5.22 Signal Averaging

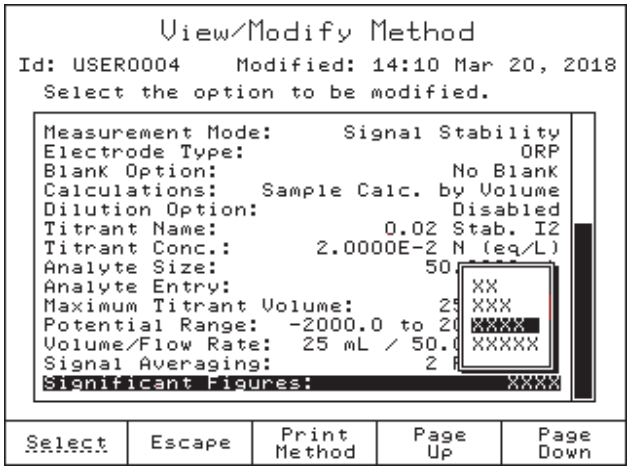
This option enables filtering on the mV/pH reading.
If *1 Reading* is selected, the filtering is disabled. The Titrator will take the last reading and place it into a "moving window" along with the last 2, 3 or 4 readings (depending on the selected option). The average of those readings is displayed and used for calculations.

Averaging more readings is helpful when a noisy signal is received from the electrode.



5.5.23 Significant Figures

This option allows you to set the format for displaying the final titration result.



5.6 Printing

To print method parameters, press Method Options from the main screen.

Press Print Method and wait a few seconds until the printer completes the job.
If no printer is connected to the dedicated socket, or if the printer is offline, an error message will appear on the display (see ***Connecting a Printer*** section, for information about connecting a printer to the Titrator).

Chapter 6. Contents

6 TITRATION MODE

6.1 Titration Start

Before beginning to perform a titration make sure that the following conditions are met:

- At least one pump is properly installed.
- A burette is inserted in the pump and filled with titrant.
- The aspiration tube is inserted in the titrant bottle and the dispensing tube is over the analyte beaker.
- The standard or sample has been carefully weighed / measured into the titration beaker.
- The electrode and the temperature probe is inserted in the analyte beaker.
- The desired method is selected as active and the parameters are set at optimum values.

6.1.1 In Progress Titration

To start a new titration, press start/stop from the main screen.
When a titration begins:

- The stirrer will turn on (if detected and enabled).
- If the pre-stirring time option is enabled, the sample will be stirred until the prescribed time elapses (see **Methods**, *Pre-Titration Stir Time* section*).*
- If the pre-titration volume option is enabled, the prescribed volume will be dispensed (see **Methods**, *Pre-Titration Volume* section*).*
- According to the *Measurement Mode* and the *Dosing Type* option, the titrator will start to deliver doses until the titration end point are detected or a titration stop condition occurs.

6.1.2 Suspend Titration

While titration is in progress, you can temporarily stop it by pressing Suspend. All the titration parameters will be frozen.
You can continue the titration by pressing Resume.

6.1.3 On-line Graph

During a titration, both the potentiometric S-shape curve and the selected derivative curve (titration with equivalence point only) can be displayed on the ***Titration Graph*** screen, by pressing View Curve. The titration ID report is also displayed inside the graph window.

The S-shape curve and the derivative curve are scaled to fit simultaneously inside the display. Also, when the titration is normally terminated (end point detected successfully), the end point volume marked with a cross is displayed on the graph.

The contents of the graph as related to an end point type, is as follows:

Equivalence End Point (pH) - the pH curve and the selected derivative vs volume is displayed (see Figure 1).

Equivalence End Point (mV) - the mV curve and the selected derivative vs volume is displayed (see Figure 2).

Fixed End Point (pH) - only the pH vs volume curve is displayed (see Figure 3).

Fixed End Point (mV) - only the mV vs volume curve is displayed (see Figure 4).

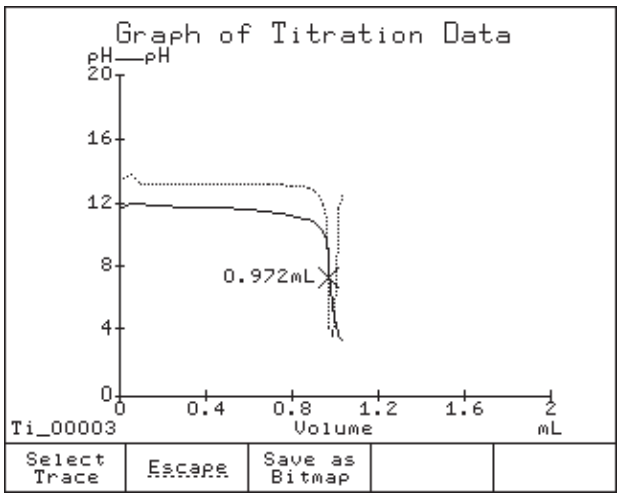


Figure 1

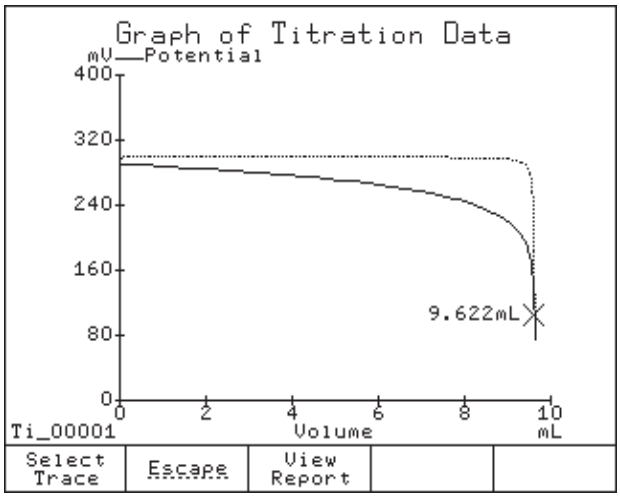


Figure 2

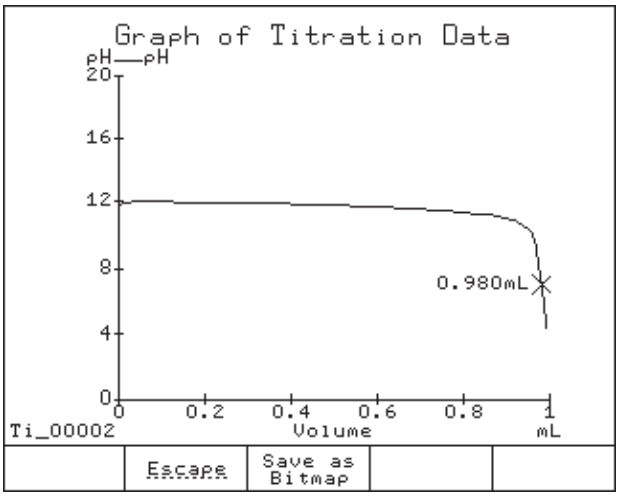


Figure 3

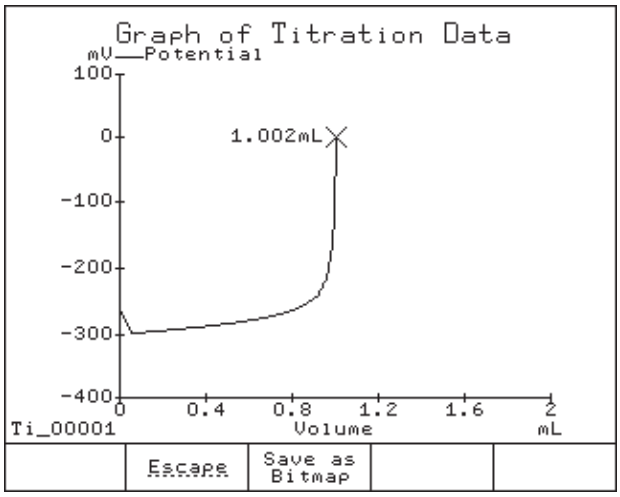


Figure 4

Select Trace - allows you to view on the ordinate axis a plot of either the mV (or pH) values or the selected derivative values (of mV or pH). Available only for titrations with equivalence end points.

Save as Bitmap - allows you to save the graph as a bitmap file. Available only when the titration is finished (after end point detection).

6.2 Titration Stop

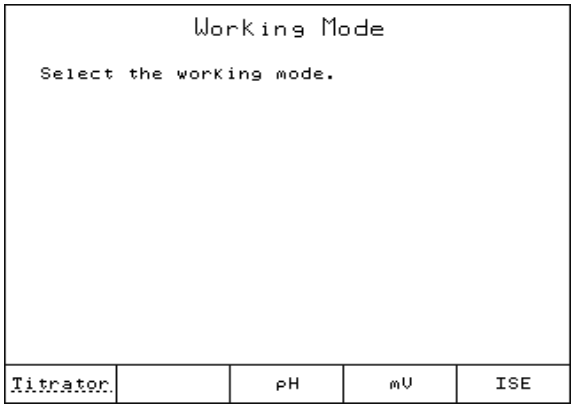
The titration can be finished in one of the modes described below:

- **Titration Completed.** The titration was successfully terminated (with end point successfully detected). This is the only mode with valid final result values.
- **Manually Terminated.** The current titration was manually terminated before end point detection was achieved.
- **Limits Exceeded.** The preset maximum titrant volume was delivered without reaching the end point. The titration is stopped with an error message.
- **Critical Error.** A critical error occurred and the titration was stopped. These errors are normally related to the dosing system. The titration is stopped with a specific error message.
- **Potential Out of Range.** The measured values from the input sensor are outside the preset range (potential range). The titration is stopped with an error message.

Chapter 7. Contents

7 pH MODE

By pressing [Mode] from the main screen, the Titrator can be switched to **Titrator**, **pH**, **mV** or **ISE** modes.



[Titrator] Switches to **Titrator** mode.

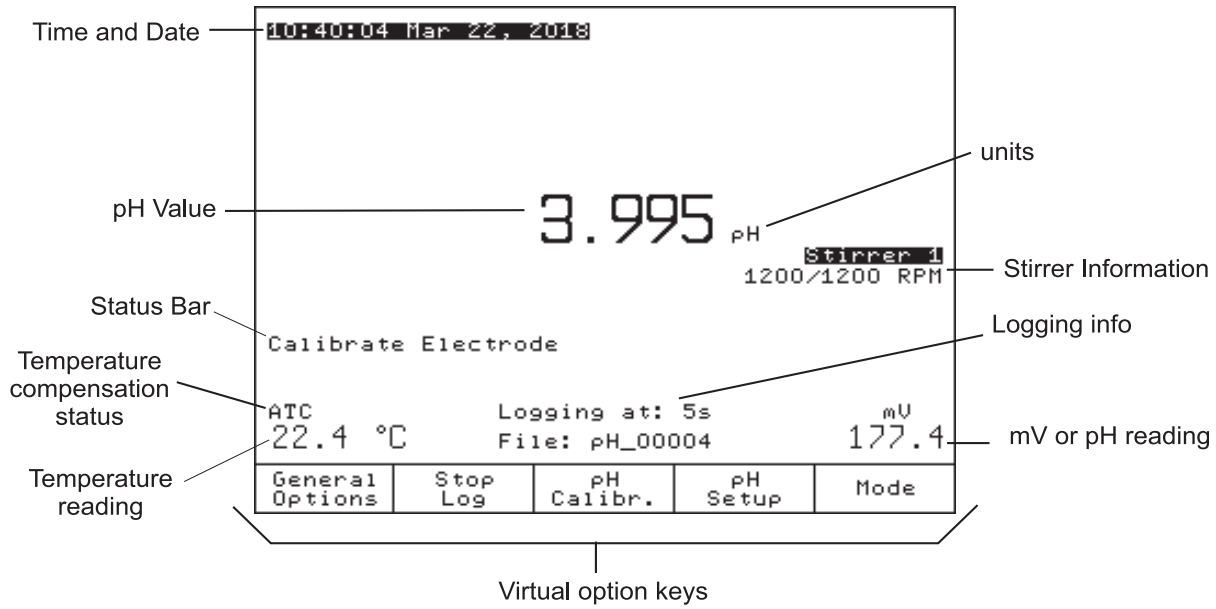
[pH] Switches to **pH** mode.

[mV] Switches to **mV** mode.

[ISE] Switches to **ISE** mode.

7.1 Display

The ***pH*** screen is shown below with short explanations of the screen segments.



pH Mode Option keys:

General Options — The General Options screen gives access to options that are not directly related to the measurement process (see *General Options* chapter for more information).

Save Reading — Stores the current pH reading (see *Manual Logging* section).

or

Start Log — Starts the pH automatic log (see *Automatic Logging* section).

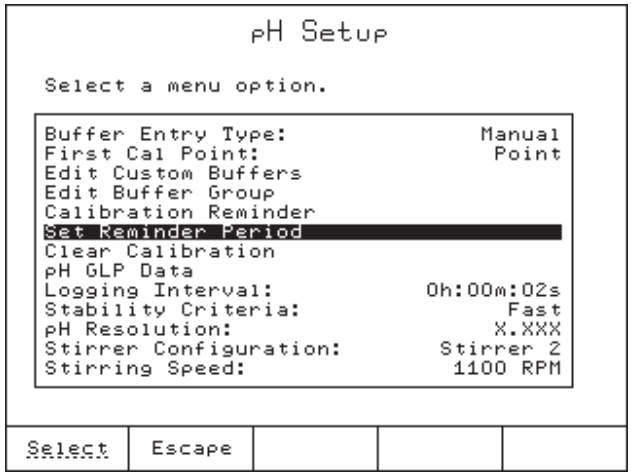
pH Calibr. — Enter the pH calibration screen (see *pH Calibration* section).

pH Setup — Enter the pH setup screen, parameters are associated with pH measurements and calibration (see *pH Setup* section).

Mode — Allow the user to switch between the available measurement modes: **Titrator**, **pH**, **mV** or **ISE** mode.

7.2 pH Setup

To access pH Setup, press [pH Setup] option key while in pH mode.



Use △ and ▽ keys to highlight the desired option.

Press [Select] or [enter] to access the selected option.

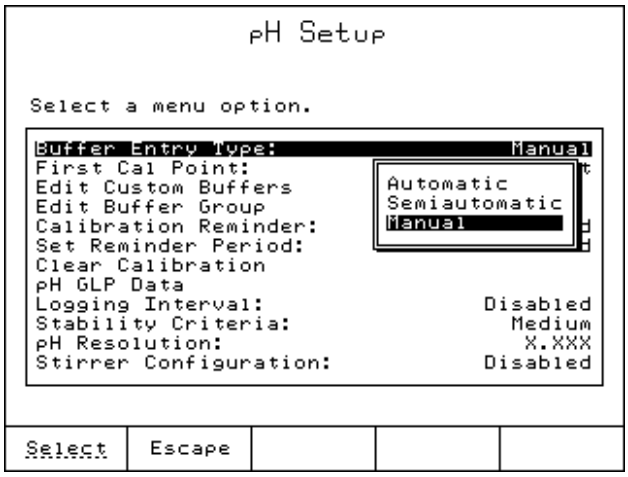
7.2.1 Buffer Entry Type

Select the pH buffer entry mode used for calibration:

Automatic - the instrument automatically selects the pH calibration point as the closest buffer from the predefined Buffer Group (see *Edit Buffer Group* section).

Semiautomatic - the instrument automatically selects the closest buffer from Available Buffers (standard and custom buffers).

Manual - the calibration buffer must be manually selected by the user during calibration from the available buffer list (standard and custom buffers).

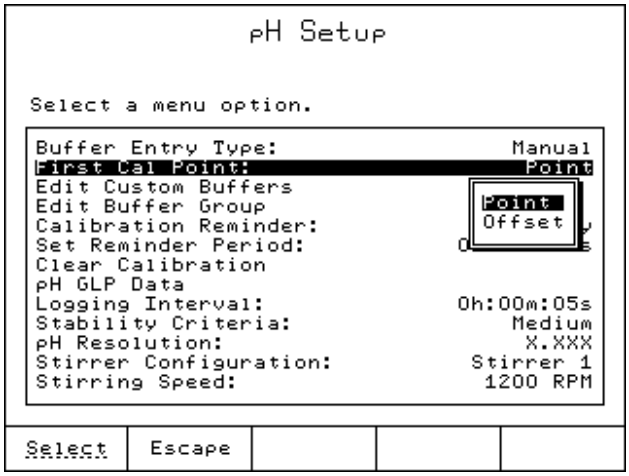


7.2.2 First Calibration Point

Two options are available for the First Calibration Point: *Point* and *Offset*.

If *Point* option is selected, the slope values adjacent to the calibration points will be reevaluated (normal calibration).

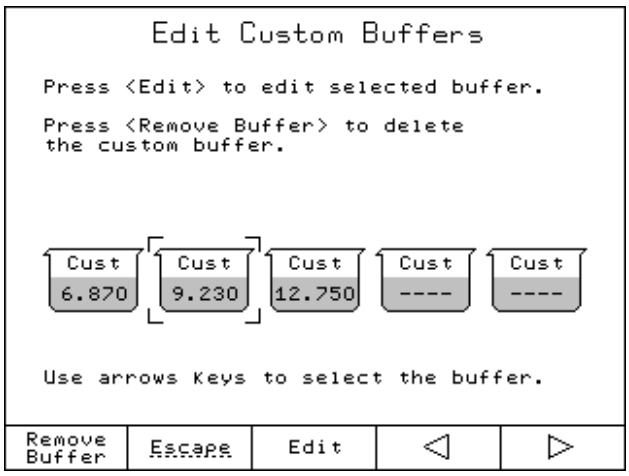
If at least a two-point calibration has been performed and an offset correction is needed, perform a one-point calibration using the *Offset* option. The existing slope values will not be changed.



7.2.3 Edit Custom Buffers

If you wish to use buffers other than the standard ones, the Edit Custom Buffers option is available, allowing you to set the desired pH buffers. Up to five pH custom buffers can be set.

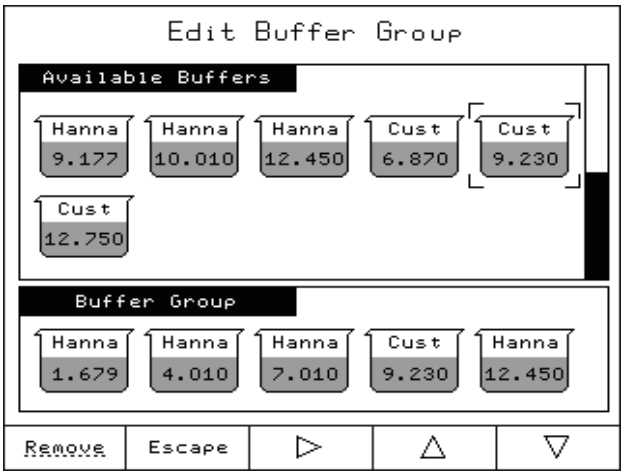
Note: *Custom buffers are not temperature compensated. The value of the buffer at the calibration temperature should be entered. The standard buffers are automatically temperature compensated.*



- Use ◁ and ▷ keys to select the desired buffer.
- Press [Remove Buffer] to delete the custom buffer.
- Press [Edit] to edit the selected buffer; use the numeric keys to edit the buffer values.
- Press [Accept] to save the value.
- Press [Escape] to return to pH Setup menu.

7.2.4 Edit Buffer Group

Select up to five buffers from the available buffers (Hanna or Custom) to be used for automatic buffer recognition (Automatic Buffer Entry Type).
Within the Buffer Group, pH values must be at least 1.5 pH far apart.
If the Buffer Group already contains five pH buffers, at least one pH buffer has to be removed in order to add another buffer.



- Use the arrow keys to select the pH buffer to be included/removed in/from the buffer group.
- Press [Add] or [Remove] to add/remove the selected pH Buffer to/from buffer group.
- Press [Escape] to return to pH Setup menu.

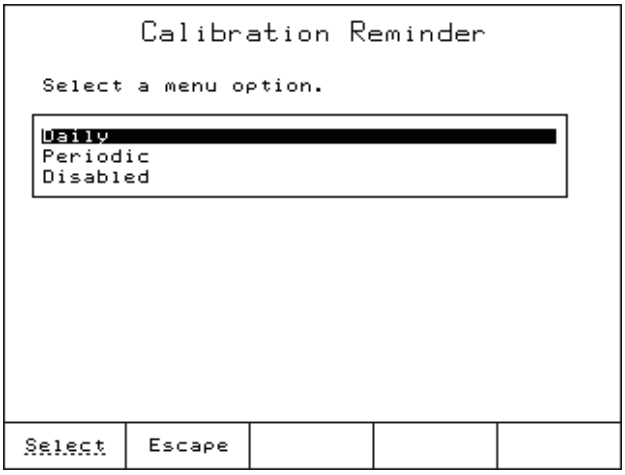
7.2.5 Calibration reminder

In order to have accurate readings, the electrode must be calibrated frequently. Three options are available for calibration reminder:

Daily - the calibration reminder will appear daily at specified time.

Periodic - the calibration reminder will appear after the set time has elapsed since the last calibration.

Disabled - the calibration reminder will not appear.

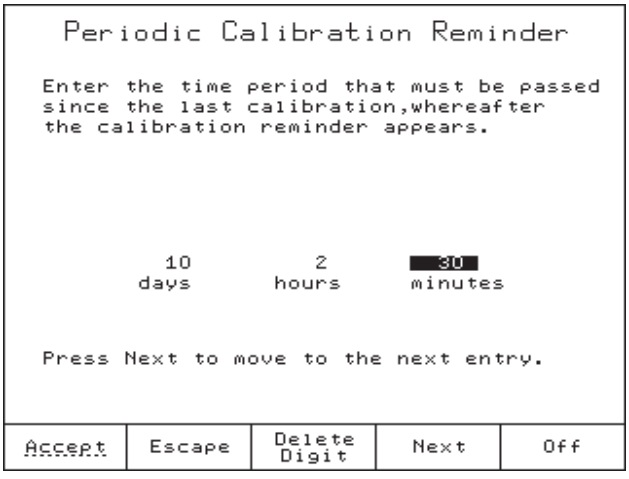


7.2.6 Set Reminder Period

If *Daily* or *Periodic* option was selected for the Calibration Reminder, the reminder period must also be set.

For a daily reminder period, the time of day can be set.

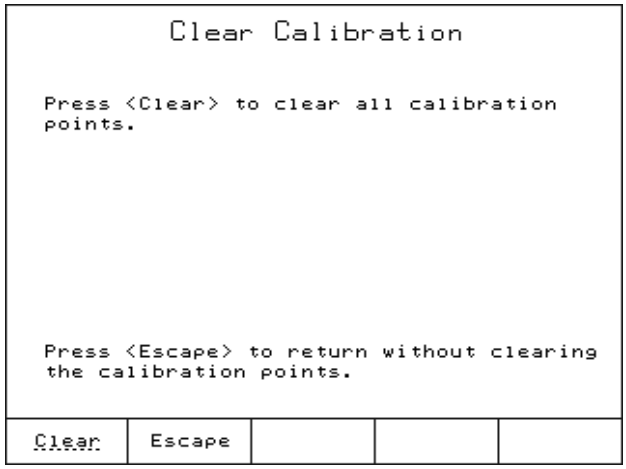
For a periodic reminder period, the number of days, hours and minutes can be set.



- Press [Next] to move the cursor to the next field.
- Press [Accept] to save the changes or [Escape] to return to the previous screen.
- Press [Off] to disable the calibration reminder and return to pH setup.

7.2.7 Clear Calibration

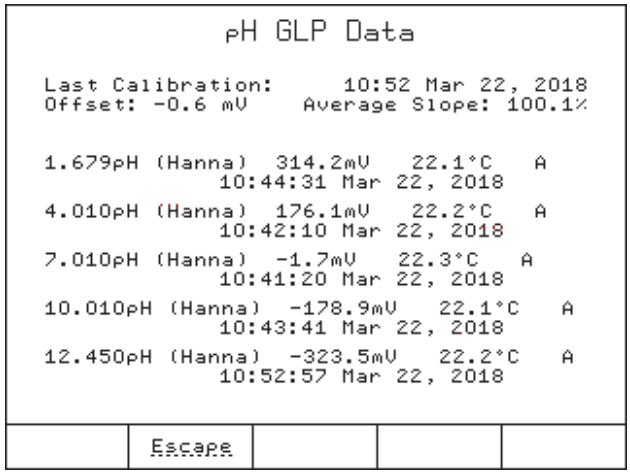
This option clears the existing pH calibration for the selected channel. If the calibration is cleared, another calibration has to be performed.



- Press [Clear] to clear the previous calibration or [Escape] to return to the previous screen without clearing the calibration.

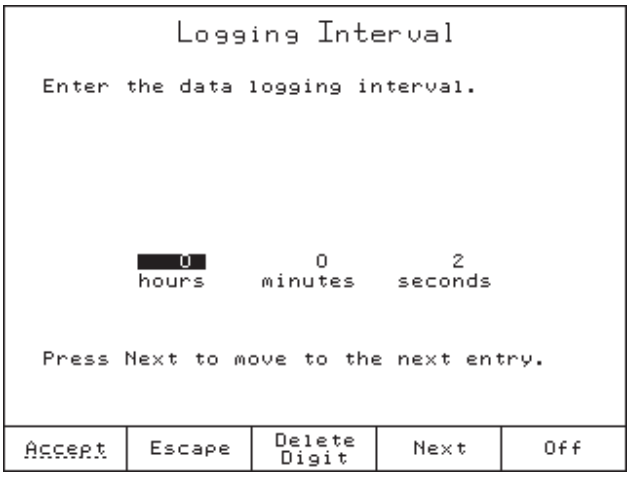
7.2.8 pH GLP Data

Displays the pH calibration data.



7.2.9 Logging Interval

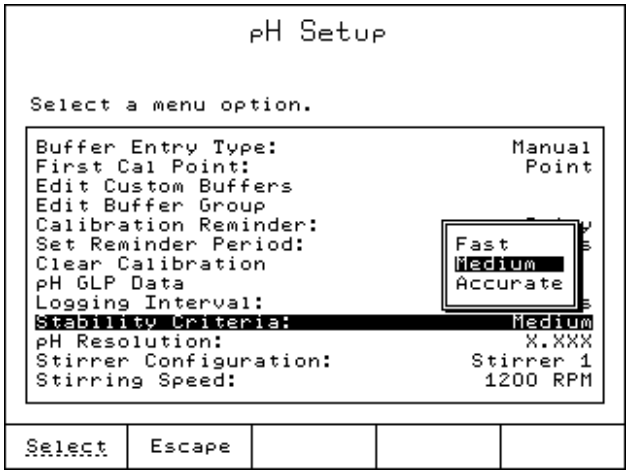
Set the logging interval to be used for automatic logging.



7.2.10 Stability Criteria

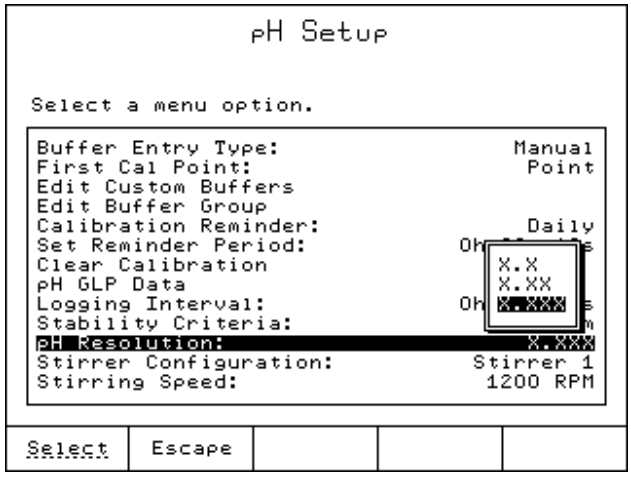
Select the signal stability criteria:

Fast - quicker results with less accuracy
Medium - medium speed results with medium accuracy
Accurate - slower results with high accuracy



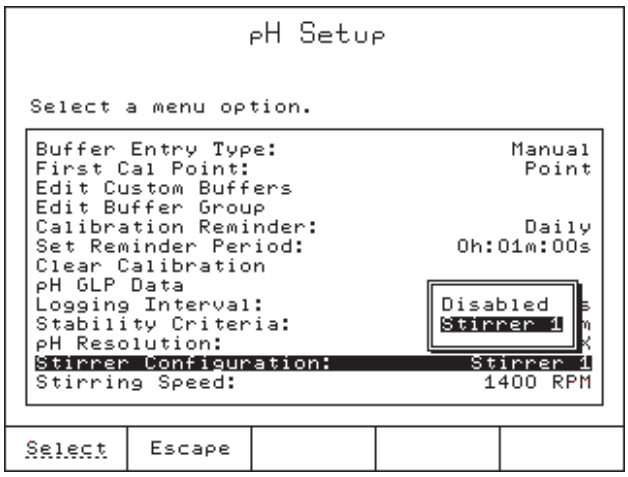
7.2.11 pH Resolution

Set the desired pH resolution: one (X.X), two (X.XX) or three (X.XXX) decimal places.



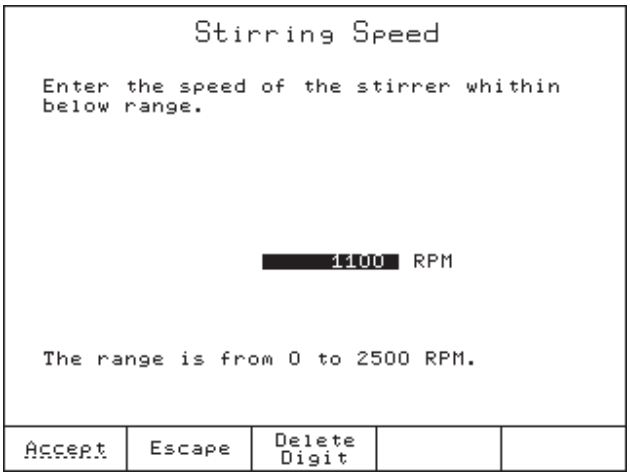
7.2.12 Stirrer Configuration

Set the stirrer configuration: Stirrer 1, Stirrer 2, or Disabled.



7.2.13 Stirring Speed

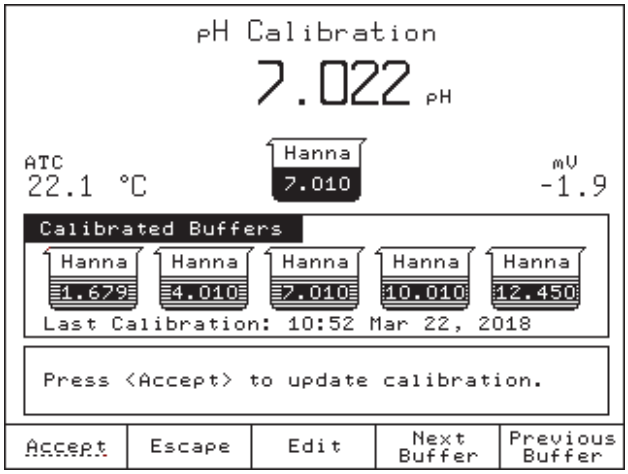
The stirring speed for the selected stirrer can be set.



7.3 pH Calibration

Calibrate the instrument often, especially if high accuracy is required.
The instrument should be recalibrated:

- Whenever the pH electrode is replaced.
- At least once a week.
- After testing aggressive chemicals.
- When "No pH Calibration" or "pH Calibration Expired" message appears on the LCD, in the Reminder messages area.



PREPARATION

Pour small quantities of the buffer solutions into clean beakers. If possible, use plastic beakers to minimize any EMC interferences.

For accurate calibration and to minimize cross-contamination, use two beakers for each buffer solution: one for rinsing the electrode and one for calibration. If you are measuring in the acidic range, use pH 7.01 or 6.86 as the first buffer and pH 4.01/3.00 or 1.68 as the second buffer. If you are measuring in the alkaline range, use pH 7.01 or 6.86 as the first buffer and pH 10.01/9.18 or 12.45 as the second buffer.

For extended range measurements (acidic and alkaline), perform a five-point calibration by selecting five buffers across the entire pH range.

CALIBRATION PROCEDURE

During calibration, the user has a choice of 8 standard buffers: pH 1.68, 3.00, 4.01, 6.86, 7.01, 9.18, 10.01, 12.45 and up to 5 custom buffers.

For accurate measurements it is recommended to perform a five-point calibration. However, at least a two-point calibration is suggested. For pH titrations, the selected buffers should bracket your end point (e.g.: if your end point value is at 8.5, use 7.01 and 9.18 for calibration).

Three buffer entry types are available: Automatic, Semiautomatic and Manual Selection (see *Buffer Entry Type* section).

To begin calibration:

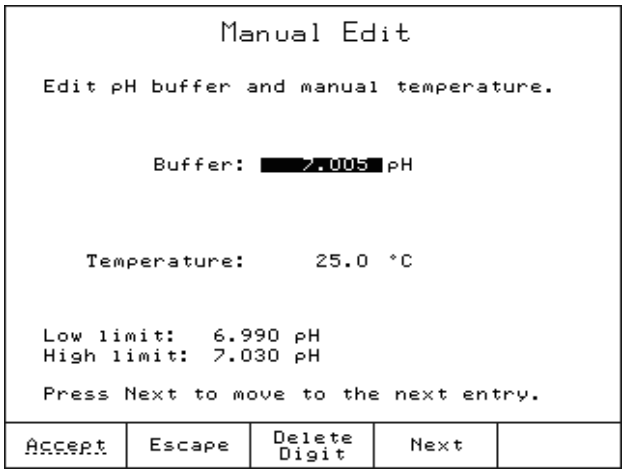
- Press pH Calibr.. If the instrument was calibrated before and the calibration was not cleared, the old calibration can be cleared by pressing Clear Cal.

Note: *It is very important to clear calibration history when a new electrode is used.*

- Immerse the pH electrode and the temperature probe approximately 4 cm (1.5") into a buffer solution and stir gently. The temperature probe should be close to the pH electrode.
- Select the pH calibration buffer value with Next Buffer or Previous Buffer.
- Press Accept to update the calibration. Once the reading has stabilized, the calibration buffer will be added to the Calibrated Buffers section.
- Rinse the pH electrode and the temperature probe, then immerse them into the next buffer solution and follow the above procedure or press Escape to exit the calibration.

Notes: • *The new calibration points will replace old ones if the difference between them is ± 0.2 pH. Buffers used in older calibrations will not have a solid background.*

- *If calibrating with a standard buffer in MTC mode, the pH value and temperature can be modified by pressing Edit. The values can be adjusted using the numeric keys. Press Accept to save the new values.*



- *In ATC mode, the pH value can be modified by pressing* Edit.
- *If the Automatic buffer entry type was selected for the calibration procedure, the instrument will automatically select the closest buffer to the measured pH value from the edit buffer group (see Buffer Group Edit section).*
- *If the Semiautomatic buffer entry type was selected for the calibration procedure, the instrument will automatically select the closest buffers to the measured pH value from all the available buffers and the buffer value can be selected with* Previous Buffer *or* Next Buffer.

CALIBRATION MESSAGES:

- **Wrong Buffer. Please check the buffer**: This message appears when the difference between the pH reading and the value of the selected calibration buffer is significant. If the message is displayed, check if you have selected the appropriate calibration buffer.
- **Wrong buffer temperature**: This message appears if the buffer temperature is out of the defined temperature range.
- **Clean the electrode or check the buffer. Press** Accept **to update calibration**: This message alerts the user that some dirt or deposits could be on the electrode.
- **Slope too low. Please check the buffer**: This message appears if the current slope is under 80% or over 110% of default slope. Recalibrate the instrument using fresh buffers.
- **Slope too high. Press** Clear Cal **to clear the old calibration**: This message appears as a result of an erroneous slope condition. Follow displayed instructions.

7.4 Logging

Data logging is available in pH mode. It can be logging on demand (Manual Logging) or automatically at predefined time intervals (Automatic Logging).
To customize the logging report:

- Press [results] to display the ***Data Parameters*** screen.
- Highlight the *Setup pH/mV/ISE Report* option and press [Select] to display the ***Setup pH/mV/ISE Report*** screen.
- Use the △ and ▽ keys to highlight the data field that you want to show/hide in the pH/mV/ISE report and then press [Select] to activate/deactivate it.
- Each field marked by "*" is an active field selected for the report.
- Press [Save Report] to save the customized report.

7.4.1 Automatic Logging

The logging interval is set in the pH / mV / ISE Setup screen.
Press [start/stop] to start the log.
The logging interval and name of logging file will be also displayed on the measure screen.
To stop the automatic logging, press [start/stop] again.

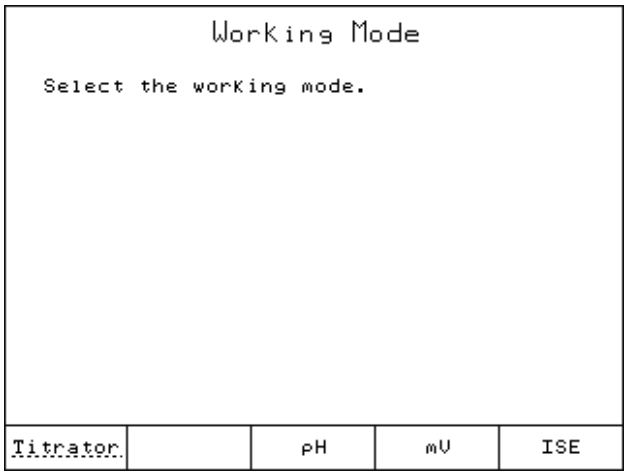
7.4.2 Manual Logging

To manually log pH readings, press [Save Reading] from the ***pH*** screen.
A new record will be added to the report every time [Save Reading] is pressed.

Chapter 8. Contents

8 mV

By pressing [Mode] from the main screen, the Titrator can be switched to **Titrator**, **pH**, **mV** or **ISE** modes.



[Titrator] Switches to **Titrator** mode.

[pH] Switches to **pH** mode.

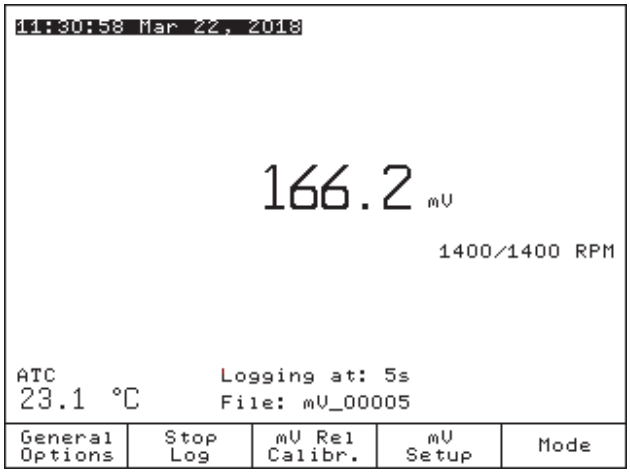
[mV] Switches to **mV** mode.

[ISE] Switches to **ISE** mode.

mV MODE

8.1 Display

The ***mV*** screen is shown below.



mV Mode Option Keys:

General Option — The General Options screen gives you access to options that are not directly related to the measurement process (See **General Options** chapter for more information).

Save Reading — Stores the current mV reading (see *Manual Logging* section).

or

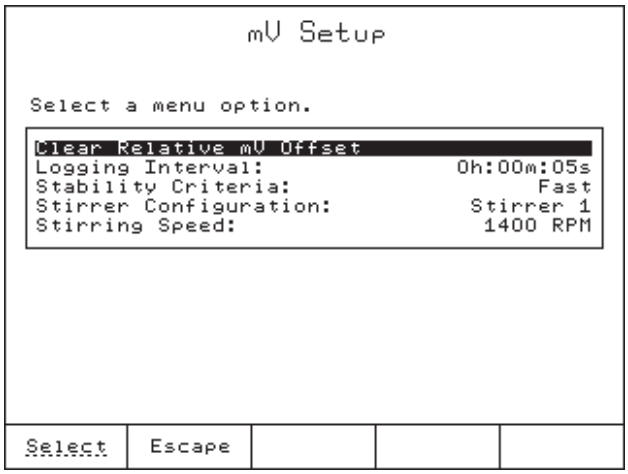
Start Log — Starts the mV automatic log (see *Automatic Logging* section).

mV Rel Calibr. — Enter the relative mV calibration screen (see *Relative mV Calibration* section).

mV Setup — Enter the mV setup screen. Parameters are associated with mV measurement and calibration.

Mode — Allows the user to switch between the available measurement modes: Titrator, pH, mV or ISE mode.

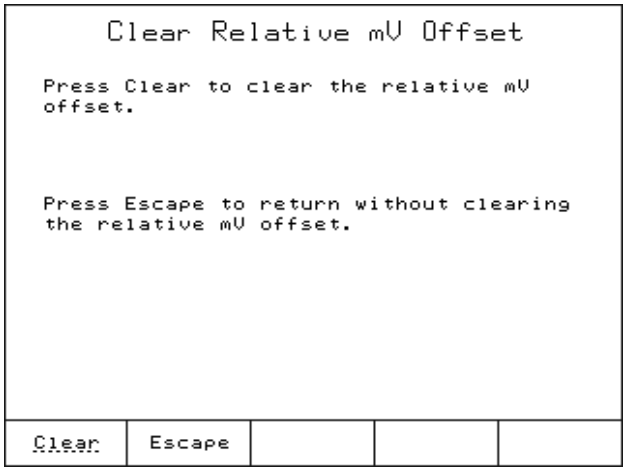
8.2 mV Setup



8.2.1 Clear Relative mV Offset

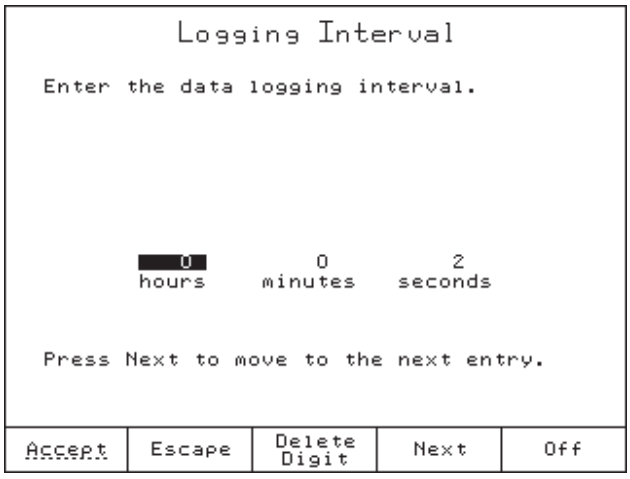
Clear the relative mV offset and return to absolute mV measurement.

- Press Clear to clear the relative mV offset or Escape to return to the previous screen.



8.2.2 Logging Interval

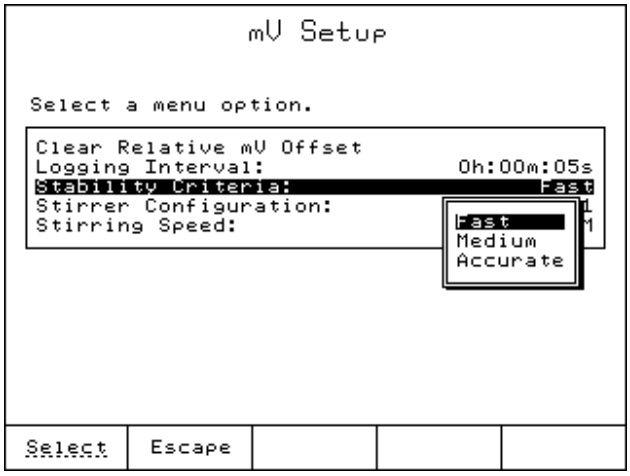
Set the logging interval.



8.2.3 Stability Criteria

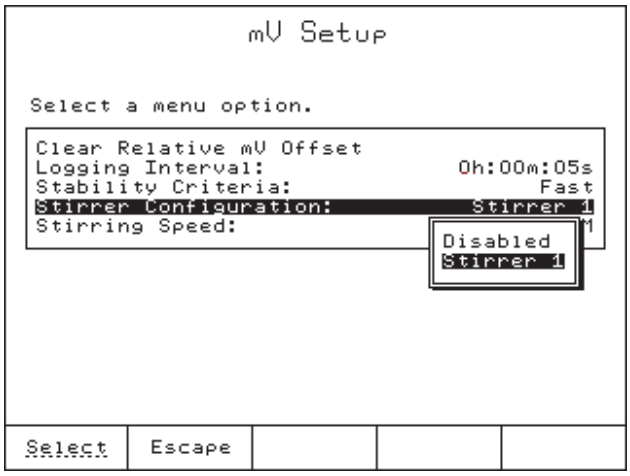
Select the signal stability criteria:

Fast - quicker results with less accuracy
Medium - medium speed results with medium accuracy
Accurate - slower results with high accuracy



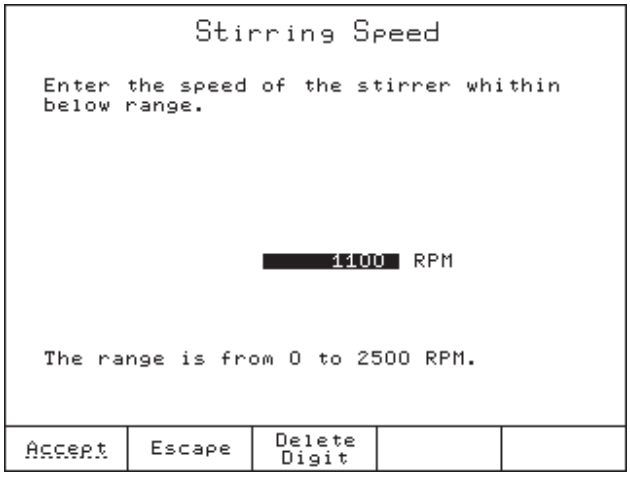
8.2.4 Stirrer Configuration

Set the stirrer configuration: Stirrer 1, Stirrer 2 or Disabled.

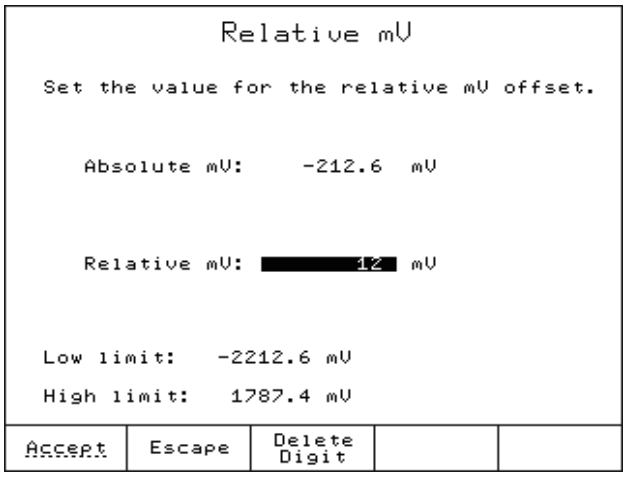


8.2.5 Stirring Speed

The stirring speed for the selected stirrer can be set.



8.3 Relative mV Calibration



- Press [Accept] to accept the value.
- Press [Delete Digit] to delete the last digit.
- Press [Escape] to cancel this operation and return to the previous screen.

8.4 Logging

Data logging is available in mV mode. It can be logging on demand (Manual Logging) or automatically at predefined time intervals (Automatic Logging).
To customize the logging report:

- Press [results] to display the ***Data Parameters*** screen.
- Highlight the *Setup pH/mV/ISE Report* option and press [Select] to display the ***Setup pH/mV/ISE Report*** screen.
- Use the △ and ▽ keys to highlight the data field that you want to show/hide in the pH/mV/ISE report and then press [Select] to activate/deactivate it.
- Each field marked by "*" is an active field selected for the report.
- Press [Save Report] to save the customized report.

8.4.1 Automatic Logging

The logging interval is set in the mV Setup screen.
Press [start/stop] to start the log.
The logging interval and name of logging file will be also displayed on the measure screen.
To stop the automatic logging, press [start/stop] again.

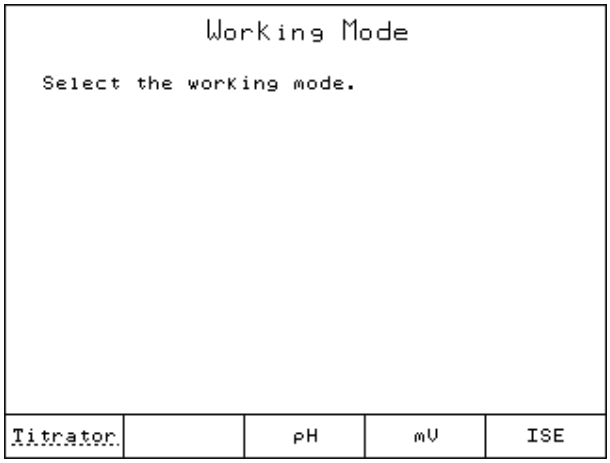
8.4.2 Manual Logging

To manually log mV readings, press [Save Reading] from the ***mV*** screen.
A new record will be added to the report every time [Save Reading] is pressed.

Chapter 9. Contents

9 ISE Mode

By pressing [Mode] from the main screen, the Titrator can be switched to **Titrator**, **pH**, **mV** or **ISE** modes.



[Titrator] Switches to **Titrator** mode.

[pH] Switches to **pH** mode.

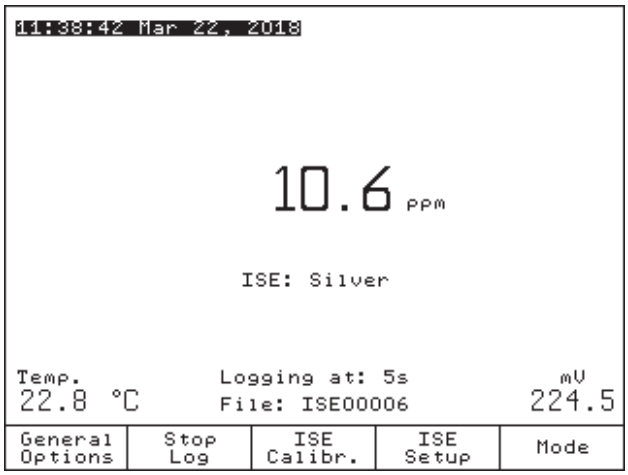
[mV] Switches to **mV** mode.

[ISE] Switches to **ISE** mode.

ISE MODE

9.1 Display

The ***ISE*** screen is shown below.



ISE Mode option keys:

General Options — The General Options screen gives access to options that are not directly related to the measurement process (see **General Options** chapter for more information).

Save Reading — Stores the current concentration reading (see *Manual Logging* section*)*.

or

Start Log — Starts the ISE automatic log (see *Automatic Logging* section).

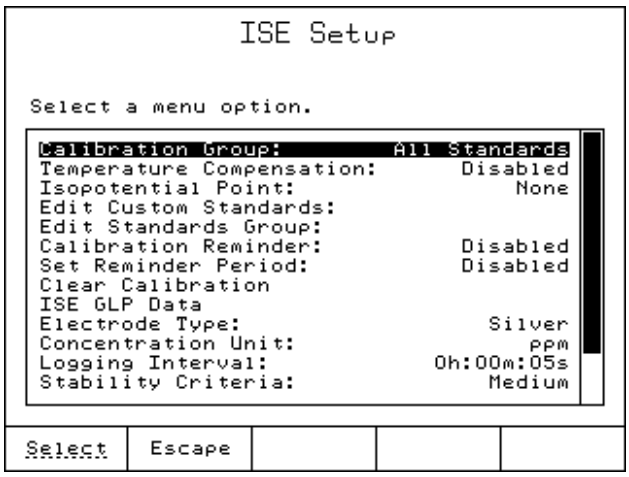
ISE Calibr. — Enter the ISE calibration screen (see *ISE Calibration* section).

ISE Setup — Enter the ISE setup screen. Parameters are associated with ISE measurements and calibration.

Mode — Allows the user to switch between the available measurement modes: Titrator, pH, mV and ISE mode.

9.2 ISE Setup

To access the ISE Setup, press ISE Setup option key in ISE mode.

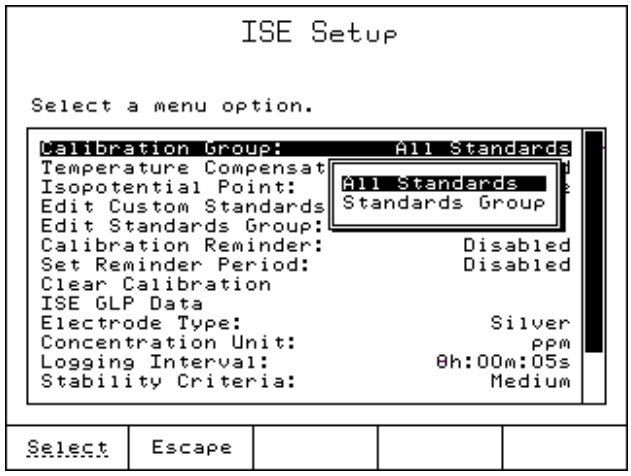


9.2.1 Calibration Group

Selecting the set of available standards to be used in calibration:

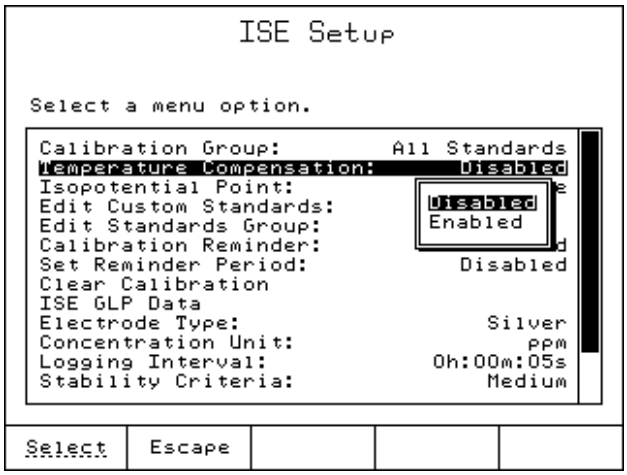
All Standards: the set of available standards includes the Standard solutions and Custom solutions.

Standards Group: the set of available standards includes the standards selected by the user.



9.2.2 Temperature Compensation

Enable or disable temperature compensation for ISE measurements.

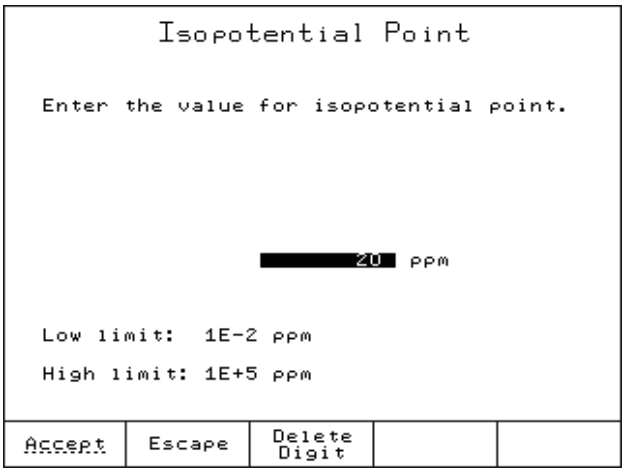


Note: *If you enabled Temperature Compensation, then the isopotential point must be set.*

9.2.3 Isopotential Point

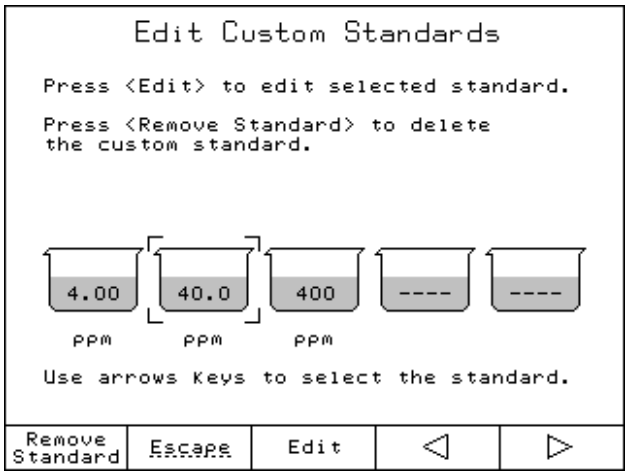
This option is available only if temperature compensation is enabled.

This option allows the user to set an isopotential point for the selected electrode. Ion selective electrodes have different isopotential points. The isopotential point is edited in ppm units only. The isopotential point should be entered if it is known and if measurements are going to be made at several temperatures.



9.2.4 Edit Custom Standards

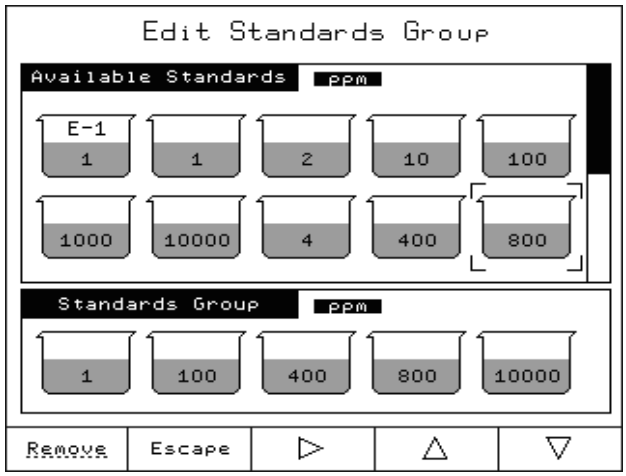
Edit the custom standard list. Up to five can be used in calibration.



- Use the ◁ and ▷ keys to select the standard.
- Press [Remove Standard] to delete the custom standard.
- Press [Edit] to edit the selected custom standard; use the numeric keys to edit the standard.

9.2.5 Edit Standard Group

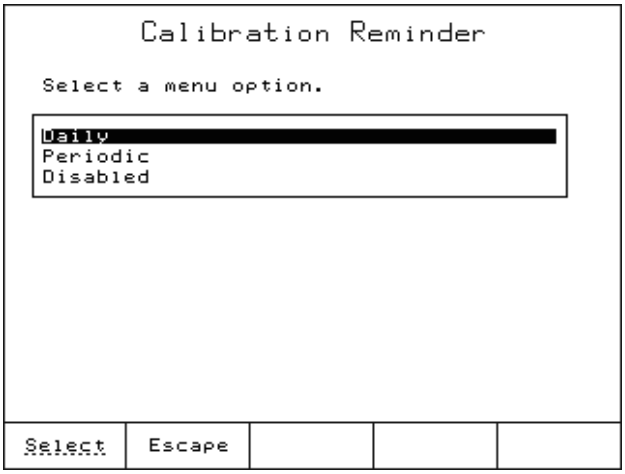
Select up to 5 standards from the available standards (Predefined and Custom) to be used during calibration.



- Use the arrow keys to select the standard to be included/removed in/from the standard group.
- Press [Add] or [Remove] to add/remove the selected standard to/from standard Group.
- Press [Escape] to return to ISE Setup menu.

9.2.6 Calibration Reminder

In order to have accurate readings, the electrode must be calibrated frequently. Three options are available for the calibration reminder:

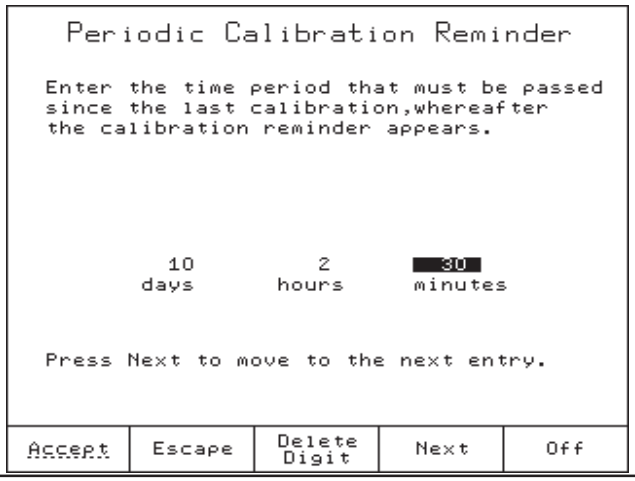


Daily - the calibration reminder will appear daily at specified time.
Periodic - the calibration reminder will appear after the set time has elapsed since the last calibration.
Disable - the calibration reminder will not appear.

9.2.7 Set Reminder Period

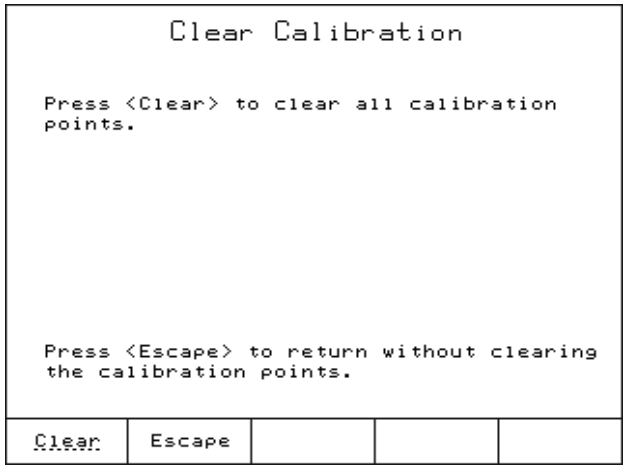
If Daily or Periodic option was selected for the Calibration Reminder, the reminder period must also be set.
For a daily reminder period the time of day can be set.
For a periodic reminder period the number of days, hours and minutes can be set.

- Press [Next] to move the cursor to the next field.
- Press [Accept] to save the changes or [Escape] to return to the previous screen.
- Press [Off] to disable the calibration reminder and return to ISE setup menu.



9.2.8 Clear Calibration

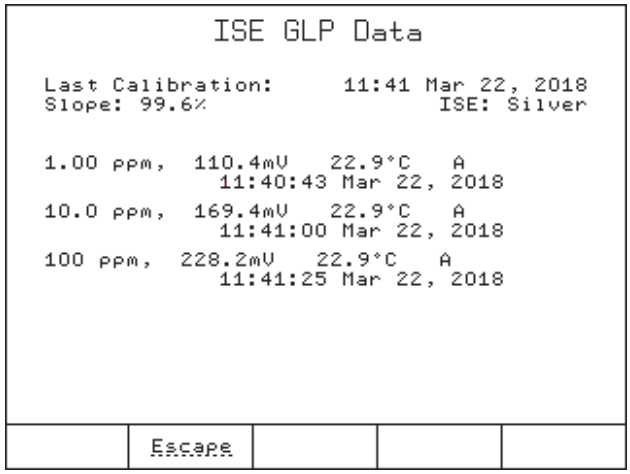
This option clears the existing ISE calibration. If the calibration is cleared, a new calibration must be done in order to take measurements.



- Press [Clear] to clear the previous calibration or [Escape] to return to the previous screen.

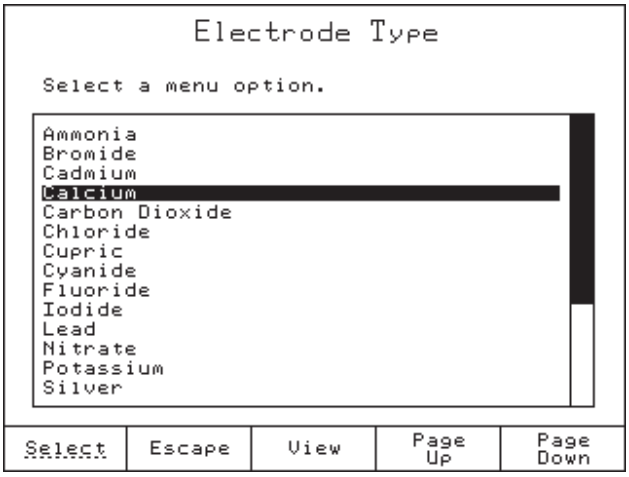
9.2.9 ISE GLP Data

Displays the ISE calibration data.



9.2.10 Electrode Type

Select the Ion Selective Electrode used for measurements from a list: Ammonia, Bromide, Cadmium, Calcium, Carbon Dioxide, Chloride, Cupric, Cyanide, Fluoride, Iodide, Lead, Nitrate, Potassium, Silver, Sodium, Sulfate, Sulfide or five custom ISE. For the standard ISE, it is possible to view the Ion constants (Name, Molar Weight and Electric Charge/Slope), while for the custom ISE, all of these constants must be manually set.



For Standard ISE:

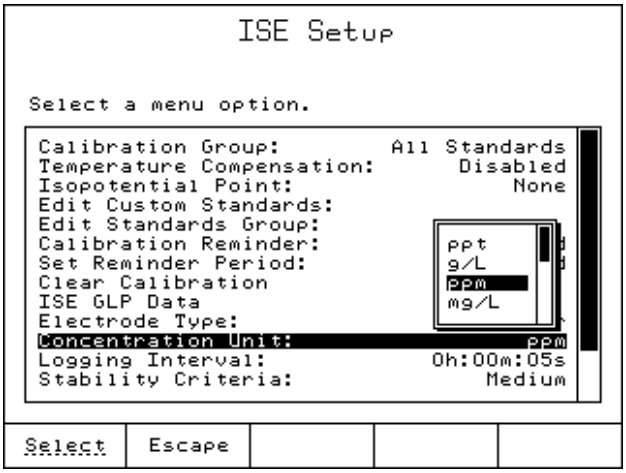
- Press [View] to see the Ion constants, press [Escape] at any time to exit Ion constants view.

For Custom ISE:

- Press [View] to edit the Ion constants for the selected custom ISE. Use the △ and ▽ keys to select the desired Ion constant and press [Select] to edit the value or [Escape] to cancel operation.
- Set the Ion Name (up to 10 characters can be entered).
- Set the appropriate molecular weight (in g / mol) using the numeric keys. Press [Accept] to save the value or press [Escape] to return to the previous screen.
- Select the appropriate Electric Charge / Slope. Use the △ and ▽ keys to select the value and then press [Select]. If the Ion electric charge is None, its slope can be manually set by pressing [Edit]. Press [Accept] to save the value or press [Escape] to return to the previous screen.

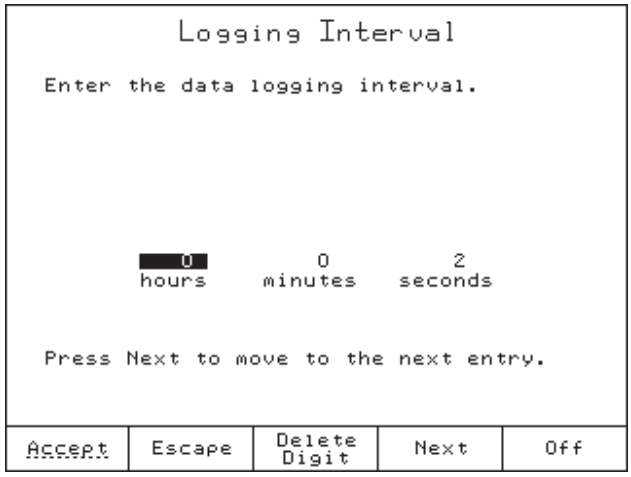
9.2.11 Concentration Unit

Select the desired concentration unit for the measured Ion or chemical compound. The available concentration units are: ppt (g/L), ppm (mg/L), ppb (µg/L), mg/mL, M (mol/L), mmol/L, %w/v or user defined.



9.2.12 Logging Interval

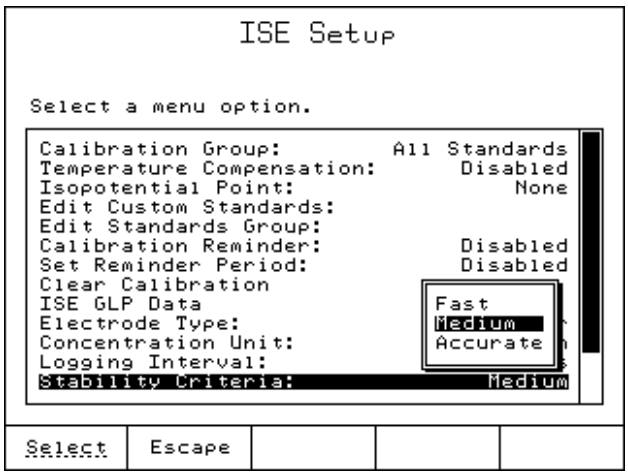
Set the logging interval to be used.



9.2.13 Stability Criteria

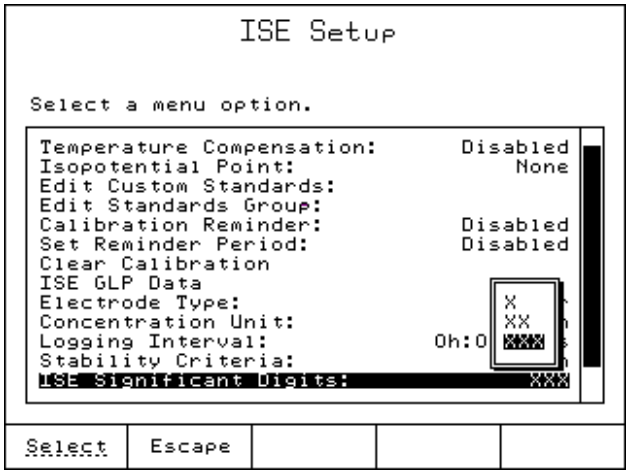
This option allows the user to select the signal stability criteria for the measured parameters:

Fast - quicker results with less accuracy
Medium - medium speed results with medium accuracy
Accurate - slower results with high accuracy



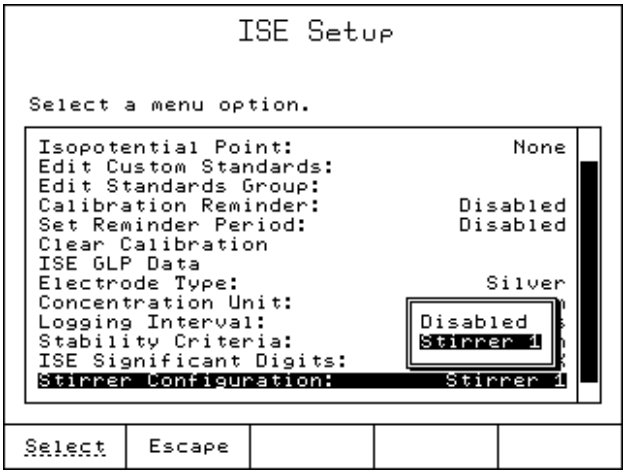
9.2.14 ISE Significant Digits

Select the number of significant digits to be displayed: one (X), two(XX) or three(XXX).



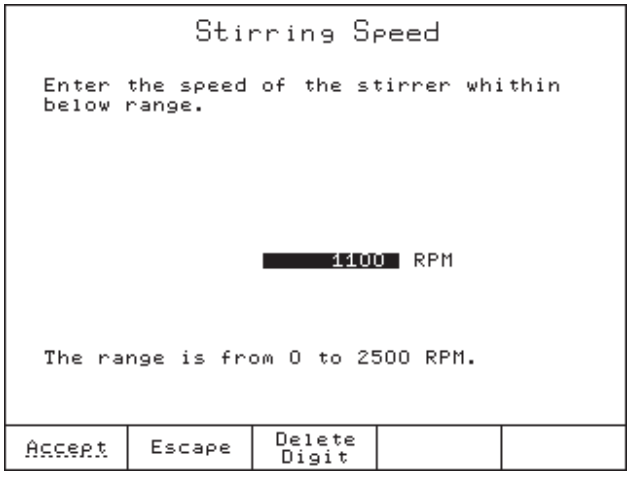
9.2.15 Stirrer Configuration

Set the stirrer configuration: Stirrer 1, Stirrer 2 (when available) or Disabled.



9.2.16 Stirring Speed

The stirring speed for the selected stirrer can be set.



9.3 ISE Calibration

It is recommended to calibrate the instruments frequently if high accuracy is required. The instrument should also be recalibrated whenever the "Calibrate Electrode" message appears on the LCD.

Due to electrode conditioning time, the electrode must be immersed for several seconds to stabilize. The user will be guided step by step during calibration with easy-to-follow messages on the display. This will make the calibration a simple and error-free procedure.

PREPARATION:
Pour small quantities of the standard solution into clean beakers. If possible, use plastic beakers to minimize any EMC interferences.

ISE MODE

For accurate calibration and to minimize cross-contamination, use two beakers for each standard solution: one for rinsing the electrode and one for calibration.

Note: *For accurate measurements, add the appropriate ISA (Ionic Strength Adjustment) to the calibration standards.*

CALIBRATION PROCEDURE:

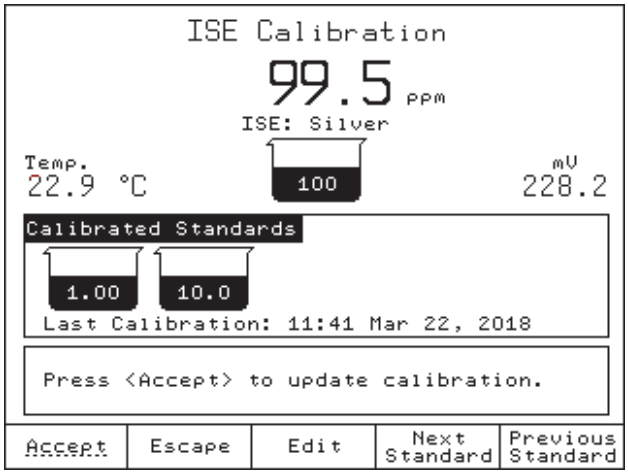
Before calibrating, make sure that the appropriate Electrode Type and concentration unit has been selected in ISE Setup.

Up to a five points calibration is possible using any combination of five memorized standard solutions and five custom solutions.

The ISE calibration and measurement can be performed with or without temperature compensation. If the temperature compensation option is enabled, the isopotential point of the electrode must be set in ISE Setup.

The current standard will be manually selected by the user from the available standards list.

The list of available standards depends of the Manual Entry setting.



To calibrate the instrument using Manual Entry:

- Press [ISE Calibr.] from the main screen. If the instrument was calibrated before and the calibration was not cleared, the old calibration can be cleared by pressing [Clear Cal].
- Immerse the Ion Selective Electrode and the temperature probe approximately 2 cm into the lowest concentrated standard solution.
- Select the concentration with [Next Standard] or [Previous Standard].
- When the reading has stabilized, press [Accept] to update the calibration. The calibration point value will be added to the Calibrated Standard list.
- Select [Next Standard] and repeat the procedure with all of the available standards.

9.4 Logging

Data logging is available in ISE mode. It can be logging on demand (Manual Logging) or automatically at predefined time intervals (Automatic Logging).
To customize the logging report:

- Press [results] to display the ***Data Parameters*** screen.
- Highlight the *Setup pH/mV/ISE Report* option and press [Select] to display the ***Setup pH/mV/ISE Report*** screen.
- Use the △ and ▽ keys to highlight the data field that you want to show/hide in the pH/mV/ISE report and then press [Select] to activate/deactivate it.
- Each field marked by "*" is an active field selected for the report.
- Press [Save Report] to save the customized report.

9.4.1 Automatic Logging

The logging interval is set in the ISE Setup screen.
Press [start/stop] to start the log.
The logging interval and name of logging file will be also displayed on the measure screen.
To stop the automatic logging, press [start/stop] again.

9.4.2 Manual Logging

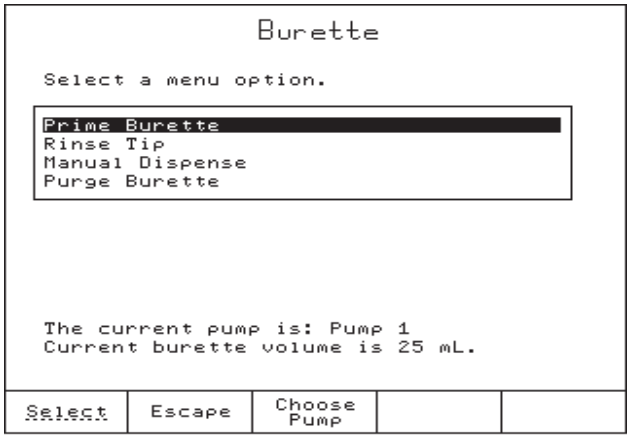
To manually log ISE readings, press [Save Reading] from the ***ISE*** screen.
A new record will be added to the report every time [Save Reading] is pressed.

Chapter 10. Contents

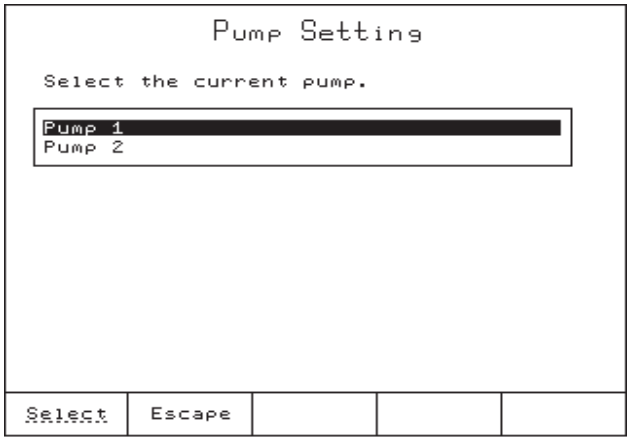
10 AUXILIARY FUNCTIONS

10.1 Burette

To access the ***Burette*** screen, press [Burette] from the main titration screen.
Highlight the desired option and then press [Select].

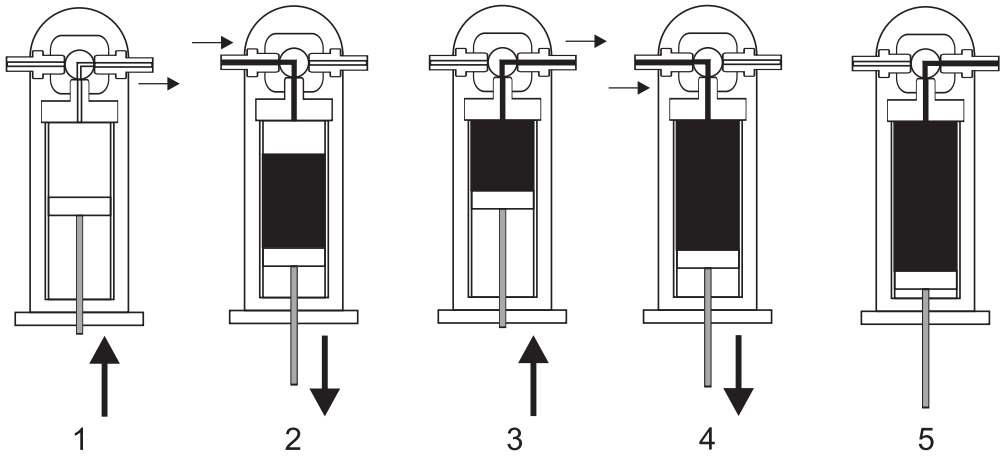


[Choose Pump] allows you to select the desired pump for burette operations (it is only active if two pumps are connected).



10.1.1 Prime Burette

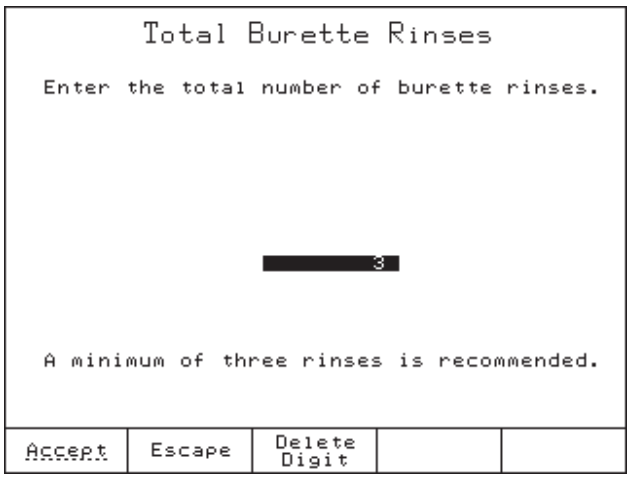
The *Prime Burette* option is used to mechanically fill the burette before starting a set of titrations. The priming process consists of several cycles of filling and emptying the burette with titrant.
Two rinse cycles of burette are shown in the figure below. The dispensing tube is connected on the right side and the aspiration tube on the left side.



Note: *Before starting this operation, the aspiration tube must be inserted in the titrant bottle. A waste container should be placed under the dispensing tip to collect the waste solution.*

To prime the burette, select *Prime Burette* from the ***Burette*** screen. Enter the number of rinses and press Accept.

The number of burette rinses can be set between 1 and 5 (we recommend at least three rinses to assure that the air bubbles are completely removed).

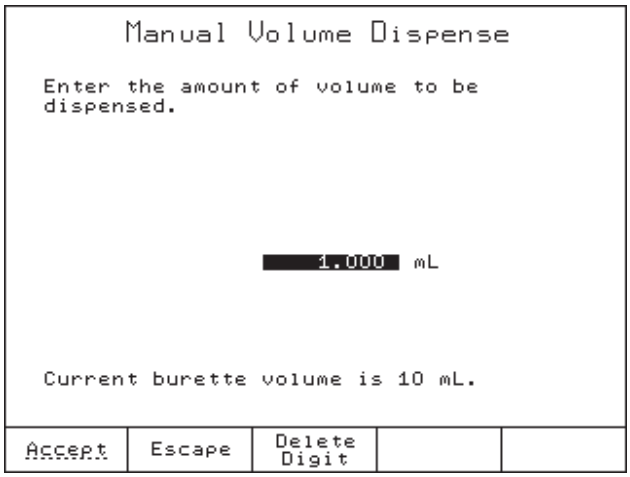


10.1.2 Rinse Tip

A 2 mL dose of titrant will be dispensed from the burette when this operation is selected. This operation will eliminate the air from the dispensing tip.

10.1.3 Manual Dispense

Manual Dispense option allows a defined titrant volume to be dosed. Select the *Manual Dispense* option and press Select. The ***Manual Volume Dispense*** screen will become active and the display will prompt you to enter the desired volume to be dispensed.



The manual dispense volume must be between the limits shown below:

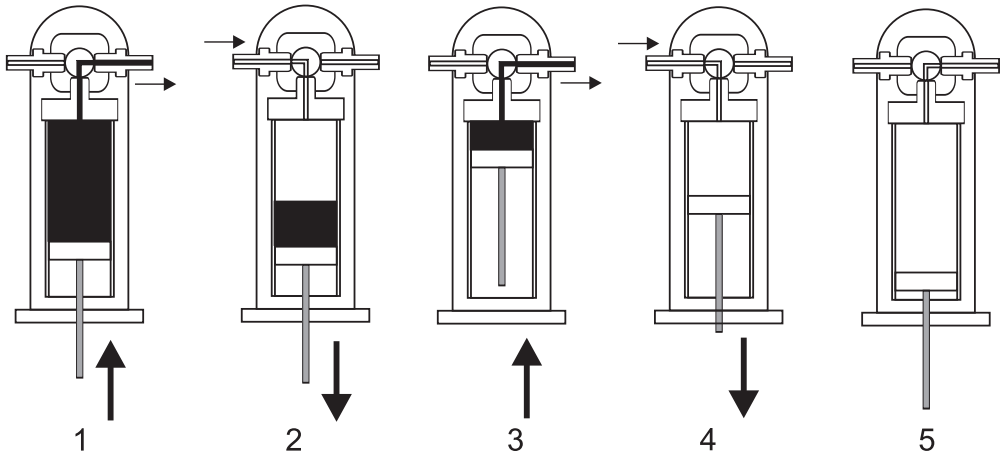
0.001 to 4.750 mL for a 5 mL burette
0.001 to 9.500 mL for a 10 mL burette
0.005 to 23.750 mL for a 25 mL burette
0.005 to 47.500 mL for a 50 mL burette

10.1.4 Purge Burette

This option allows the burette to be emptied before cleaning and/or storing the burette. The burette is flushed twice.

Note: *Before starting this operation, remove the aspiration tube from the titrant bottle.*

The figures below show the steps in a purge burette operation.

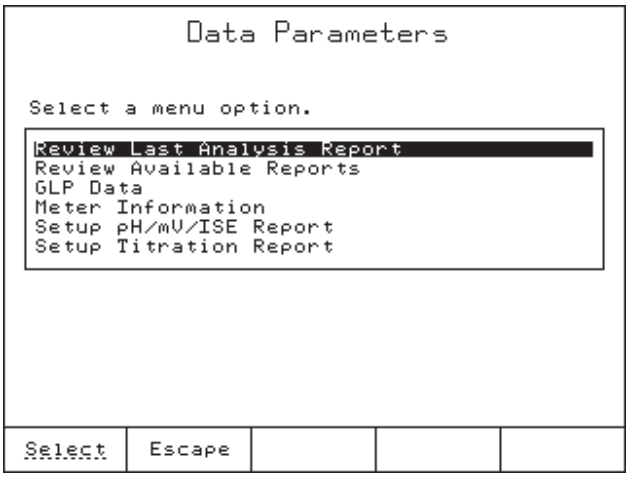


10.2 Stirrer

The stirrer can be turned on and off by pressing [stir].
The stirring speed is set within the method parameters (see **Titration Methods**, *Stirring Speed* section).
During the titration process, the stirring speed can be manually adjusted by using the △ and ▽ keys.

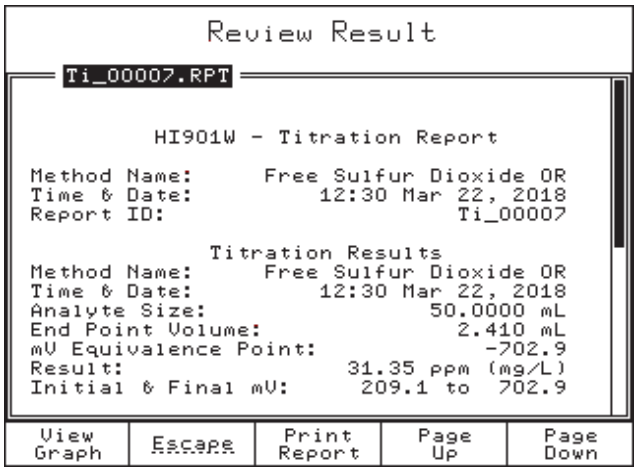
10.3 Results

From the ***Data Parameters*** screen, you can access the following options:



10.3.1 Review Last Analysis Report

The last analysis report can be reviewed.
The titration graph can be reviewed by selecting [View Graph].



The information seen in the report is based on the selections made in the ***Setup Titration Report*** screen.
The following option keys are available:

[View Graph] Review the titration graph. The potentiometric titration curve is displayed. If the *Equivalence End Point* option was selected, the derivative curve (1st derivative, 2nd derivative) is simultaneously displayed. Pressing [Select Trace] will change the vertical axes scale units.

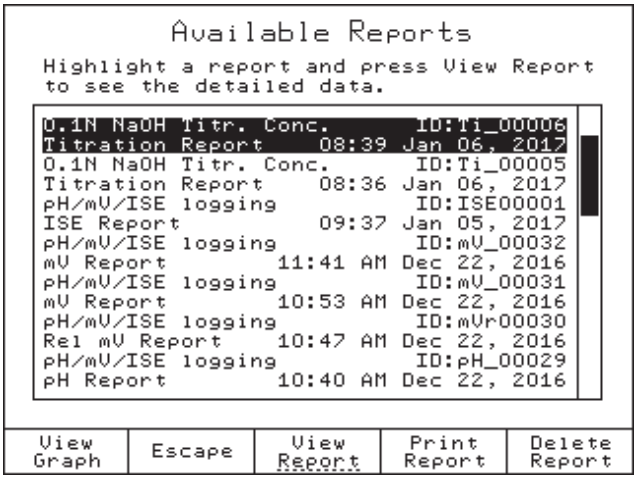
[Print Report] Print the titration report.

[Escape] Return to the previous screen.

[Page Up] [Page Down] Keys can be used to scroll through the pages.

10.3.2 Review Available Reports

Up to 100 reports can be saved on the Titrator. To view one of the saved reports, highlight a report and then press [View Report].
All of the saved reports can be reviewed and printed.



The report contains only the information selected in the ***Setup Titration Report*** and ***Setup pH/mV/ISE Report*** screens during report configuration.

The following option keys are available:

[View Graph] Review the selected graph.

[View Report] Review the selected report.

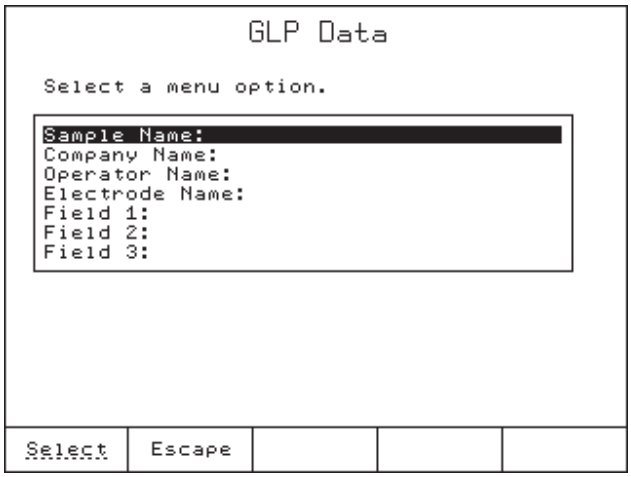
[Print Report] Print the selected report.

[Delete Report] Delete the selected report.

[Escape] Return to the previous screen.

10.3.3 GLP Data

Enter up to 20 alphanumeric characters for each option from ***GLP Data*** screen.

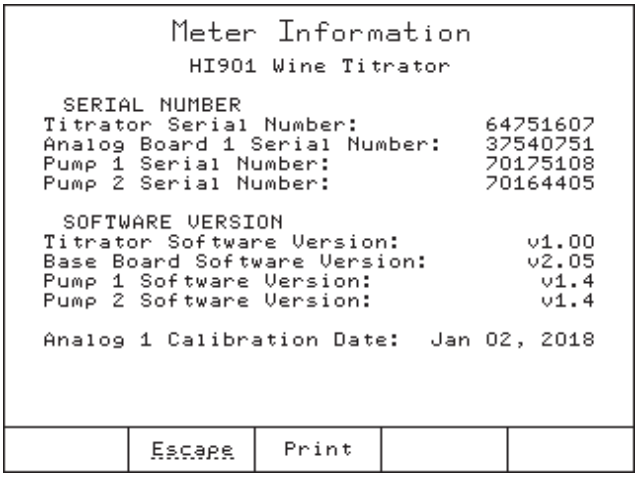


Sample Name	Allows the sample name to be recorded in each report. The sample name will increase by one, with each new titration or logging report, if the last character is a number.
Company Name	Allows the company name to be recorded in each report.
Operator Name	Allows the operator name to be recorded in each report.
Electrode Name	Allows the electrode name to be recorded in each report.
Fields 1, 2, 3	Allows any additional information to be recorded in each report.

The fields must be selected from ***Setup Titration Report*** screen (see *Setup pH/mV/ISE* section and *Setup Titration Report* section) in order to be displayed in the titration report.

10.3.4 Meter Information

Displays titrator configuration data.



Titrator Serial Number: The serial number of the Titrator base board.

Analog Board 1 Serial Number: The serial number of the analog board.

Pump 1 (and/or 2) Serial Number: The serial number of the connected pump.

Titrator Software Version: The current software version installed on the Titrator.

Base Board Software Version: The current software version present on the base board of the Titrator.

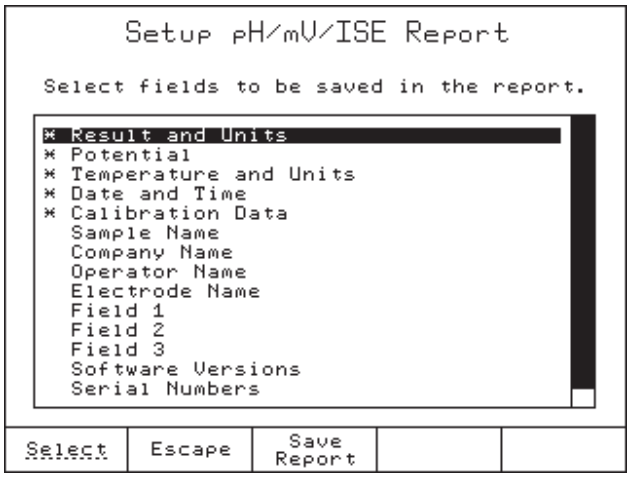
Pump 1 (and/or 2) Software Version: The current software version for the pump.

Analog 1 (and/or 2) Calibration Date: Manufacturer calibration date of the analog board.

Note: *If more than 1 year elapsed from the calibration date of the analog board 1 and/or 2, the message* **Analog 1 Calibration Due** *will appear on the main screen. The analog board needs to be recalibrated.*

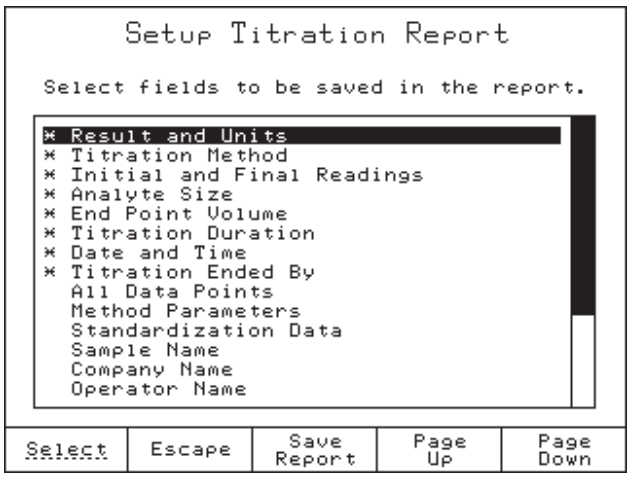
10.3.5 Setup pH/mV/ISE Report

Customize a unique report to record the pH, mV, and ISE measurements. An asterisk means that it will be included in the report.



10.3.6 Setup Titration Report

Customize a unique report to record the titration results. An asterisk means that it will be included in the titration report.



Chapter 11. Contents

11 MAINTENANCE, PERIPHERALS

The 25-mL burette included with the Titrator exceeds the ISO 8655 standard for accurate delivery of liquids by a motor-driven piston burette.

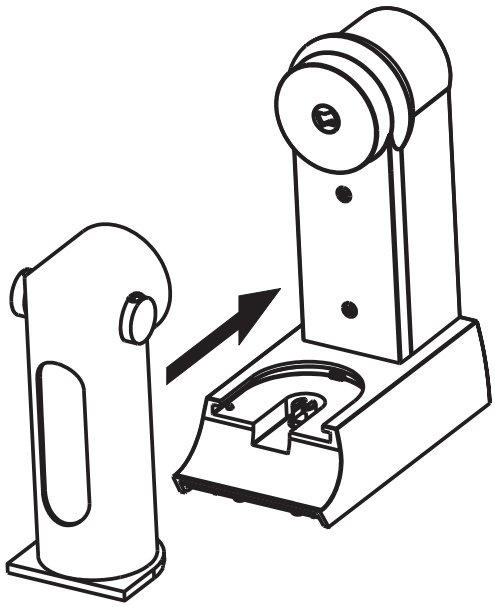
11.1 Burette Maintenance

11.1.1 Burette Assembly

The burette is delivered with a 25-mL syringe inside and with all of the accessories mounted (see **Setup**, *Unpacking* section for burette assembly details). The burette assembly consists of a rigid housing which holds the glass syringe, a 3-way valve and titrant tubing.

11.1.2 Changing the Burette

Remove the burette from the pump assembly by sliding it forward and then slide the new burette into place (see the picture below).

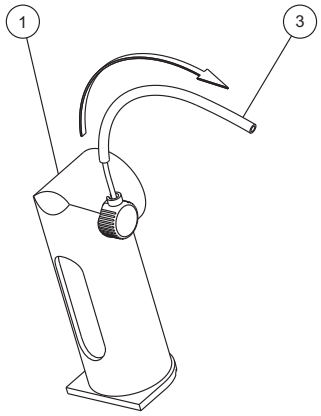


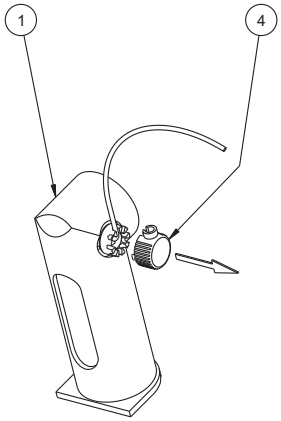
11.1.3 Disassembling the Burette

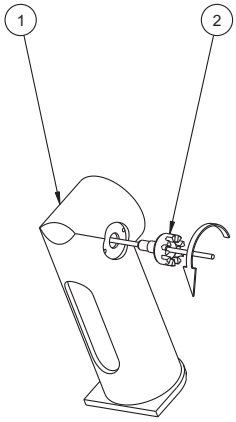
The aspiration and the dispensing tubes have fittings and tube protectors. The aspiration tube will be mounted in the left side and the dispensing tube will be mounted in the right side of the burette.

To remove the dispensing tube and the aspiration tube follow these steps:

- Slide up the tube protector (3).
- Remove the tube lock (4) from the burette holder.
- Unscrew the fitting (2).
- Remove the tube.



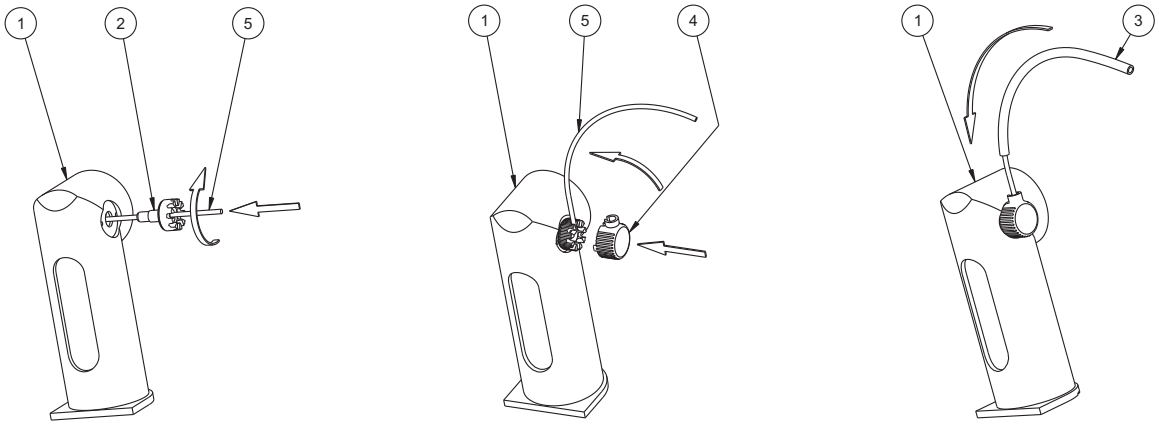




11.1.4 Assembling the Burette

To attach the dispensing tube and the aspiration tube, follow these steps:

- Insert the flat-shaped end of the dispensing tube into the valve outlet and screw in the fitting so that the highest of its 9 cuts stays vertically in the final position (2).
- Bend the tube up into the vertical position to enter the highest cut of the fitting (5).
- Put on the tube lock on the fitting (4).
- Slide down tightly the tube protector (3) into the dedicated gap of the tube lock.



11.1.5 Cleaning the Burette

To clean the burette, follow these steps:

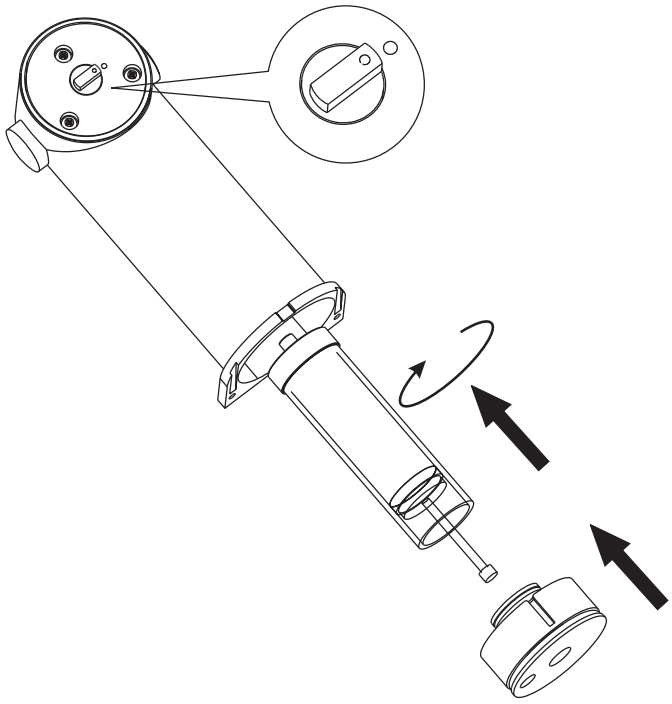
- If the burette is filled with titrant, remove the aspiration tube from the titrant bottle and purge burette (see **Auxiliary Functions**, *Purge Burette* section).
- Insert the aspiration tube into cleaning solution, deionized water or titrant solvent.
- Prime burette to fill the burette (use 2 rinses) (see **Auxiliary Functions**, *Prime Burette* section).
- During second refilling of the burette remove the aspiration tube out of the cleaning solution, deionized water, or solvent and allow the air to replace the liquid in the burette. This will clean the aspiration tube.

If this simple cleaning procedure is not adequate, continue with these steps:

- Slide the burette out from the pump assembly.
- Remove the dispensing and aspiration tubes. Clean them separately or insert new ones.
- Remove the protective cap from the bottom of the burette assembly by using the special tool.
- Remove the syringe from the burette assembly by unscrewing it with your fingers.
- Extract the piston from the syringe.
- Clean both the piston and the syringe with appropriate cleaning solution. Rinse with deionized water.
- Remove the excess liquid.

Warning: *Avoid contacting the titrant with bare hands.*
Avoid spilling titrant.
Clean the external side of the syringe and piston to remove aggressive chemicals.
Do not touch the white PTFE part of the piston or internal walls of the burette with bare hands or greasy materials.

- Reinsert the piston into the syringe.
- Reinsert the syringe by screwing it in the valve with your fingers.
- Reinsert the protective cap to the bottom of the burette assembly. Carefully position the cap into the burette.
- Slide the burette into the burette stand. Notice the position of the piston shaft to the pump couple.
- Priming the burette three times with new titrant is recommended.



11.1.6 Burette Preparation (Titrant Filling)

Before starting a titration, the burette must be properly filled with titrant in order to obtain an accurate and repeatable result. To fill the burette, follow the next steps and recommendations:

- If necessary, clean the burette and make sure it is empty.
- From the main screen press Burette.
- Highlight *Prime Burette* option and press Select.
- Enter the number of times the burette needs to be rinsed (minimum three rinses are recommend allowing air bubbles to be evacuated).
- Press Accept.

To avoid the presence of the air bubbles inside the burette, make sure to have a continuous liquid flow inside the burette. A little air just above the liquid level at the first filling is normal. The next filling will evacuate all of the air; no air will be left in the valve.
Sometimes during this process, slight finger tapping on the tubes is helpful to remove any residual air bubbles from the tubes.
If air bubbles are still present:

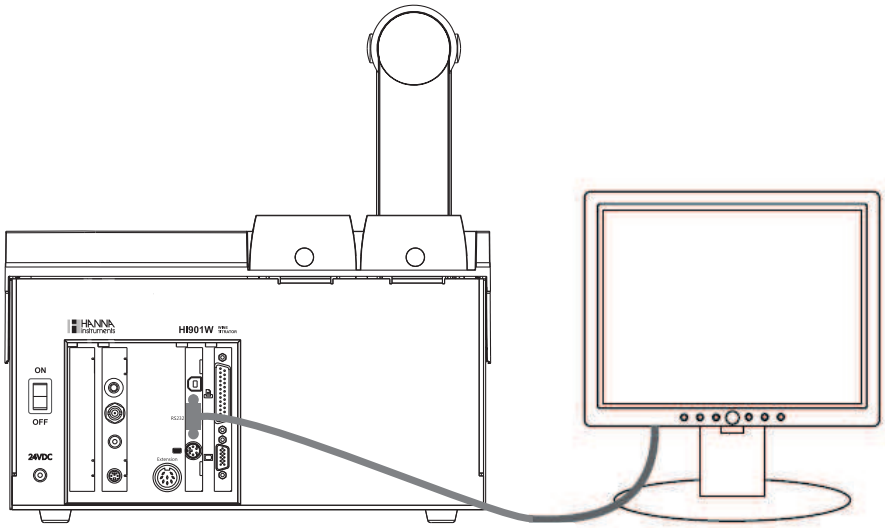
- Remove the aspiration tube from the titrant bottle.
- Repeat burette preparation procedure.
- If this is not successful, clean the burette again.

11.2 Peripherals

Warning! *Connection/disconnection of POWER, PUMP ASSEMBLY, EXTERNAL PC DISPLAY, PRINTER, RS232 INTERFACE must only be done when Titrator and external devices are turned off.*

11.2.1 Connecting an External Display

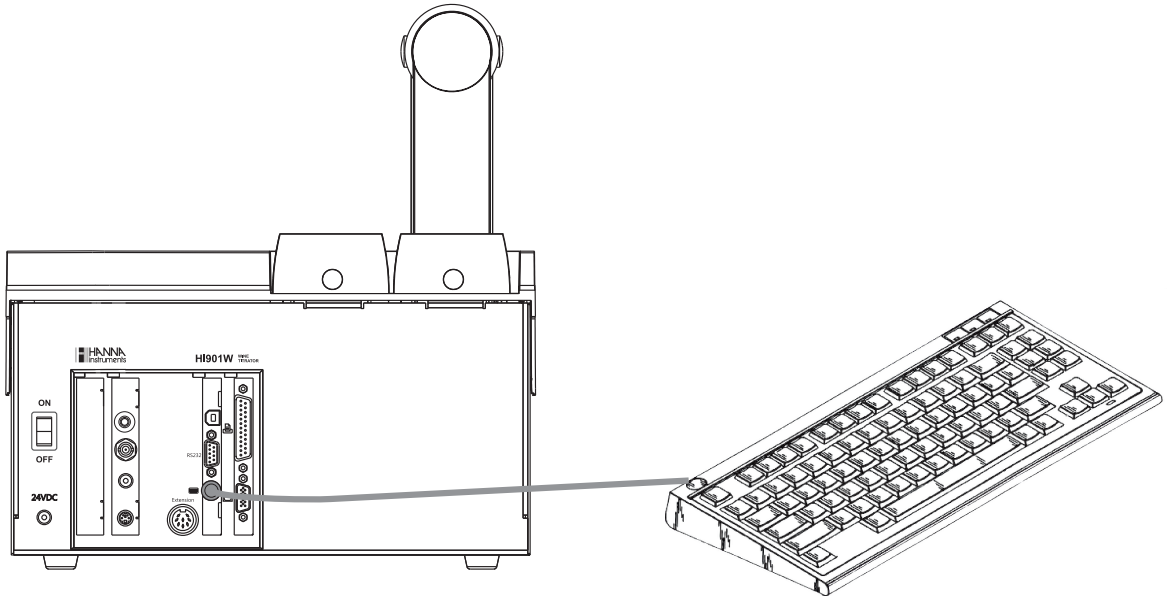
The information shown on the Titrator display can be viewed also on a Standard VGA display connected with a 15-pins cable, as presented below.



Connect the external display to the display socket.
Turn on the Titrator and then the external display.

11.2.2 Connecting an External PC Keyboard

This connection allows you to use an external PS/2 PC Keyboard in addition to the titrator's keypad.



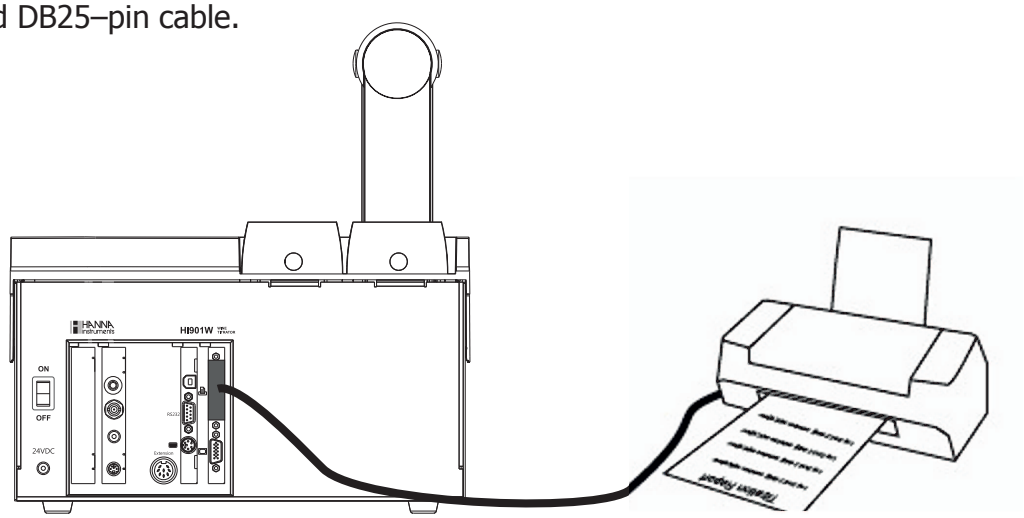
Connect an external PC Keyboard (PS/2 connector).

The correspondence between the titrator's keypad and the United States 101-type external keyboard are:

External PC Keyboard (United States 101)	**Titrator Keypad**
Function Key **F-1**	?
Function Key **F-2**	stir
Function Key **F-3**	results
Function Key **F-4**	device
Function Key **F-5**	Option Key **1** (from left to right)
Function Key **F-6**	Option Key **2** (from left to right)
Function Key **F-7**	Option Key **3** (from left to right)
Function Key **F-8**	Option Key **4** (from left to right)
Function Key **F-9**	Option Key **5** (from left to right)
Function Key **F-10**	start/ stop
Arrow Key: **Up**	△
Arrow Key: **Down**	▽
Arrow Key**: Left**	◁
Arrow Key**: Right**	▷
Page Up	Page Up
Page Down	Page Down
Numeric Keys**: 0 to 9**	0 **to** 9
Tab	Tab
Enter	enter , enter
Alphanumeric Keys	Allow alphanumeric entries.

11.2.3 Connecting a Printer

A variety of parallel printers can be connected to the parallel port of the Titrator using a standard DB25–pin cable.



Warning: *The Titrator and the external printer must be both turned OFF before they are connected.*

Connect the external printer to the standard 25–pin Socket.
Turn on the Titrator and then the printer.

Appendix 1. Contents

HI 901C TECHNICAL SPECIFICATIONS

mV	Range	- 2000.0 to 2000.0 mV
	Resolution	0.1 mV
	Accuracy	±0.1 mV
pH	Range	- 2.000 to 20.000 pH
	Resolution	0.1 / 0.01 / 0.001 pH
	Accuracy	±0.001 pH
ISE	Range	$1x10^{-6}$ to $9.99x10^{10}$
	Resolution	1 / 0.1 / 0.01
	Accuracy	±0.5% (monovalent ion) ±1.0% (divalent ion)
Temperature	Range	- 5.0 to 105.0 °C 23.0 to 221.0 °F 268.2 to 378.2 K
	Resolution	0.1 °C / 0.1 °F / 0.1 K
	Accuracy	±0.1 °C / ±0.2 °F / ±0.1 K
Burette Sizes	Resolution	0.001 mL
	Accuracy	±0.005 mL (5 mL Burette) ±0.010 mL (10 mL Burette) ±0.025 mL (25 mL Burette) ±0.050 mL (50 mL Burette)
Graphic Display		5.7” graphical color display with backlight.
Languages		English, Portuguese, Spanish.
Titration Methods		up to 100 (standard and user methods)

Burette size auto-detection and interchangeable burettes. The Titrator automatically detects the size of the burette when it is slid into the pump assembly.

Propeller Stirrer with Programmable Stir Speed. The stirring speed can be set between 200 and 2500 RPM with 100 RPM resolution.

Flow Rate: user-selectable (see **Titration Methods**, *Volume/Flow Rate* section).

mV / pH / ISE Measurement modes.

Automatically Temperature Compensated pH Measurements.

pH Calibration with up to 5 buffers using *Auto-Entry* or *Manual-Entry* options; temperature compensated buffers are stored internally for *Auto-Entry* option.

Relative mV calibration: single point offset.

ISE Calibration: with up to 5 standards.

APPENDIX 1

Potentiometric Titrations: Acid-Base (pH or mV-Mode), Redox, Precipitation, Complexometric, Non-Aqueous, Ion-Selective, Argentometric.

Titer Determination.

Fixed mV or pH End Point Detection.

Single Equivalence Point Detection with the 1st or 2nd Derivatives of the titration curve.

Flexible Concentration Calculations with many concentration units.

Graph Display during titration, graphs of the stored titration data (mV-Volume or pH-Volume titration curve, 1st derivative curve or 2nd derivative curve, in pH-mode or mV-mode) and pH/mV values versus time-data logging results.

Data Storage: up to 100 complete titration and pH/mV/ISE reports.

Files Copied to and Restored from USB Storage Device: Standard Methods, User Methods, Titration and pH/mV/ISE Logging Reports and Bitmap Files can be transferred to a PC using a USB storage device.

Peripheral Units:

External VGA Display

External PC Keyboard

Printer

GLP Conformity: Good Laboratory Practice and Instrumentation Data storage and printing capabilities.

Mains: 100-240 Vac, 50/60 Hz

Power Draw: 0.5 Amps

Enclosure Material: ABS plastic and Steel

Keypad: Polycarbonate

Dimensions: Width x Depth x Height = 390 x 350 x 380 mm

Weight: approx. 20 lbs. (9 Kg) (with 1 pump, stirrer and sensors)

Operating Environment: 10 to 40 °C, up to 95% relative humidity

Storage Environment: -20 to 70 °C, up to 95% relative humidity

Appendix 2. Contents

A2 ACCESSORIES

A2.1 Solutions

A2.1.1 pH Calibration Solutions

HI7001M —> pH 1.68 Buffer Solution, 230 mL
HI7001L —> pH 1.68 Buffer Solution, 500 mL
HI7004M —> pH 4.01 Buffer Solution, 230 mL
HI7004L —> pH 4.01 Buffer Solution, 500 mL
HI7006M —> pH 6.86 Buffer Solution, 230 mL
HI7006L —> pH 6.86 Buffer Solution, 500 mL
HI7007M —> pH 7.01 Buffer Solution, 230 mL
HI7007L —> pH 7.01 Buffer Solution, 500 mL
HI7009M —> pH 9.18 Buffer Solution, 230 mL
HI7009L —> pH 9.18 Buffer Solution, 500 mL
HI7010M —> pH 10.01 Buffer Solution, 230 mL
HI7010L —> pH 10.01 Buffer Solution, 500 mL

A2.1.2 pH Calibration Solutions in FDA Approved Bottle

HI8004L —> pH 4.01 Buffer Solution, 500 mL
HI8006L —> pH 6.86 Buffer Solution, 500 mL
HI8007L —> pH 7.01 Buffer Solution, 500 mL
HI8009L —> pH 9.18 Buffer Solution, 500 mL
HI8010L —> pH 10.01 Buffer Solution, 500 mL

A2.1.3 pH Technical Calibration Solutions

HI5016 —> pH 1.68 Buffer Solution, 500 mL
HI5003 —> pH 3.00 Buffer Solution, 500 mL
HI5004 —> pH 4.01 Buffer Solution, 500 mL
HI5068 —> pH 6.86 Buffer Solution, 500 mL
HI5007 —> pH 7.01 Buffer Solution, 500 mL
HI5091 —> pH 9.18 Buffer Solution, 500 mL
HI5010 —> pH 10.01 Buffer Solution, 500 mL
HI5124 —> pH 12.45 Buffer Solution, 500 mL

A2.1.4 pH Millesimal Calibration Solutions

HI6016 —> pH 1.679 Buffer Solution, 500 mL
HI6003 —> pH 3.000 Buffer Solution, 500 mL
HI6004 —> pH 4.010 Buffer Solution, 500 mL
HI6004-01 —> pH 4.010 Buffer Solution, 1 L

HI6068 —> pH 6.862 Buffer Solution, 500 mL
HI6007 —> pH 7.010 Buffer Solution, 500 mL
HI6007-01 —> pH 7.010 Buffer Solution, 1 L
HI6091 —> pH 9.177 Buffer Solution, 500 mL
HI6010 —> pH 10.010 Buffer Solution, 500 mL
HI6010-01 —> pH 10.010 Buffer Solution, 1 L
HI6124 —> pH 12.450 Buffer Solution, 500 mL

A2.1.5 Electrode Cleaning Solutions

HI7061M —> General Purpose Solution, 230 mL
HI7061L —> General Purpose Solution, 500 mL
HI7073M —> Protein Cleaning Solution, 230 mL
HI7073L —> Protein Cleaning Solution, 500 mL
HI7074M —> Inorganic Cleaning Solution, 230 mL
HI7074L —> Inorganic Cleaning Solution, 500 mL
HI7077M —> Oil & Fat Cleaning Solution, 230 mL
HI7077L —> Oil & Fat Cleaning Solution, 500 mL

A2.1.6 Electrode Cleaning Solutions in FDA Approved Bottle

HI8061L —> General Purpose Solution, 500 mL
HI8073L —> Protein Cleaning Solution, 500 mL
HI8077L —> Oil & Fat Cleaning Solution, 500 mL

A2.1.7 Electrode Storage Solutions

HI70300M —> Storage Solution, 230 mL
HI70300L —> Storage Solution, 500 mL

A2.1.8 Electrode Storage Solutions in FDA Approved Bottle

HI80300M —> Storage Solution, 230 mL
HI80300L —> Storage Solution, 500 mL

A2.1.9 Refilling Electrolyte Solutions

HI7071 —> 3.5M KCl + AgCl Electrolyte, 30 mL, for single junction electrodes
HI7072 —> 1M KNO_3 Electrolyte, 30 mL
HI7075 —> KNO_3 and KCl Electrolyte, 30 mL
HI7076 —> 1M NaCl Electrolyte, 30 mL
HI7078 —> $(NH_4)_2SO_4$ Electrolyte, 30 mL
HI7082 —> 3.5M KCl Electrolyte, 30 mL, for double junction electrodes

A2.1.10 Refilling Electrolyte Solutions in FDA Approved Bottle

HI8071 —> 3.5M KCl + AgCl Electrolyte, 30 mL, for single junction electrodes
HI8082 —> 3.5M KCl Electrolyte, 30 mL, for double junction electrodes

A2.1.11 ORP Pretreatment Solutions

HI7091M —> Reducing Pretreatment Solution, 230 mL
HI7091L —> Reducing Pretreatment Solution, 500 mL
HI7092M —> Oxidizing Pretreatment Solution, 230 mL
HI7092L —> Oxidizing Pretreatment Solution, 500 mL

A2.1.12 Titration Reagents

HI70429 —> 0.05 M $AgNO_3$ Titration Reagent, 1 L
HI70433 —> 0.01 N Stabilized Iodine Titration Reagent, 1 L
HI70439 —> 0.1 M $Na_2S_2O_3$ Titration Reagent, 1 L
HI70440 —> 0.02 N Stabilized Iodine Titration Reagent, 1 L
HI70441 —> 0.04 N Stabilized Iodine Titration Reagent, 1 L
HI70448 —> 0.02 M $AgNO_3$ Titration Reagent, 1 L
HI70449 —> 0.02 M EDTA Titration Reagent, 1 L
HI70455 —> 0.01 N NaOH Titration Reagent, 1 L
HI70456 —> 0.1 N NaOH Titration Reagent, 1 L
HI70457 —> 1 N NaOH Titration Reagent, 1 L
HI70458 —> 0.01 M H_2SO_4 Titration Reagent, 1 L
HI70459 —> 0.05 M H_2SO_4 Titration Reagent, 1 L
HI70462 —> 0.01 N HCl Titration Reagent, 1 L
HI70463 —> 0.1 N HCl Titration Reagent, 1 L
HI70464 —> 1 N HCl Titration Reagent, 1 L

A2.1.13 Ion Selective Electrode Calibration Solutions

HI4001-01 —> 0.1 M Ammonia Standard
HI4001-02 —> 100 ppm Ammonia Standard (as N)
HI4001-03 —> 1000 ppm Ammonia Standard (as N)
HI4002-01 —> 0.1 M Bromide Standard
HI4003-01 —> 0.1 M Cadmium Standard
HI4004-01 —> 0.1 M Calcium Standard
HI4005-01 —> 0.1 M Carbon Dioxide Standard
HI4005-03 —> 1000 ppm Carbon Dioxide Standard (as $CaCO_3$)
HI4007-01 —> 0.1 M Chloride Standard
HI4007-02 —> 100 ppm Chloride Standard
HI4007-03 —> 1000 ppm Chloride Standard
HI4008-01 —> 0.1 M Cupric Standard
HI4010-01 —> 0.1 M Fluoride Standard
HI4010-02 —> 100 ppm Fluoride Standard
HI4010-03 —> 1000 ppm Fluoride Standard
HI4011-01 —> 0.1 M Iodide Standard
HI4012-01 —> 0.1 M Lead Standard

HI4012-21 —> 0.1 M Sulfate Standard
HI4013-01 —> 0.1 M Nitrate Standard
HI4013-02 —> 100 ppm Nitrate Standard
HI4013-03 —> 1000 ppm Nitrate Standard
HI4014-01 —> 0.1 M Potassium Standard
HI4015-01 —> 0.1 M Silver Standard

A2.2 Sensors

A2.2.1 pH Electrodes

HI1043B
Glass-body, double junction, refillable, combination pH electrode.
Use: strong acid and base, paint and solvents

HI1053B
Glass-body, triple ceramic, conic shape, refillable, combination pH electrode.
Use: emulsions, fats and creams, soil and semi-solids samples

HI1083B
Glass-body, micro, Viscolene, nonrefillable, combination pH electrode.
Use: biotechnology and micro titration

HI1131B
Glass-body, double junction, refillable, combination pH electrode.
Use: general purpose

HI1330B
Glass-body, semimicro, single junction, refillable, combination pH electrode.
Use: laboratory, vials, and test tubes

HI1331B
Glass-body, semimicro, single junction, refillable, combination pH electrode.
Use: flasks

HI1230B
Plastic-body (PEI), double junction, gel-filled, combination pH electrode.
Use: general purpose

HI2031B
Glass-body, conical tip, refillable, combination pH electrode.
Use: dairy and semi-solid products

HI1332B
Plastic-body (PEI), double junction, refillable, combination pH electrode.
Use: chemicals, field applications and quality control testing.

FC100B
Plastic-body (PVDF), double junction, refillable, combination pH electrode.
Use: sauces, juices, dairy products and other liquids or slurry forms of food

FC200B
Plastic-body (PVDF), single junction, conical tip, non-refillable Viscolene electrolyte, combination pH electrode.
Use: dairy, dough, ground meats and other semi-solid food

FC210B
Glass-body, double junction, conical tip, non-refillable Viscolene electrolyte, combination pH electrode.
Use: milk, yogurt, and cream

FC220B
Glass-body, single junction, refillable, combination pH electrode.
Use: milk, yogurt, cream, sauce, and fruit juices

FC911B
Plastic-body (PVDF), double junction, refillable, combination pH electrode.
Use: sauce, juices, dairy products and other liquid or slurry forms of food

HI1413B
Glass-body, single junction, flat tip, non-refillable Viscolene electrolyte, combination pH electrode.
Use: surfaces, skin, leather, paper, and emulsions

A2.2.2 ORP Electrodes

HI3131B
Glass-body, refillable, combination platinum ORP electrode.
Use: laboratories and general purpose

HI3230B
Plastic-body (PEI), gel-filled, combination platinum ORP electrode.
Use: municipal water and quality control

HI4430B
Plastic-body (PEI), gel-filled, combination gold ORP electrode.
Use: oxidants and ozone

A2.2.3 Half-cell Electrodes

HI2110B
Glass-body, single half-cell pH electrode.
Use: general purpose

HI5311
Glass-body, Ag/AgCl reference half-cell electrode, double junction, refillable with 4mm banana plug with 1m (3.3') cable.
Use: general purpose with wide temperature range

HI5315
Plastic-body (PEI), double junction, Ag/AgCl reference half-cell electrode, refillable with 4mm plug with 1 m (3.3') cable.
Use: Ion Selective Electrodes

HI5412
Glass-body, single Calomel reference half-cell electrode, refillable with 4mm plug with 1m (3.3') cable.
Use: general purpose with constant temperature range

A2.2.4 Ion Selective Electrodes

HI4101 Ammonia ISE

HI4002 / HI4102 Bromide ISE

HI4003 / HI4103 Cadmium ISE

HI4004 / HI4104 Calcium ISE

HI4105 Carbon Dioxide ISE

HI4007 / HI4107 Chloride ISE

HI4008 / HI4108 Cupric ISE

HI4009 / HI4109 Cyanide ISE

HI4010 / HI4110 Fluoride ISE

HI4011 / HI4111 Iodide ISE

HI4012 / HI4112 Lead ISE

HI4013 / HI4113 Nitrate ISE

HI4014 / HI4114 Potassium ISE

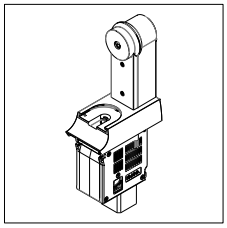
HI4015 / HI4115 Silver / Sulfide ISE

FC300B Sodium

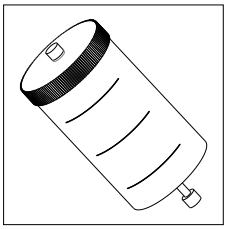
A2.2.5 Temperature Sensor

HI7662-T
Temperature probe with 1 m (3.3') paneled cable.

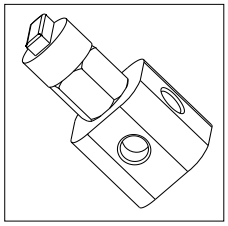
A2.3 Titrator components



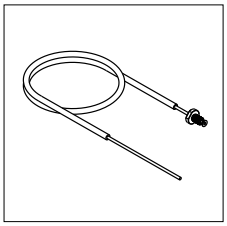
Pump Assembly
HI900100



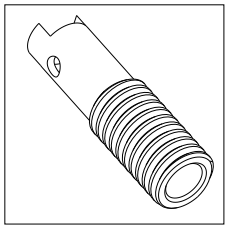
50 mL Syringe
HI900250



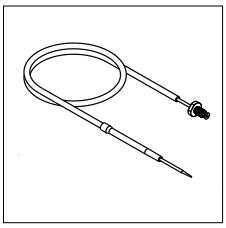
3 Way Valve
HI900260



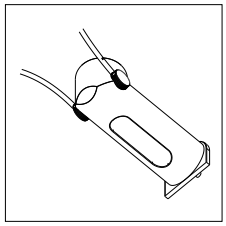
Aspiration Tube with fitting and protection tube
HI900270



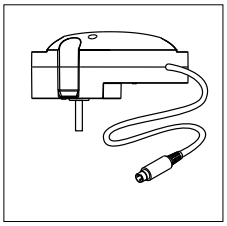
Tool for burette cap removal
HI900942



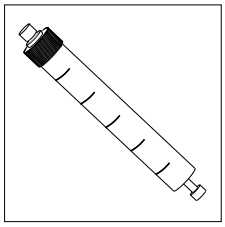
Dispensing Tube with normal dispensing tip, fitting, protection tube and tube guide
HI900280



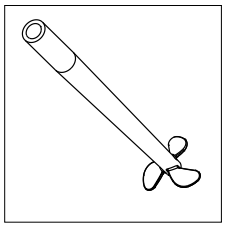
Burette with:
5 mL syringe - **HI900105**
10 mL syringe - **HI900110**
25 mL syringe - **HI900125**
50 mL syringe - **HI900150**



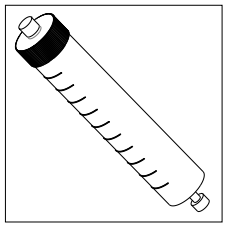
Overhead Stirrer +
3 propellers
HI900301



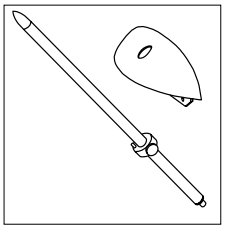
5 mL Syringe
HI900205



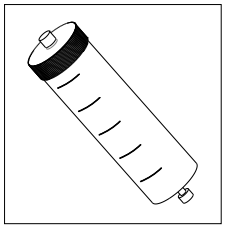
Propeller
HI900302



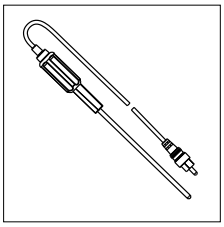
10 mL Syringe
HI900210



Stirrer Support and Stand
HI900320

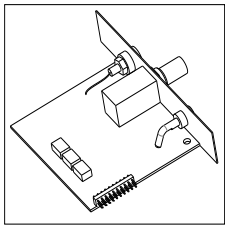


25 mL Syringe
HI900225

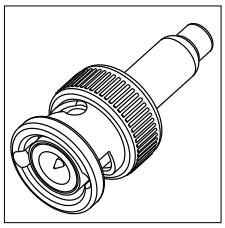


Temperature Probe
HI7662-T

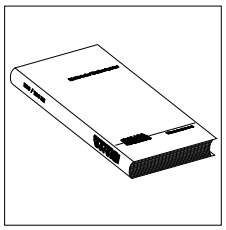
APPENDIX 2



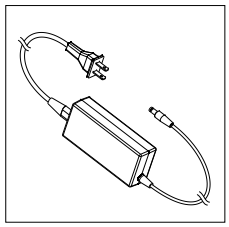
Potentiometric Analog Board
HI900401



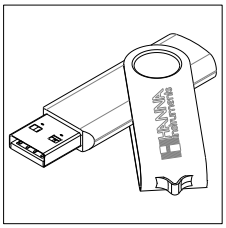
Shorting Cap
HI900945



Instruction Manual Binder
HI900801W



Power Adapter
HI900946



USB Storage Device
HI900900W

0.1N Sodium Hydroxide Titrant Concentration

Description:
Method for the standardization (titer determination) of 0.1N Sodium Hydroxide (NaOH) titrant solution against Potassium Hydrogen Phthalate (KHP). The results are expressed in **N (eq/L)**.

Reference:
AOAC Official Methods of Analysis, Official Method 936.16

Electrode:
- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

Reagents:
- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70401 Potassium Hydrogen Phthalate (20 g)
- HI70436 Deionized Water (1 gal)

Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- HI740036P 100-mL Plastic Beakers (10 pcs)
- Analytical Balance with 0.0001 g resolution

Device Preparation:
- Connect the pH electrode and temperature probe to the titrator.
- Install a 25-mL burette filled with 0.1N sodium hydroxide (HI70456) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight 'HI0001EN 0.1N Sodium Hydroxide' and press *"Select".*

Electrode Preparation:
- Press "*Mode*" from the main screen and press *"pH".*
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation:
- Crush approximately 3 grams of potassium hydrogen phthalate (HI70401) and dry it for 2 hours at 120°C. Cool to room temperature in a desiccator.
- Place a clean 100-mL plastic beaker on the analytical balance.
- Zero the balance.
- Carefully weigh approximately 0.20 grams of dried potassium hydrogen phthalate into the beaker.

  NOTE: Ensure that all of the potassium hydrogen phthalate is on the bottom of the beaker.
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Remove the beaker from the balance and add deionized water to the 50-mL mark on the beaker.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press *"Start".* You will be prompted to enter the analyte size (weight of potassium hydrogen phthalate). Use the numeric keypad to enter the exact weight and press *"Enter"* to start the analysis.
  NOTE: Ensure that the potassium hydrogen phthalate dissolves completely during the pre-titration stir time. Erroneous results may occur if the sample does not dissolve completely prior to titration. If necessary the pre-titration stir time can be increased.
- At the end of the titration, after detection of the equivalence point, 'titration complete' will appear with the titrant concentration. The result is expressed in **N (eq/L) of sodium hydroxide**.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

NOTE: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1N sodium hydroxide titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1N sodium hydroxide.
- Select *"Method Options"* from the main screen.
- Using the arrow keys, highlight 'Titrant Conc.' and press *"Select".*
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press *"Accept".*
- Press *"Escape"* to exit the Method Options screen and select 'Save Method' option.

0.1N Sodium Hydroxide Titrant Concentration

Method Parameters:

Name:	0.1N Sodium Hydroxide
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.030 mL
max Vol:	0.500 mL
delta E:	4.500 mV
End Point Mode:	pH 1EQ point, 1st Dev
Recognition Options:	
Threshold:	500 mV/mL
Range:	NO
Filtered Derivatives:	NO
Pre-Titration Volume:	5.000 mL
Pre-Titration Stir Time:	60 Sec
Measurement Mode:	Signal Stability
delta E:	0.3 mV
delta t:	1.5 Sec
t-min wait:	3 Sec
t-max wait:	30 Sec
Electrode Type:	pH
Blank Option:	No Blank
Calculations:	Stdz. Titrant by Weight
Dilution Option:	Disabled
Titrant Name:	0.1N NaOH
Analyte Size:	0.200 g
Analyte Entry:	Manual
Maximum Titrant Volume:	15.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	1 Reading
Final Result Format:	XXXXX

Calculations:

Calculations:	Stdz. Titrant by Weight
Titrant units:	N (eq/L)
Titrant volume dosed:	V (L)
Standard weight:	0.200 g
Titrant/Standard:	1.000 eq/mol
MW of standard:	204.23 g/mol

$$N\ (eq/L) = \frac{0.200*1.000}{204.23*V(L)}$$

Results:

Titration Report

Method Name:	0.1N Sodium Hydroxide
Time & Date:	17:03 Jun 07, 2016
Titration ID:	Ti_00053

Titration Results

Method Name:	0.1N Sodium Hydroxide
Time & Date:	17:03 Jun 07, 2016
Analyte size:	0.20920 g
End Point Volume:	10.215 mL
pH Equivalence Point:	8.394
Results:	0.10027 N(eq/L)
Initial and Final pH:	4.173 to 9.570
Titration Duration:	6:25 [mm:ss]

Titration went to Completion

Operator name: _____

0.1M Sodium Thiosulfate Titrant Concentration

Description:
Method for the standardization (titer determination) of 0.1M Sodium Thiosulfate ($Na_2S_2O_3$) titrant solution against Potassium Iodate (KIO_3). The results are expressed in **M (mol/L)**.

Reference:
Standard Methods for the Examination of Water and Wastewater 19th Edition, Method 4500-Cl B

Electrode:
- HI3131B Combination ORP Electrode

Reagents:
- HI70439 0.1M Sodium Thiosulfate (1 L)
- HI70407 Potassium Iodate (20 g)
- HI70425 16% Sulfuric Acid (500 mL)
- HI70468 Potassium Iodide (35 g)
- HI70436 Deionized Water (1 gal)

Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI740036P 100-mL Plastic Beakers (10 pcs)
- Analytical Balance 0.0001 g
- 100-mL Class-A Volumetric Flask
- 10-mL Class-A Volumetric Pipette

Device Preparation:
- Connect the ORP electrode to the titrator.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight 'HI0003EN 0.1M Sodium Thiosulfate' and press *"Select".*
- Install a 25-mL burette filled with 0.1M sodium thiosulfate (HI70439) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.

Electrode Preparation:
- Prepare the ORP electrode according to the procedure in the manual.

Sample Preparation:
- Crush approximately 2 grams of potassium iodate (HI70407) and dry it for 2 hours at 120°C. Cool to room temperature in a desiccator.
- Carefully weigh approximately 0.35 grams of dried potassium iodate.
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Carefully transfer the salt to a 100-mL class-A volumetric flask. Add approximately 80 mL of deionized water, and mix to dissolve. Once the salt is completely dissolved bring the flask to volume with deionized water, mix well.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of the solution to a clean 100-mL plastic beaker. Add deionized water to the 50-mL mark on the beaker.
- Add 5 mL of 16% sulfuric acid (HI70425) and 1.5 grams of potassium iodide (HI70468) to the beaker.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press *"Start".* You will be prompted to enter the analyte size (weight of potassium iodate). Use the numeric keypad to enter the exact weight and press *"Enter"* to start the analysis.
- At the end of the titration, after detection of the equivalence point, 'titration complete' will appear with the titrant concentration. The result is expressed in **M (mol/L) of sodium thiosulfate**.
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

NOTE: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1M sodium thiosulfate titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1M sodium thiosulfate.
- Select *"Method Options"* from the main screen.
- Using the arrow keys, highlight Titrant Conc.' and press *"Select".*
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press 'Accept'.
- Press *"Escape"* to exit the Method Options screen and select 'Save Method' option.

0.1M Sodium Thiosulfate Titrant Concentration

Method Parameters:

Name:	0.1M Sodium Thiosulfate
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirrer Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.030 mL
max Vol:	0.600 mL
delta E:	6.500 mV
End Point Mode:	mV 1EQ point, 1st Dev
Recognition Options:	
Threshold:	50 mV/mL
Range:	NO
Filtered Derivatives:	NO
Pre-Titration Volume:	5.000 mL
Pre-Titration Stir Time:	0 Sec
Measurement Mode:	Signal Stability
delta E:	0.3 mV
delta t:	2.0 Sec
t-min wait:	2 Sec
t-max wait:	20 Sec
Electrode Type:	ORP
Blank Option:	No Blank
Calculations:	Stdz. Titrant by Weight
Dilution Option:	Enabled
Final Dilution Volume:	100.000 mL
Aliquot Volume:	10.000 mL
Titrant Name:	0.1M Na2S2O3
Analyte Size:	0.35000 g
Analyte Entry:	Manual
Maximum Titrant Volume:	15.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	1 Reading
Final Result Format:	XXXXX

Calculations:

Calculations:	Stdz. Titrant by Weight
Titrant units:	M (mol/L)
Titrant volume dosed:	V (L)
Standard weight:	0.350 g
Dilution Factor:	0.100
Final Dilution volume:	100.000 mL
Aliquot Volume:	10.000 mL
Titrant/Standard:	6.000 mol/mol
MW of standard:	214.00 g/mol

$$M\ (mol/L) = \frac{0.350 * 0.100 * 6.000}{214.00 * V(L)}$$

Results:

Titration Report

Method Name:	0.1M Sodium Thiosulfate
Time & Date:	17:10 Jun 22, 2016
Titration ID:	Ti_00073

Titration Results

Method Name:	0.1M Sodium Thiosulfate
Time & Date:	17:10 Jun 22, 2016
Analyte size:	0.35020 g
End Point Volume:	9.635 mL
mV Equivalence Point:	233.0
Results:	0.10191 M (mol/L)
Initial and Final mV:	361.8 to 173.4
Titration Duration:	2:51 [mm:ss]

Titration went to Completion

Operator name: _____

0.01N Sodium Hydroxide Titrant Concentration

Description:
Method for the standardization (titer determination) of 0.01N Sodium Hydroxide (NaOH) titrant solution against Potassium Hydrogen Phthalate (KHP). The results are expressed in **N (eq/L)**.

Reference:
AOAC Official Methods of Analysis, Official Method 936.16

Electrode:
- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

Reagents:
- HI70455 0.01N Sodium Hydroxide (1 L)
- HI70401 Potassium Hydrogen Phthalate (20 g)
- HI70436 Deionized Water (1 gal)

Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- HI740036P 100-mL Plastic Beakers (10 pcs)
- Analytical Balance with 0.0001 g resolution

Device Preparation:
- Connect the pH electrode and temperature probe to the titrator.
- Install a 25-mL burette filled with 0.01N sodium hydroxide (HI70455) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight 'HI0015EN 0.01N Sodium Hydroxide' and press *"Select".*

Electrode Preparation:
- Press *"Mode"* from the main screen and press *"pH".*
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation:
- Crush approximately 3 grams of potassium hydrogen phthalate (HI70401) and dry it for 2 hours at 120°C. Cool to room temperature in a desiccator.
- Place a clean 100-mL plastic beaker on the analytical balance.
- Zero the balance.
- Carefully weigh approximately 0.02 grams of dried potassium hydrogen phthalate into the beaker.

  NOTE: Ensure that all of the potassium hydrogen phthalate is on the bottom of the beaker.
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Remove the beaker from the balance and add deionized water to the 50-mL mark on the beaker.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5-6 mm below the surface. If necessary add extra deionized water.
- **NOTE:** The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press *"Start"*. You will be prompted to enter the analyte size (weight of potassium hydrogen phthalate). Use the numeric keypad to enter the exact weight and press *"Enter"* to start the analysis.

  NOTE: Ensure that the potassium hydrogen phthalate dissolves completely during the pre-titration stir time. Erroneous results may occur if the sample does not dissolve completely prior to titration. If necessary the pre-titration stir time can be increased.
- At the end of the titration, after detection of the equivalence point, 'titration complete' will appear with the titrant concentration. The result is expressed in **N (eq/L) of sodium hydroxide**.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

NOTE: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.01N sodium hydroxide titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.01N sodium hydroxide.
- Select *"Method Options"* from the main screen.
- Using the arrow keys, highlight 'Titrant Conc.' and press *"Select".*
- Use the numeric keypad to enter the standardized (titer) value of the titrant then *press "Accept".*
- Press *"Escape"* to exit the Method Options screen and select 'Save Method' option.

0.01N Sodium Hydroxide Titrant Concentration

Method Parameters:

Name:	0.01N Sodium Hydroxide
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.030 mL
max Vol:	0.500 mL
delta E:	4.500 mV
End Point Mode:	pH 1EQ point, 1st Dev
Recognition Options:	
Threshold:	185 mV/mL
Range:	NO
Filtered Derivatives:	NO
Pre-Titration Volume:	5.000 mL
Pre-Titration Stir Time:	60 Sec
Measurement Mode:	Signal Stability
delta E:	0.3 mV
delta t:	1.5 Sec
t-min wait:	3 Sec
t-max wait:	30 Sec
Electrode Type:	pH
Blank Option:	No Blank
Calculations:	Stdz. Titrant by Weight
Dilution Option:	Disabled
Titrant Name:	0.01N NaOH
Analyte Size:	0.0200 g
Analyte Entry:	Manual
Maximum Titrant Volume:	15.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	1 Reading
Final Result Format:	XXXXX

Calculations:

Calculations:	Stdz. Titrant by Weight
Titrant units:	N (eq/L)
Titrant volume dosed:	V (L)
Standard weight:	2.000E-2 g
Titrant/Standard:	1.000 eq/mol
MW of standard:	204.23 g/mol

$$N\ (eq/L) = \frac{2.000E\text{-}2 * 1.000}{204.23 * V(L)}$$

Results:

Titration Report

Method Name:	0.01N Sodium Hydroxide
Time & Date:	15:35 Jun 09, 2016
Titration ID:	Ti_00015

Titration Results

Method Name:	0.01N Sodium Hydroxide
Time & Date:	15:35 Jun 09, 2016
Analyte size:	0.01960 g
End Point Volume:	10.555 mL
pH Equivalence Point:	8.146
Results:	0.00909 N(eq/L)
Initial and Final pH:	4.987 to 8.640
Titration Duration:	7:02 [mm:ss]

Titration went to Completion

Operator name: _____

0.02N Iodine Titrant Concentration

Description:
Method for the standardization (titer determination) of 0.02N Iodine (I_2) titrant solution against Sodium Thiosulfate ($Na_2S_2O_3$). The results are expressed in **N (eq/L)**.

Reference:
Standard Methods for the Examination of Water and Wastewater 19th Edition, Method 4500-Cl C

Electrode:
- HI3131B Combination ORP Electrode

Reagents:
- HI70440 0.02N Stabilized Iodine (1 L)
- HI70403 Sodium Thiosulfate (20 g)
- HI70444 25% Sulfuric Acid (500 mL)
- HI70404 KI Powder Packets (100 pcs)
- HI70436 Deionized Water (1 gal)

Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI740036P 100-mL Plastic Beakers (10 pcs)
- Analytical Balance 0.0001 g
- 100-mL Class-A Volumetric Flask
- 10-mL Class-A Volumetric Pipette

Device Preparation:
- Connect the ORP electrode to the titrator.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight 'HI0204EN 0.02N Iodine' and press *"Select"*.
- Install a 25-mL burette filled with 0.02N stabilized iodine (HI70440) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.

Electrode Preparation:
- Prepare the ORP electrode according to the procedure in the manual.

Sample Preparation:
- Carefully weigh approximately 0.40 grams of sodium thiosulfate (HI70403).
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Carefully transfer the salt to a 100-mL class-A volumetric flask. Add approximately 80 mL of deionized water, and mix to dissolve. Once the salt is completely dissolved bring the flask to volume with deionized water, mix well.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of the solution to a clean 100-mL plastic beaker. Add deionized water to the 50-mL mark on the beaker.
- Add 7 mL of 25% sulfuric acid (HI70444) and one HI70404 powder packet to the beaker.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press *"Start"*. You will be prompted to enter the analyte size (weight of sodium thiosulfate). Use the numeric keypad to enter the exact weight and press *"Enter"* to start the analysis.
- At the end of the titration, after detection of the equivalence point, 'titration complete' will appear with the titrant concentration. The result is expressed in **N (eq/L) iodine**.
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

NOTE: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.02N iodine titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.02N iodine.
- Select *"Method Options"* from the main screen.
- Using the arrow keys, highlight 'Titrant Conc.' and press *"Select".*
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press *"Accept".*
- Press *"Escape"* to exit the Method Options screen and select 'Save Method' option.

0.02N Iodine Titrant Concentration

Method Parameters:

Name:	0.02N Iodine
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirrer Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.050 mL
max Vol:	0.500 mL
delta E:	5.000 mV
End Point Mode:	mV 1EQ point, 1st Dev
Recognition Options:	
Threshold:	30 mV/mL
Range:	NO
Filtered Derivatives:	NO
Pre-Titration Volume:	5.000 mL
Pre-Titration Stir Time:	30 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	1.0 Sec
t-min wait:	2 Sec
t-max wait:	20 Sec
Electrode Type:	ORP
Blank Option:	No Blank
Calculations:	Stdz. Titrant by Weight
Dilution Option:	Enabled
Final Dilution Volume:	100.000 mL
Aliquot Volume:	10.000 mL
Titrant Name:	0.02N I2
Analyte Size:	0.4000 g
Analyte Entry:	Manual
Maximum Titrant Volume:	15.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	1 Reading
Final Result Format:	XXXXX

Calculations:

Calculations:	Stdz. Titrant by Weight
Titrant units:	M (mol/L)
Titrant volume dosed:	V (L)
Standard weight:	0.400 g
Dilution Factor:	0.100
Final Dilution volume:	100.000 mL
Aliquot Volume:	10.000 mL
Titrant/Standard:	1.000 eq/mol
MW of standard:	248.18 g/mol

$$M\ (mol/L) = \frac{0.400 * 0.100 * 1.000}{248.18 * V(L)}$$

Results:

Titration Report

Method Name:	0.02N Iodine
Time & Date:	15:17 Jun 22, 2016
Titration ID:	Ti_00060

Titration Results

Method Name:	0.02N Iodine
Time & Date:	15:17 Jun 22, 2016
Analyte size:	0.45320 g
End Point Volume:	9.261 mL
mV Equivalence Point:	316.9
Results:	0.01972 N(eq/L)
Initial and Final mV:	212.2 to 375.2
Titration Duration:	3:34[mm:ss]

Titration went to Completion

Operator name: _____

Total Titratable Acidity in Wine, pH 7.00

Description:
Method for the determination of total titratable acidity (TA) in wine, by titration of a 10-mL degassed sample to a fixed end point of pH 7.00. The results are expressed in **g/L of tartaric acid**.

Reference:
AOAC International, Official Method 962.12 Acidity (Titratable) of Wines

Electrode:
- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

Reagents:
- HI 70456 0.1N Sodium Hydroxide (1 L)
- HI 70436 Deionized Water (1 gal)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- HI740036P 100-mL Plastic Beakers (10 pcs)
- 10-mL Class-A Volumetric Pipette

Device Preparation:
- Connect the pH electrode and temperature probe to the titrator.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight 'HI3204EN Titratable Acidity pH7.0' method and press *"Select".*
- Install a 25-mL burette with 0.1N sodium hydroxide (HI70456) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
  For the determination of the exact concentration of the 0.1N sodium hydroxide, follow HI0001EN 0.1N Sodium Hydroxide Titrant Concentration.

Electrode Preparation:
- Press *"Mode"* from the main screen and press *"pH".*
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation:
- Degas the wine sample to remove CO_2 by passing nitrogen through it, using an ultrasonic bath, or by stirring and applying vacuum. Degassing can also be achieved by boiling the wine for several seconds. The wine must be cooled to room temperature before it is used.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of degassed sample to a clean 100-mL plastic beaker. Add deionized water to the 50-mL mark on the beaker.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press *"Start".* The titrator will start the analysis.
- At the end of titration, when pH 7.00 is reached, 'titration complete' will appear with the total titratable acidity concentration. The result is expressed in **g/L of tartaric acid**.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Total Titratable Acidity in Wine, pH 7.00

Method Parameters:

Name:	Titratable Acidity pH7.0
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.050 mL
max Vol:	0.800 mL
delta E:	8.000 mV
End Point Mode:	Fixed 7.000 pH
Pre-Titration Volume:	3.000 mL
Pre-Titration Stir Time:	15 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	1.5 Sec
t-min wait:	2 Sec
t-max wait:	20 Sec
Electrode Type:	pH
Blank Option:	No Blank
Calculations:	Sample Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.1N NaOH
Titrant Conc.:	0.1000 N (eq/L)
Analyte Size:	10.0000 mL
Analyte Entry:	Fixed
Maximum Titrant Volume:	25.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	1 Reading
Final Result Format:	XXX

Calculations:

Calculations:	Sample Calc. by Volume
Titrant units:	N (eq/L)
Titrant volume dosed:	V (L)
Final result units:	g/L
Titrant conc.:	0.100 N (eq/L)
Sample/Titrant:	0.500 mol/eq
MW of sample:	150.09 g/mol
Sample volume:	10.000 mL

$$ {}^{g}/_{L} = \frac{V(L)*0.100*0.5*150.09*1000}{10.000} $$

Results:

Titration Report

Method Name:	Titratable Acidity pH7.0
Time & Date:	11:15 Aug 05, 2016
Titration ID:	Ti_00011

Titration Results

Method Name:	Titratable Acidity pH7.0
Time & Date:	11:15 Aug 05, 2016
Analyte size:	10.000 mL
End Point Volume:	7.273 mL
pH Fixed End Point:	7.000
Results:	5.15 g/L
Initial and Final pH:	3.273 to 7.049
Titration Duration:	2:40 [mm:ss]

Titration went to Completion

Operator name: _____

Total Titratable Acidity in Wine, pH 8.20

Description:
Method for the determination of total titratable acidity (TA) in wine, by titration of a 10-mL degassed sample to a fixed end point of pH 8.20. The results are expressed in **g/L of tartaric acid**.

Reference:
AOAC International, Official Method 962.12 Acidity (Titratable) of Wines

Electrode:
- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

Reagents:
- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70436 Deionized Water (1 gal)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- HI740036P 100-mL Plastic Beakers (10 pcs)
- 10-mL Class-A Volumetric Pipette

Device Preparation:
- Connect the pH electrode and temperature probe to the titrator.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight 'HI3205EN Titratable Acidity pH8.2' method and press *"Select".*
- Install a 25-mL burette with 0.1N sodium hydroxide (HI70456) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
  For the determination of the exact concentration of the 0.1N sodium hydroxide, follow HI0001EN 0.1N Sodium Hydroxide Titrant Concentration.

Electrode Preparation:
- Press *"Mode"* from the main screen and press *"pH".*
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation:
- Degas the wine sample to remove CO_2 by passing nitrogen through it, using an ultrasonic bath, or by stirring and applying vacuum. Degassing can also be achieved by boiling the wine for several seconds. The wine must be cooled to room temperature before it is used.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of degassed sample to a clean 100-mL plastic beaker. Add deionized water to the 50-mL mark on the beaker.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press *"Start".* The titrator will start the analysis.
- At the end of titration, when pH 8.20 is reached, 'titration complete' will appear with the total titratable acidity concentration. The result is expressed in **g/L of tartaric acid**.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Total Titratable Acidity in Wine, pH 8.20

Method Parameters:

Name:	Titratable Acidity pH8.2
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.050 mL
max Vol:	0.800 mL
delta E:	8.000 mV
End Point Mode:	Fixed 8.200 pH
Pre-Titration Volume:	3.000 mL
Pre-Titration Stir Time:	15 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	1.5 Sec
t-min wait:	2 Sec
t-max wait:	20 Sec
Electrode Type:	pH
Blank Option:	No Blank
Calculations:	Sample Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.1N NaOH
Titrant Conc.:	0.1000 N (eq/L)
Analyte Size:	10.0000 mL
Analyte Entry:	Fixed
Maximum Titrant Volume:	25.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	1 Reading
Final Result Format:	XXX

Calculations:

Calculations:	Sample Calc. by Volume
Titrant units:	N (eq/L)
Titrant volume dosed:	V (L)
Final result units:	g/L
Titrant conc.:	0.100 N (eq/L)
Sample/Titrant:	0.500 mol/eq
MW of sample:	150.09 g/mol
Sample volume:	10.000 mL

$$ {}^{g}/_{L} = \frac{V(L)*0.10000*0.5*150.09*1000}{10.000} $$

Results:

Titration Report

Method Name:	Titratable Acidity pH8.2
Time & Date:	14:49 Sep 21, 2016
Titration ID:	Ti_00046

Titration Results

Method Name:	Titratable Acidity pH8.2
Time & Date:	14:49 Sep 21, 2016
Analyte size:	10.000 mL
End Point Volume:	7.915 mL
pH Fixed End Point:	8.2000
Results:	5.64 g/L
Initial and Final pH:	3.472 to 8.293
Titration Duration:	4:08 [mm:ss]

Titration went to Completion

Operator name: _____

Volatile Acidity in Wine

Description:
Method for the determination of volatile acidity in wine, by titration of a distillate that is collected from a steam distillation apparatus (Volatile Acid / Cash Still). The results are expressed in **g/L of acetic acid**.
Sulfur Dioxide interferences are eliminated by addition of hydrogen peroxide prior to the steam distillation.

Reference:
Wine Analysis and Production Acetic Acid: Steam Distillation of Volatile Acid using Still

Electrode:
- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

Reagents:
- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70432 3% Hydrogen Peroxide (25 mL)
- HI70436 Deionized Water (1 gal)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- Volatile Acid (Cash) Still
- 10-mL Class-A Volumetric Pipette
- 150-mL Glass Beaker

Device Preparation:
- Connect the pH electrode and temperature probe to the titrator.
- Press "*Select Method*" from the main screen. Use the arrow keys to highlight the 'HI3208EN Volatile Acidity' method and press "*Select*".
- Install the 25-mL burette with 0.1N sodium hydroxide (HI70456) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all air has been removed completely.
  For the determination of the exact concentration of the 0.1N sodium hydroxide, follow the HI0001EN 0.1N Sodium Hydroxide Titrant Concentration.

Electrode Device:
- Press *"Mode"* from the main screen and press *"pH"*.
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation:
- Use the class-A volumetric pipette to transfer exactly 10.00 mL of degassed sample to the distillation flask. Add 0.5 mL of 3% hydrogen peroxide (HI70432). Steam distill the sample until approximately 100 mL of distillate has been collected in the 150-mL glass beaker.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press *"Start".* The titrator will start the analysis.
- At the end of titration, when pH 8.20 is reached, 'titration completed' will appear with the volatile acidity concentration. The result is expressed in **g/L of acetic acid**.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Volatile Acidity in Wine

Method Parameters:

Name:	Volatile Acidity
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.010 mL
max Vol:	0.100 mL
delta E:	6.000 mV
End Point Mode:	Fixed 8.200 pH
Pre-Titration Volume:	0.000 mL
Pre-Titration Stir Time:	20 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	1.6 Sec
t-min wait:	2 Sec
t-max wait:	20 Sec
Electrode Type:	pH
Blank Option:	No Blank
Calculations:	Sample Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.01N NaOH
Titrant Conc.:	1.0000E-2 N (eq/L)
Analyte Size:	10.000 mL
Analyte Entry:	Fixed
Maximum Titrant Volume:	25.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	3 Readings
Final Result Format:	XXX

Calculations:

Calculations:	Sample Calc. by Volume
Titrant units:	N (eq/L)
Titrant volume dosed:	V L)
Final result units:	g/L
Titrant conc.:	1.0000E-2 eq/L
Sample/Titrant:	1.000 mol/eq
MW of sample:	60.050 g/mol
Sample volume:	10.000 m

L

$$g/L = \frac{V(L)*1.0000E-2 *1.0*60.05*1000}{10.000}$$

Results:

Titration Report

Method Name:	Volatile Acid
Time & Date:	12:43 Mar 28, 2018
Titration ID:	Ti_00009

Titration Results

Method Name:	Volatile Acid
Time & Date:	12:43 Mar 28, 2018
Analyte size:	10.000 mL
End Point Volume:	0.402 mL
pH Fixed End Point:	8.200
Results:	0.024 g/L
Initial and Final pH:	5.534 to 8.204
Titration Duration:	9:12 [mm:ss]

Titration went to Completion

Operator name: _____

Free Sulfur Dioxide

Orienting Ripper Method

Description:
Method for the determination of free sulfur dioxide (SO_2) in wine, following the Orienting Ripper Method. The result is expressed in **ppm (mg/L) of sulfur dioxide**.

Reference:
Wine Analysis and Production Sulfur Dioxide: Ripper Titrametric Method Using Iodine

Electrode:
- HI3131B Combination ORP electrode

Primary Reagents:
- HI70440 0.02N Stabilized Iodine (1 L)
- HI70444 25% Sulfuric Acid (500 mL)
- HI70404 KI Powder Packets (100 pcs)
- HI70436 Deionized Water (1 gal)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Filling Solution (30 mL x 4)
- HI740036 100-mL Plastic Beakers (10 pcs)
- 50-mL Class-A Pipette

Device Preparation:
- Connect the ORP electrode to the titrator.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight the 'HI3213EN Free Sulfur Dioxide' method and press *"Select".*
- Install a 25-mL burette with 0.02N iodine (HI70440) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
  For the determination of the exact concentration of the 0.02N iodine, follow HI0204EN 0.02N Iodine Titrant Concentration.

Electrode Preparation:
- Prepare the ORP electrode according to the procedure in the manual.

Sample Preparation:
- Use a class-A glass pipette to transfer exactly 50.00 mL of wine to a clean 100-ml plastic beaker.
- Add 5 to 7 mL of 25% sulfuric acid (HI70444) and one HI70404 powder packet.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5-6 mm below the surface.

  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press *"Start".* The titrator will start the analysis.
- At the end of titration, when the equivalence point is reached, 'titration complete' will appear with the sulfur dioxide concentration. The result is expressed in **ppm (mg/L) of sulfur dioxide.**
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Free Sulfur Dioxide

Orienting Ripper Method

Method Parameters:

Name:	Free Sulfur Dioxide
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.015 mL
max Vol:	0.200 mL
delta E:	0.800 mV
End Point Mode:	mV 1EQ point, 1st Der
Recognition Options:	
Threshold:	200 mV/mL
Range:	NO
Filtered Derivatives:	NO
Pre-Titration Volume:	0.000 mL
Pre-Titration Stir Time:	30 Sec
Measurement Mode:	Signal Stability
delta E:	1.0 mV
delta t:	3.0 Sec
t-min wait:	5 Sec
t-max wait:	45 Sec
Electrode Type:	ORP
Blank Option:	No Blank
Calculations:	Sample Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.02N Iodine
Titrant Conc.:	2.0000E-2 N (eq/L)
Analyte Size:	50.000 mL
Analyte Entry:	Fixed
Maximum Titrant Volume:	25.000 mL
Stirring Speed:	1400 rpm
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	2 Readings
Final Result Format:	XXXX

Calculations:

Calculations:	Sample Calc. by Volume
Titrant units:	N (eq/L)
Titrant volume dosed:	V L)
Final result units:	ppm (mg/L)
Titrant conc.:	2.000E-2 eq/L
Sample/Titrant:	0.500 mol/eq
MW of sample:	64.063 g/mol
Sample volume:	50.000 mL

$$ppm = \frac{V(L)*1000*0.02*0.5*64.063}{0.050}$$

Results:

Titration Report

Method Name:	Free Sulfur Dioxide
Time & Date:	13:44 Feb 16, 2017
Titration ID:	Ti_00013

Titration Results

Method Name: Report

Method Name:	Free Sulfur Dioxide
Time & Date:	13:44 Feb 16, 2017
Analyte size:	50.000 mL
End Point Volume:	1.225 mL
pH Equivalence Point:	254.4
Results:	15.36 ppm (mg/L)
Initial and Final mV:	2361.1 to 315.3
Titration Duration:	1:39 [mm:ss]

Titration went to Completion

Operator name: _____

Total Sulfur Dioxide

Orienting Ripper Method

Description:
Method for the determination of total sulfur dioxide (SO_2) in wine, following the Orienting Ripper Method. The result is expressed in **ppm (mg/L) of sulfur dioxide**.

Reference:
Wine Analysis and Production Sulfur Dioxide: Ripper Titrametric Method Using Iodine

Electrode:
- HI3131B Combination ORP electrode

Primary Reagents:
- HI70440 0.02N Stabilized Iodine (1 L)
- HI70435 5M Sodium Hydroxide (500 mL)
- HI70444 25% Sulfuric Acid (500 mL)
- HI70404 KI Powder Packets (100 pcs)
- HI70436 Deionized Water (1 gal)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Filling Solution (30 mL x 4)
- HI740036 100-mL Plastic Beakers (10 pcs)
- 50-mL Class-A Pipette

Device Preparation:
- Connect the ORP electrode to the titrator.
- Press "*Select Method*" from the main screen. Use the arrow keys to highlight the 'HI3216EN Total Sulfur Dioxide' method and press "*Select*".
- Install a 25-mL burette with 0.02N iodine (HI70440) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
  For the determination of the exact concentration of the 0.02N iodine, follow HI0204EN 0.02N Iodine Titrant Concentration.

Electrode Preparation:
- Prepare the ORP electrode according to the procedure in the manual.

Sample Preparation:
- Use a class-A glass pipette to transfer exactly 50.00 mL of wine to a clean 100-ml plastic beaker.
- Add 5 mL of 5M sodium hydroxide (HI70435). Cover the beaker and swirl. Allow the sample to sit for approximately 20 minutes.
- Add 10 mL of 25% sulfuric acid (HI70444) and one HI70404 powder packet.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5-6 mm below the surface.
- **NOTE:** The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press "*Start*". The titrator will start the analysis.
- At the end of titration, when the equivalence point is reached, 'titration complete' will appear with the sulfur dioxide concentration. The result is expressed in **ppm (mg/L) of sulfur dioxide.**
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Total Sulfur Dioxide

Orienting Ripper Method

Method Parameters:

Name:	Total Sulfur Dioxide
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.020 mL
max Vol:	0.400 mL
delta E:	2.500 mV
End Point Mode:	mV 1EQ point, 1st Der
Recognition Options:	
Threshold:	50 mV/mL
Range:	NO
Filtered Derivatives:	YES
Pre-Titration Volume:	0.000 mL
Pre-Titration Stir Time:	30 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	2.0 Sec
t-min wait:	2 Sec
t-max wait:	20 Sec
Electrode Type:	ORP
Blank Option:	No Blank
Calculations:	Sample Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.02N Iodine
Titrant Conc.:	2.0000E-2 N (eq/L)
Analyte Size:	50.000 mL
Analyte Entry:	Fixed
Maximum Titrant Volume:	25.000 mL
Stirring Speed:	1400 rpm
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	3 Readings
Final Result Format:	XXXX

Calculations:

Calculations:	Sample Calc. by Volume
Titrant units:	N (eq/L)
Titrant volume dosed:	V L)
Final result units:	ppm (mg/L)
Titrant conc.:	2.000E-2 eq/L
Sample/Titrant:	0.500 mol/eq
MW of sample:	64.063 g/mol
Sample volume:	50.000 mL

$$ppm = \frac{V(L)*1000*0.02*0.5*64.063}{0.050}$$

Results:

Titration Report

Method Name:	Total Sulfur Dioxide
Time & Date:	14:40 Feb 20, 2016
Titration ID:	Ti_00123

Titration Results

Method Name:	Total Sulfur Dioxide
Time & Date:	14:40 Feb 20, 2016
Analyte size:	50.000 mL
End Point Volume:	4.290 mL
pH Equivalence Point:	261.6
Results:	54.40 ppm (mg/L)
Initial and Final mV:	253.7 to 281.8
Titration Duration:	4:53 [mm:ss]

Titration went to Completion

Operator name: _____

Reducing Sugar - Blank

Description:
Method for the determination of reducing sugar in wine. An excess of copper(II) is added to a sample and upon heating, the reducing sugars present in the sample reduce the copper(II) to copper(I). Potassium iodide is added to the sample and reacts with the remaining excess copper(II) to form iodine. The iodine is titrated with sodium thiosulfate.
A blank titration is performed to quantify and correct for the other constituents in wine that react with the Fehling reagent and therefore, decrease copper.
The reduction of copper(II) is neither stoichiometric nor linear, thus a calibration factor must be determined to standardize the correlation between copper(II) and reducing sugar.
This analysis involves a three-part procedure to determine the blank, the calibration factor, and finally the concentration of reducing sugars. This method, HI3217EN, is used to determine the blank value. The result is expressed as **L of titrant**.

Reference:
Zoecklein, et al. Wine Analysis and Production. Reducing Sugars: Titrametric-Rebelein (Gold Coast) Method

Electrode:
- HI3131B Combination ORP Electrode

Reagents:
- HI70439 0.1M Sodium Thiosulfate (1 L)
- HI70446 Fehling A (500 mL)
- HI70447 Fehling B (500 mL)
- HI70425 16% Sulfuric Acid (500 mL)
- HI70437 Concentrated KI (500 mL)
- HI70436 Deionized Water (1 gal)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI731342 2000µL Automatic Pipette
- HI731352 2000µL Automatic Pipette Tips (4pcs)
- 150-mL Glass Beaker
- 5-mL Class-A pipette
- 10-mL Class-A pipette
- Hot Plate

Device Preparation:
- Connect the ORP electrode to the titrator.
- Press "*Select Method*" from the main screen. Use the arrow keys to highlight 'HI3217EN Reducing Sugar - Blank' method and press "*Select*".
- Install a 25-mL burette with 0.1M sodium thiosulfate (HI70439) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.

  For the determination of the exact concentration of the 0.1M sodium thiosulfate, follow HI0003EN 0.1M Sodium Thiosulfate Titrant Concentration.

Electrode Preparation:
- Prepare the ORP electrode according to the procedure in the manual.

Sample Preparation:
- Add 2000 µL of deionized water to a 150-mL glass beaker using the automatic pipette.
- Use a class-A volumetric pipette to transfer exactly 5.00 mL of Fehling A (HI70446) and exactly 5.00 mL of Fehling B (HI70447) to the 150-mL glass beaker
- Heat the mixture to a boil for approximately 2 minutes.
- Cool the solution to room temperature by running the beaker under tap water or submersing the bottom of the beaker in a water bath.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of concentrated potassium iodide (HI70437) to the 150-mL glass beaker.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of 16% sulfuric acid (HI70425) to the 150-mL glass beaker.
- Add approximately 70 mL of deionized water to bring the total volume to 100 mL.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press "*Start*". The titrator will start the analysis.
- At the end of titration, when the equivalence point is reached, 'titration complete' will appear with the result. The result is expressed in **L of titrant**
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Reducing Sugar - Blank

Method Parameters:

```
Name:                  Reducing Sugar - Blank
Method Revision:                         3.0
Stirrer Configuration:
     Stirrer:                      Stirrer 1
     Stirring Speed:                1400 RPM
Pump Configuration:
     Titrant Pump:                     Pump 1
Dosing Type:                         Dynamic
     min Vol:                      0.020 mL
     max Vol:                      0.500 mL
     delta E:                      6.500 mV
End Point Mode:       mV 1 EQ Point, 1st Der
Recognition Options:
     Threshold:                    75 mV/mL
     Range:                              NO
     Filtered Derivatives:              YES
Pre-Titration Volume:              10.000 mL
Pre-Titration Stir Time:              30 Sec
Measurement Mode:           Signal Stability
     delta E:                       0.5 mV
     delta t:                      2.5 Sec
     t-min wait:                     3 Sec
     t-max wait:                    20 Sec
Electrode Type:                         ORP
Blank Option:                      No Blank
Calculations:            No Formula (L only)
Titrant Name:                  0.1M Na2S2O3
Maximum Titrant Volume:            20.000 mL
Potential Range:       -2000.0 to 2000.0 mV
Volume/Flow Rate:           25 mL/50 mL/min
Signal Averaging:                2 Readings
Final Result Format:                   XXXX
```

Calculations:

Final Results unit in L

V = volume dispensed in liters

Results:

```
            Titration Report
Method Name:          Reducing Sugar - Blank
Time & Date:               9:52 Dec 20, 2016
Titration ID:                       Ti_00048

            Titration Results
Method Name:          Reducing Sugar - Blank
Time & Date:               9:52 Dec 20, 2016
End Point Volume:                 0.01228 L
mV Equivalence Point:                 221.2
Results:                          0.01228 L
Initial and Final mV:         353.1 to 174.4
Titration Duration:             3:32 [mm:ss]
Titration went to Completion
Operator name: _____________________________
```

Reducing Sugar - Calibration Factor

Description:
Method for the determination of reducing sugar in wine. An excess of copper(II) is added to a sample and upon heating, the reducing sugars present in the sample reduce the copper(II) to copper(I). Potassium iodide is added to the sample and reacts with the remaining excess copper(II) to form iodine. The iodine is titrated with sodium thiosulfate.
A blank titration is performed to quantify and correct for the other constituents in wine that react with the Fehling reagent and therefore, decrease copper.
The reduction of copper(II) is neither stoichiometric nor linear, thus a calibration factor must be determined to standardize the correlation between copper(II) and reducing sugar.
This analysis involves a three-part procedure to determine the blank, the calibration factor, and finally the concentration of reducing sugars. This method, HI3218EN, is used to determine the calibration factor. The result is expressed as **g/L of reducing sugar**.

Reference:
Zoecklein, et al. Wine Analysis and Production.
Reducing Sugars: Titrametric-Rebelein (Gold Coast) Method

Electrode:
- HI3131B Combination ORP Electrode

Reagents:
- HI70439 0.1M Sodium Thiosulfate (1 L)
- HI70446 Fehling A (500 mL)
- HI70447 Fehling B (500 mL)
- HI70425 16% Sulfuric Acid (500 mL)
- HI70437 Concentrated KI (500 mL)
- HI70436 Deionized Water (1 gal)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI731342 2000µL Automatic Pipette
- HI731352 2000µL Automatic Pipette Tips (4pcs)
- 150-mL Glass Beaker
- 100-mL Class-A Volumetric Flask
- 5-mL Class-A Pipette
- 10-mL Class-A Pipette
- Hot Plate
- Analytical Balance with 0.0001g resolution
- 10 g Glucose Standard
- 20 g Glucose Standard

Device Preparation:
- Connect the ORP electrode to the titrator.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight 'HI3218EN Reducing Sugar – Cal.' method and press *"Select"*.
- Install a 25-mL burette with 0.1M sodium thiosulfate (HI70439) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
  For the determination of the exact concentration of the 0.1M sodium thiosulfate, follow HI0003EN 0.1M Sodium Thiosulfate Titrant Concentration.
- Update the blank value. Press *"Method Options"* from the main screen. Use the arrow keys to highlight 'Blank Value' and press *"Select"*. Use the numeric keypad to enter the blank value obtained from HI3217 Reducing Sugar – Blank and press *"Accept"*. Press *"Escape"* to exit the Method Options screen and select 'Save Method' option.

Electrode Preparation:
- Prepare the ORP electrode according to the procedure in the manual.

Glucose Standard Preparation:
- 10 g Glucose Standard – Weigh 1.0 g of glucose, transfer to 100-mL class-A volumetric flask. Add deionized water to dissolve, bring to volume, cap and mix. This solution is not stable and should not be made in advance.
- 20 g Glucose Standard – Weigh 2.0 g of glucose, transfer to 100-mL class-A volumetric flask. Add deionized water to dissolve, bring to volume, cap and mix. This solution is not stable and should not be made in advance.

Sample Preparation:
- Add 2000µL of 10 g/L glucose standard to a 150-mL glass beaker using the automatic pipette.
- Use a class-A volumetric pipette to transfer exactly 5.00 mL of Fehling A (HI70446) and exactly 5.00 mL of Fehling B (HI70447) to the 150-mL glass beaker
- Heat the mixture to a boil for approximately 2 minutes.
- Cool the solution to room temperature by running the beaker under tap water or submersing the bottom of the beaker in a water bath.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of concentrated potassium iodide (HI70437) to the 150-mL beaker.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of 16% sulfuric acid (HI70425) to the 150-mL beaker.
- Add approximately 70 mL of deionized water to bring the total volume to 100 mL.
- Repeat this procedure with the 20 g/L glucose standard.

Reducing Sugar - Calibration Factor

Analysis:

- Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press "*Start*". The titrator will start the analysis.
- At the end of titration, when the equivalence point is reached, 'titration complete' will appear with the result. The result is expressed as **g/L reducing sugar.**
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.
- Repeat this procedure with the 20 g/L glucose standard.

Calibration Factor:

The Calibration factor is determined by dividing concentration of the standard by the titrated value. These values are then added together and divided by 2.

Example:

10 g/L Glucose Standard titrated as 11.24 g/L:

$$\frac{10.00}{11.24}=0.8897$$

20 g/L Glucose Standard titrated as 21.74 g/L:

$$\frac{20.00}{21.74}=0.9199$$

Calibration Factor:

$$\frac{(0.8897+0.9199)}{2}=0.9048$$

Reducing Sugar - Calibration Factor

Method Parameters:

Name:	Reducing Sugar – Cal.
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.020 mL
max Vol:	0.500 mL
delta E:	6.500 mV
End Point Mode:	mV 1 EQ Point, 1st Der
Recognition Options:	
Threshold:	75 mV/mL
Range:	NO
Filtered Derivatives:	YES
Pre-Titration Volume:	0.000 mL
Pre-Titration Stir Time:	30 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	2.5 Sec
t-min wait:	3 Sec
t-max wait:	20 Sec
Electrode Type:	ORP
Blank Option:	Blank – V
Blank Value:	1.2500E-2 L
Calculations:	Generic Formula
Dilution Option:	Disabled
Titrant Name:	0.1M Na2S2O3
Titrant Conc.:	0.1000 C
Analyte Size:	2.0000 mL
Analyte Entry:	Fixed
Maximum Titrant Volume:	20.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	2 Readings
Final Result Format:	XXXX

Calculations:

Calculations:	Generic Formula
Titrant volume dosed:	V (L)
Final result units:	g/L
Titrant conc.:	0.100 C
Blank Volume:	1.250E-2 L
Factor 1:	36.000
Factor 2:	1.0000E3
Factor 3:	1.000
Sample size:	2.000

$$g/L = \frac{(1.250E\text{-}2\text{-}V(L))*0.10*36.0*1.0E3*1.0}{2.000}$$

Results (10 g/L Glucose Standard):

Titration Report

Method Name:	Reducing Sugar – Cal.
Time & Date:	11:39 Dec 20, 2016
Titration ID:	Ti_00037

Titration Results

Method Name:	Reducing Sugar – Cal.
Time & Date:	11:39 Dec 20, 2016
Analyte Size:	2.000 mL
End Point Volume:	6.792 mL
mV Equivalence Point:	205.1
Results:	21.74 g/L
Initial and Final mV:	332.2 to 168.9
Titration Duration:	6:45 [mm:ss]

Titration went to Completion

Operator name: _____

Results (20 g/L Glucose Standard):

Titration Report

Method Name:	Reducing Sugar – Cal.
Time & Date:	12:04 Dec 20, 2016
Titration ID:	Ti_00053

Titration Results

Method Name:	Reducing Sugar – Cal.
Time & Date:	12:04 Dec 20, 2016
Analyte Size:	2.000 mL
End Point Volume:	0.959 mL
mV Equivalence Point:	217.5
Results:	21.74 g/L
Initial and Final mV:	312.8 to 170.3
Titration Duration:	3:28 [mm:ss]

Titration went to Completion

Operator name: _____

Reducing Sugar in Wine

Description:
Method for the determination of reducing sugar in wine. An excess of copper(II) is added to a sample and upon heating, the reducing sugars present in the sample reduce the copper(II) to copper(I). Potassium iodide is added to the sample and reacts with the remaining excess copper(II) to form iodine. The iodine is titrated with sodium thiosulfate.
A blank titration is performed to quantify and correct for the other constituents in wine that react with the Fehling reagent and therefore, decrease copper.
The reduction of copper(II) is neither stoichiometric nor linear, thus a calibration factor must be determined to standardize the correlation between copper(II) and reducing sugar.
This analysis involves a three-part procedure to determine the blank, the calibration factor, and finally the concentration of reducing sugars. This method, HI3219EN, is used to determine the reducing sugar in wine. The result is expressed as **g/L of reducing sugar**.

Reference:
Zoecklein, et al. Wine Analysis and Production.
Reducing Sugars: Titrametric-Rebelein (Gold Coast) Method

Electrode:
- HI3131B Combination ORP Electrode

Reagents:
- HI70439 0.1M Sodium Thiosulfate (1 L)
- HI70446 Fehling A (500 mL)
- HI70447 Fehling B (500 mL)
- HI70425 16% Sulfuric Acid (500 mL)
- HI70437 Concentrated KI (500 mL)
- HI70436 Deionized Water (1 gal)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI731342 2000µL Automatic Pipette
- HI731352 2000µL Automatic Pipette Tips (4pcs)
- 150-mL Glass Beaker
- 5-mL Class-A Pipette
- 10-mL Class-A Pipette
- Hot Plate

Device Preparation:
- Connect the ORP electrode to the titrator.
- Press "*Select Method*" from the main screen. Use the arrow keys to highlight 'HI3219EN Reducing Sugar' method and press "*Select*".
- Install a 25-mL burette with 0.1M sodium thiosulfate (HI70439) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
  For the determination of the exact concentration of the 0.1M sodium thiosulfate, follow HI0003EN 0.1M Sodium Thiosulfate Titrant Concentration.
- Update the blank value. Press "*Method Options*" from the main screen. Use the arrow keys to highlight 'Blank Value' and press "*Select*". Use the numeric keypad to enter the blank value obtained from HI3217 Reducing Sugar – Blank and press "*Accept*".
- Update the calibration value. Press "*Method Options*" from the main screen. Use the arrow keys to highlight 'Calculations' and press "*Select*". Use the arrow keys to highlight 'Edit Variable Values' and press "*Select*". Use the arrow keys to highlight 'F1 -> General Factor' and press "*Select*". Use the numeric keypad to enter the calibration value obtained from HI3218 Reducing Sugar – Cal. and press "*Accept*". Press "*Escape*" to exit the Method Options screen and select 'Save Method' option.

Electrode Preparation:
- Prepare the ORP electrode according to the procedure in the manual.

Sample Preparation:
- Add 2000 µL of wine to a 150-mL glass beaker using the automatic pipette.
- Use a class-A volumetric pipette to transfer exactly 5.00 mL of Fehling A (HI70446) and exactly 5.00 mL of Fehling B (HI70447) to the 150-mL glass beaker.
- Heat the mixture to a boil for approximately 2 minutes.
- Cool the solution to room temperature by running the beaker under tap water or submersing the bottom of the beaker in a water bath.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of concentrated potassium iodide (HI70437) to the 150-mL glass beaker.
- Use a class-A volumetric pipette to transfer exactly 10.00 mL of 16% sulfuric acid (HI70425) to the 150-mL glass beaker.
- Add approximately 70 mL of deionized water to bring the total volume to 100 mL.

Reducing Sugar in Wine

Analysis:

- Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5-6 mm below the surface. If necessary add extra deionized water.
  NOTE: The dispensing tip should be in contact with the surface of the sample (slightly submerged).
- Press "*Start*". The titrator will start the analysis.
- At the end of titration, when the equivalence point is reached, 'titration complete' will appear with the result. The result is expressed as **g/L residual sugar.**
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Reducing Sugar in Wine

Method Parameters:

Name:	Reducing Sugar
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.020 mL
max Vol:	0.600 mL
delta E:	6.500 mV
End Point Mode:	mV 1 EQ Point, 1st Der
Recognition Options:	
Threshold:	75 mV/mL
Range:	NO
Filtered Derivatives:	YES
Pre-Titration Volume:	0.000 mL
Pre-Titration Stir Time:	30 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	2.5 Sec
t-min wait:	3 Sec
t-max wait:	20 Sec
Electrode Type:	ORP
Blank Option:	Blank – V
Blank Value:	1.250E-2 L
Calculations:	Generic Formula
Dilution Option:	Disabled
Titrant Name:	0.1M Na2S2O3
Titrant Conc.:	0.1000 C
Analyte Size:	2.0000 mL
Analyte Entry:	Fixed
Maximum Titrant Volume:	20.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	2 Readings
Final Result Format:	XXXX

Calculations:

Calculations:	Generic Formula
Titrant volume dosed:	V (L)
Final result units:	g/L
Titrant conc.:	0.100 C
Blank Volume:	1.250E-2 L
Factor 1:	0.924
Factor 2:	36.000
Factor 3:	1.0000E3
Sample size:	2.000E-3

$$g/L = \frac{(1.250E\text{-}2 - V(L)) * 0.1 * 0.924 * 36.0 * 1.0E3}{2.000E\text{-}3}$$

Results:

Titration Report

Method Name:	Reducing Sugar
Time & Date:	14:55 Dec 22, 2016
Titration ID:	Ti_00001

Titration Results

Method Name:	Reducing Sugar
Time & Date:	14:55 Dec 22, 2016
Analyte Size:	2.000 mL
End Point Volume:	1.025 mL
mV Equivalence Point:	218.2
Results:	19.81 g/L
Initial and Final mV:	317.9 to 166.2
Titration Duration:	3:33 [mm:ss]

Titration went to Completion

Operator name: _____

Formol Number – pH Adjustment

Description:
Method for the determination of assimilable nitrogen in wine. This method provides an approximate value that can be used as an index of the must nutritional value. This analysis is a two-part procedure: the first is a pH adjustment and the second is the concentration of fermentable nitrogen. The result is expressed as **mL of titrant.**

Reference:
Wine Analysis and Production Formol Number (Nitrogen) in Wine

Electrode:
- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

Reagents:
- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70457 1.0N Sodium Hydroxide (1 L)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- 100-mL Class-A Pipette
- 250-mL Glass Beaker

Device Preparation:
- Connect the pH electrode and temperature probe to the titrator.
- Press *"Select Method"* from the main screen. Use the arrow keys to highlight 'HI3230EN Formol Number - pH adj.' method and press *"Select"*.
- Install a 25-mL burette with 0.1N sodium hydroxide (HI70456) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
  For the determination of the exact concentration of the 0.1N sodium hydroxide, follow HI0001EN 0.1N Sodium Hydroxide Titrant Concentration.

Electrode Preparation:
- Press *"Mode"* from the main screen and press *"pH".*
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation:
- Use a class-A volumetric pipette to transfer exactly 100.00 mL of wine to a clean 250-mL glass beaker. Rinse the pipette into the beaker with approximately 50 mL of deionized water. The total volume of should be roughly 150 mL.
- Add approximately 6.0 mL of 1N sodium hydroxide (HI70457).

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5-6 mm below the surface.
- Press *"Start"*. The titrator will start the analysis.
- At the end of titration, when pH 8.00 is reached, 'titration complete' will appear with the results. The result is expressed in **mL of titrant.**
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Save the pH adjusted sample and continue with 'HI3231EN Formol Number'

Formol Number – pH Adjustment

Method Parameters:

Name:	Formol Number – pH adj.
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1200 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.010 mL
max Vol:	0.300 mL
delta E:	8.000 mV
End Point Mode:	Fixed 8.000 pH
Pre-Titration Volume:	0.000 mL
Pre-Titration Stir Time:	30 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	1.0 Sec
t-min wait:	2 Sec
t-max wait:	20 Sec
Electrode Type:	pH
Blank Option:	No Blank
Calculations:	No Formula (mL only)
Dilution Option:	Disabled
Titrant Name:	0.1N NaOH
Maximum Titrant Volume:	25.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	3 Readings
Final Result Format:	XXXX

Calculations:

Final Results unit in mL

mL = V * 1000

V = volume dispensed in liters

Results:

Titration Report

Method Name:	Formol Number – pH adj.
Time & Date:	16:17 Feb 22, 2017
Titration ID:	Ti_00060

Titration Results

Method Name:	Formol Number – pH adj.
Time & Date:	16:17 Feb 22, 2017
End Point Volume:	13.265 mL
pH Fixed End Point:	8.000
Results:	13.265 mL
Initial and Final pH:	5.491 to 8.000
Titration Duration:	5:04 [mm:ss]

Titration went to Completion

Operator name: _____

Formol Number

Description:
Method for the determination of assimilable nitrogen in wine. This method provides an approximate value that can be used as an index of the must nutritional value. This analysis is a two-part procedure: the first is a pH adjustment and the second is the concentration of fermentable nitrogen. The result is expressed as **ppm (mg/L) of fermentable nitrogen.**

Reference:
Wine Analysis and Production Formol Number (Nitrogen) in Wine

Electrode:
- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

Reagents:
- HI70456 0.1N Sodium Hydroxide (1 L)
- HIFORM 37% Formaldehyde (500 mL)

Other Accessories:
- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- 25-mL Class-A Pipette
- 100-mL Class-A Pipette
- 250-mL Glass Beaker
- 200-mL Volumetric Flask

Device Preparation:
- Connect the pH electrode and temperature probe to the titrator.
- Press "*Select Method*" from the main screen. Use the arrow keys to highlight 'HI3230EN Formol Number' method and press "*Select*".
- Install a 25-mL burette with 0.1N sodium hydroxide (HI70456) on pump-one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
  For the determination of the exact concentration of the 0.1N sodium hydroxide, follow HI0001EN 0.1N Sodium Hydroxide Titrant Concentration.

Electrode Preparation:
- Press *"Mode"* from the main screen and press *"pH"*.
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

Sample Preparation:
- Use the pH adjusted sample from HI3230 Formol Number – pH adjustment.
- Transfer the pH adjusted sample to a 200-mL class-A volumetric flask. Bring the sample up to volume with deionized water, cap and mix well.
- Use a class-A volumetric pipette to transfer exactly 100.00 mL of sample to a clean 250-mL glass beaker.
- Use a class-A volumetric pipette to transfer exactly 25.00 mL of pH adjusted 37% formaldehyde to the beaker.
  NOTE: The formaldehyde should be adjusted or readjusted to pH 8.00, or to the specified endpoint before it is added to the sample. If no drop in pH is observed after the formaldehyde is added, the sample does not contain nitrogen in a quantifiable amount.

Analysis:
- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5-6 mm below the surface.
- Press "*Start*". The titrator will start the analysis.
- At the end of titration, when pH 8.00 is reached, 'titration complete' will appear with the results. The result is expressed in **ppm (mg/L) of fermentable nitrogen.**
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Formol Number

Method Parameters:

Name:	Formol Number
Method Revision:	3.0
Stirrer Configuration:	
Stirrer:	Stirrer 1
Stirring Speed:	1200 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Dosing Type:	Dynamic
min Vol:	0.010 mL
max Vol:	0.250 mL
delta E:	8.000 mV
End Point Mode:	Fixed 8.000 pH
Pre-Titration Volume:	0.000 mL
Pre-Titration Stir Time:	20 Sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	1.0 Sec
t-min wait:	2 Sec
t-max wait:	20 Sec
Electrode Type:	pH
Blank Option:	No Blank
Calculations:	Sample Calc. by Volume
Dilution Option:	Enabled
Final Dilution Volume:	200.000 mL
Aliquot Volume:	100.000 mL
Titrant Name:	0.1N NaOH
Titrant Conc.:	0.1000 N (eq/L)
Analyte Size:	100.00 mL
Analyte Entry:	Fixed
Maximum Titrant Volume:	25.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50 mL/min
Signal Averaging:	3 Readings
Final Result Format:	XXXXX

Calculations:

Calculations:	Sample Calc. by Volume
Titrant units:	N (eq/L)
Titrant volume dosed:	V (L)
Final result units:	ppm (mg/L)
Titrant conc.:	0.100 N (eq/L)
Sample/Titrant:	1.000 mol/eq
MW of sample:	14.016 g/mol
Sample volume:	100.00 mL

$$ppm = \frac{V(L)*0.100*1.000*14.016*1000}{100.00}$$

Results:

Titration Report

Method Name:	Formol Number
Time & Date:	15:49 Jan 30, 2017
Titration ID:	Ti_00077

Titration Results

Method Name:	Formol Number
Time & Date:	15:49 Jan 30, 2017
Analyte size:	100.00 mL
End Point Volume:	4.792 mL
pH Fixed End Point:	8.000
Results:	288 ppm (mg/L)
Initial and Final pH:	6.562 to 8.006
Titration Duration:	3:32 [mm:ss]

Titration went to Completion

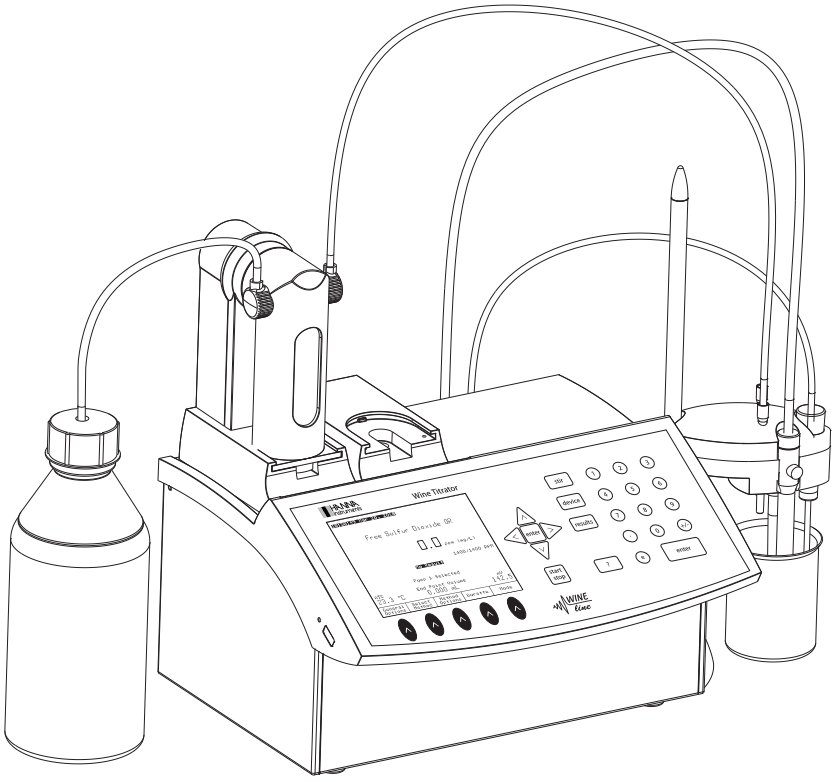
Operator name: _____

General Titration Applications Brochure

HI901 Wine

AUTOMATIC POTENTIOMETRIC TITRATOR

Revision 1.00



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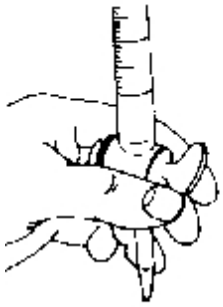
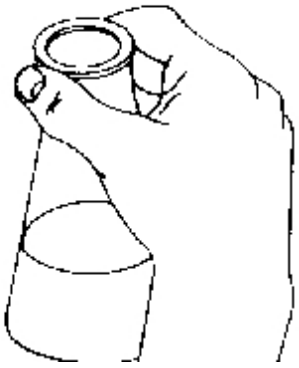
TITRATION THEORY

HI 901 and HI 902

AUTOMATIC POTENTIOMETRIC TITRATOR





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Contents

1 GENERAL REVIEW OF TITRATION THEORY

1.1 Introduction to Titrations

A titration is a quantitative, volumetric procedure used in analytical chemistry to determine the concentration of an analyte (the species being measured) in solution. The concentration of the analyte is determined by slowly adding a titrant (reagent) to the solution. As the titrant is added, a chemical reaction occurs between the titrant and the analyte.

Titration reactions are relatively fast, simple reactions that can be expressed using a chemical equation. The titration reaction continues as the titrant is added until all of the analyte is consumed and the analyte reacts completely and quantitatively with the titrant.

The point at which all of the analyte has been reacted is called the equivalence point, also known as the theoretical or stoichiometric endpoint. This point is accompanied by an abrupt physical change in the solution, which sharply defines the endpoint of the reaction. The physical change associated with the titration endpoint can be produced by the titrant or an indicator and can be detected either visually or by some other physical measurement.

Titrations cannot be used to determine the quantity of all analytes. The chemical reaction between the titrant and analyte must fulfill four requirements:

- The reaction must be fast and occur within approximately one second after the titrant is added
- The reaction must go to completion
- The reaction must have well-known stoichiometry (reaction ratios)
- A convenient endpoint or inflection point

Titrations are highly precise and can provide many advantages over alternative methods. Titrations are quickly performed and require relatively simple apparatus and instrumentation.

1.2 Uses of Titrations

Titrations can be used in many applications, including:

- Acid content of plant effluents, food (e.g.: cheese and wine), plating and etching baths, petroleum products, drugs
- Base content of fertilizer (containing ammonia), bleach, minerals
- Hardness in water
- Metal content of alloys, minerals, ores, clays, waters, plating baths, paints, paper, plant materials, biological fluids, petroleum products
- Moisture content in foodstuffs, petrochemicals, pharmaceutical products, and plastics
- Redox reagent concentrations such as available chlorine in potable water, peroxide, traces of oxidants and reductants in food, reductants in high temperature or high pressure boiler water, vitamin analysis

1.3 Advantages and Disadvantages of Titrations

Some advantages of titrations as an analytical technique are:

- More precise results than many instrumental methods, such as measurement by electrode, the accuracy of the measurement is up to 0.1%
- Simple methods, reasonable capital costs, and easy training
- Suitability to measure major components of a mixture or product
- Automation can reduce time and labor spent on each analysis

Some disadvantages of titrations are:

- Time it takes to prepare standards and titrants
- Good technique is required to achieve precise results (training and practice required)
- Not suitable for determining trace or minor components of a mixture or product
- Limited dynamic range, it may require additional sample preparations (dilution) and repeat analyses

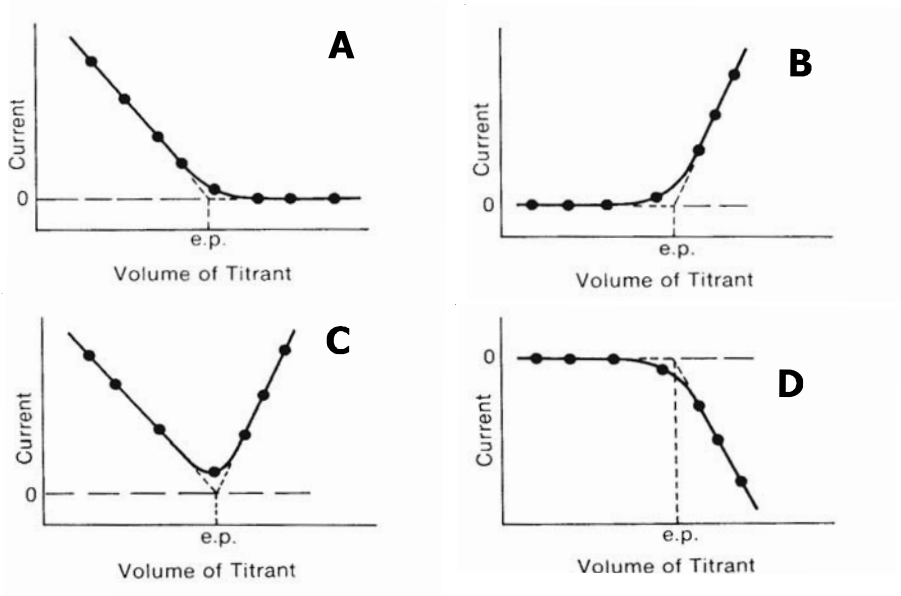
2 TYPES OF TITRATIONS

2.1 Titrations According to The Measurement Method

2.1.1 Amperometric Titrations

An amperometric titration is performed by placing two electrodes (often a metal ISE and a reference electrode) into the sample solution and holding the potential of the metal electrode at a selected voltage. The current that flows, due to the oxidation or reduction of a reactant or product, is plotted vs. volume of titrant to provide the titration curve and locate the equivalence point. Changes in the current are due to changes in the concentration of a particular species (being oxidized or reduced at the electrode).
Generally the reaction between the analyte and titrant forms a new species. Depending on the titration, the reactants are electroactive and the products are not, or vice-versa. Amperometric titration curves look like two straight lines intersecting at the equivalence point, this is due to the change in the electroactivity of the solution.
Many metal ions can be amperometrically titrated using a precipitation, complexation or redox reaction. Some metal ions and species that can be determined in this manner include silver, barium, halides, potassium, magnesium, palladium, molybdate, sulfate, tungstate, zinc, bismuth, cadmium, fluoride, indium, thallium, iodine, and gold.
Figure 1 shows four amperometric titrations and their endpoints. In graph “A” the analyte is electroactive and gives current but the reacted species does not. In “B” the reactant is not active but the titrant is. In “C” both the analyte and titrant are active and both give current flow. Graph “D” shows the same situation as “B”; however, the current has an opposite sign (the titrant is reduced).

Figure 1



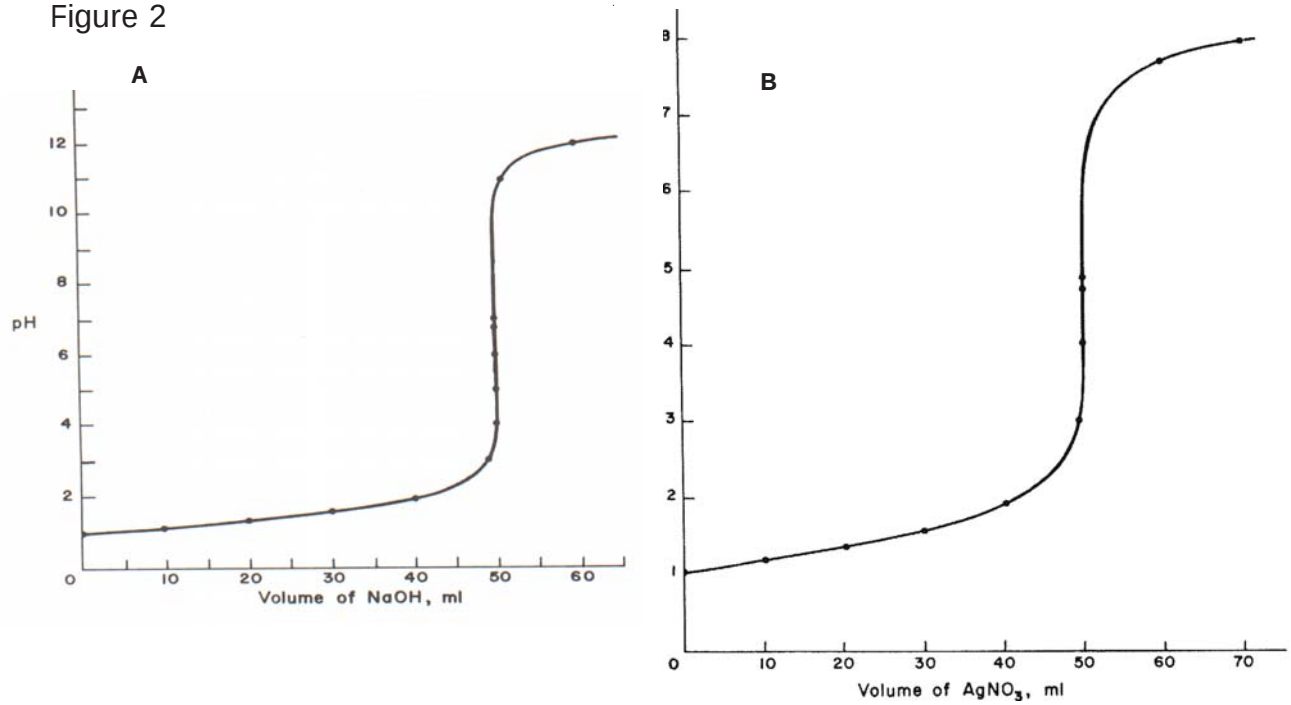
2.1.2 Potentiometric Titrations

Potentiometric titrations are done by measuring the voltage across the solution using an electrode system. An electrode system consists of an indicator electrode and a reference electrode. As titrant is added the variations in the potential of the indicator electrode, with respect to the reference electrode, are monitored to show the progress of the titration.
Potentiometry is the measurement of a potential under conditions of zero current flow. The

measured potential can then be used to determine the analytical quantity of interest, generally a component concentration of the analyte solution. The potential that develops in the electrochemical cell is the result of the free energy change that would occur if the chemical phenomena were to proceed until the equilibrium condition has been satisfied.

There are many types of titrations where potentiometry can be used,e.g., pH electrodes for acid-base titrations, platinum ORP electrodes in redox titrations, ion selective electrodes, such as chloride or fluoride for a specific ion titration, and silver electrodes for argentometric (silver-based) titrations. An example of potetiometric titrations are shown below. Figure 2 “A” is the pH of a solution vs. the volume of titrant and “B” is the potential from a chloride electrode vs. the volume of $AgNO_3$.

Figure 2



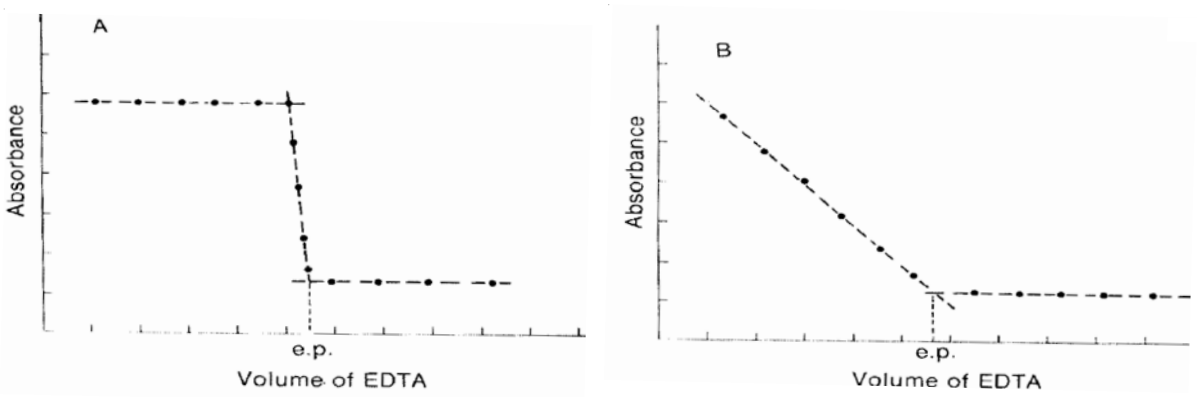
2.1.3 Spectrophotometric Titrations

The name comes from the method used to detect the endpoint of the titration, not its chemistry. Highly colored indicators that change color during the course of the titration are available for many titrations. More accurate data on the titration curve can be obtained if the light absorption is monitored instrumentally using a light source, a simple monochromator and a photodetector, rather than visually determining the color or light absorption change. Light absorption by either an indicator or by one of the reactants or products can be used to monitor the titration.

In the first titration curve, Figure 3 “A”, the absorption of a metal-indicator complex is being monitored. The absorption is constant while the metal is complexed by the EDTA titrant. The metal indicator complex was stripped, causing a sharp break in the titration curve. The point where all the metal is complexed and stripped from the indicator is the equivalence point. This point is marked by “e.p.” on the graph.

In the second titration curve, Figure 3 “B”, the metal complex is being measured while being titrated with EDTA. The new complex being formed is not colored and does not absorb light. The extrapolated intersection of the two lines determines the equivalence point.

Figure 3



2.2 Titrations According to The Reaction Type

2.2.1 Acid-Base Titrations

Acid–base titrations are the most common type of titrations. They are based upon a reaction between an acid and a base, a stoichiometric neutralization, or the exchange of protons. Virtually all acid-base titrations are carried out using a strong acid or a strong base as the titrant. The endpoint of a titration carried out with a weak acid or a weak base would be difficult to detect due to a small change in pH at the equivalence point.

Chemical indicators can be used to determine the endpoint. The indicator will change color to signify that the end of the titration has been reached. The color of the indicator is dependent upon the concentration of ions in the solution. An acid-base indicator is composed of a conjugate weak acid-weak base pair, where the two forms exhibit different colors depending on the pH of the solution. For an indicator, the acid ionization constant K_a is usually written as:

$$K_a = \frac{[H_3O^+][In^-]}{[HIn]}$$

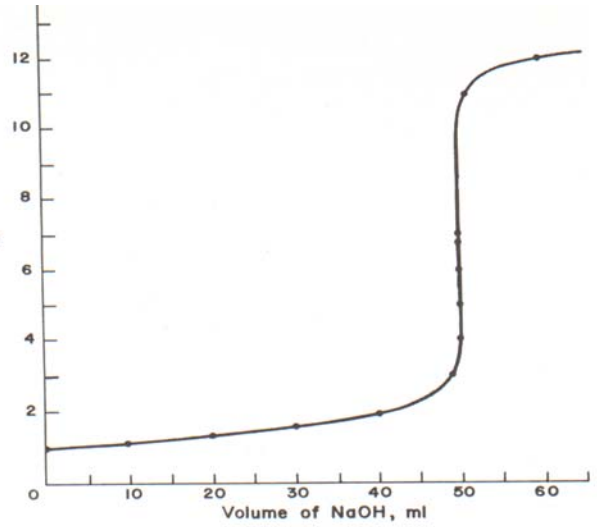
HIn is the acid form of the indicator and In^- is the base form. At the center of the change region, the ratio of $[In^-]$ to [HIn] is one, $[H_3O^+]=K_a$ and $pH=pK_a$. The color change region is usually ±1 pH unit around this point. Table 1 contains a list of some aqueous acid-base chemical indicators, as well as the pH range, the pK_a and the expected color (acid and base form). When choosing the proper indicator you should select one that has a pK_a as close to the endpoint of the titration.

When chemical indicators are not suitable, a potentiometric pH titration can also be used. The pH of the solution is plotted versus the volume of titrant added. Figure 4 shows a traditional strong acid-strong base titration curve. The graph shows the

Table 1

pH Range	Indicator	pKa	Acid Form	Base Form
0.0 - 1.6	Methyl Violet		Yellow	Blue
1.2 - 2.8	Thymol Blue	1.65	Red	Yellow
3.2 - 4.4	Methyl Orange	3.46	Red	Yellow
3.8 - 5.4	Bromocresol Green	4.90	Yellow	Blue
4.8 - 6.0	Methyl Red	5.00	Red	Yellow
5.2 - 6.8	Chlorophenol Blue	6.25	Yellow	Red
6.0 -7.6	Bromothymol Blue	7.30	Yellow	Blue
6.6 - 8.0	Phenol Red	8.00	Yellow	Red
7.4 -9.0	Metacresol Purple	8.30	Yellow	Purple
8.0 - 9.6	Thymol Blue	9.20	Yellow	Blue
8.2 - 10.0	Phenolphthalein	9.50	Clear	Pink
9.4 -10.6	Thymolphthalein		Clear	Blue
10.1 - 12.0	Alizarin Yellow R		Yellow	Red
11.4 - 12.6	Indigo Carmine		Blue	Yellow

Figure 4



volume of NaOH added to an acidic solution and the resulting pH of the solution. Note the abrupt change in the pH at the equivalence point.

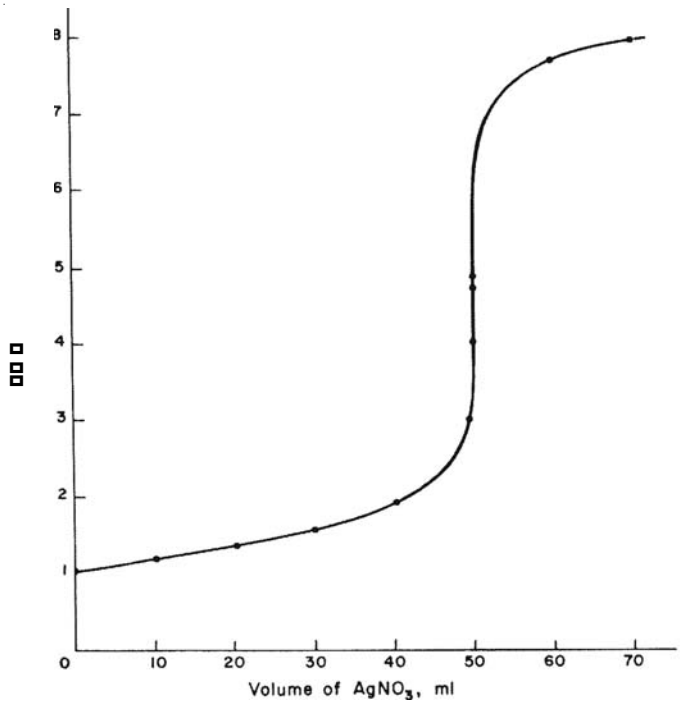
2.2.2 Argentometric Titrations

Argentometric titrations use silver (nitrate) as the titrant and are generally precipitation titrations, as many silver salts are insoluble. These titrations are commonly used to titrate and determine the concentration of bromide, chloride, cyanide, iodide, and sulfide.

Argentometric titrations can be done with Mohr's indicator (when all of the chloride has reacted, a red silver chromate precipitate is formed) or the titration can be easily followed with a silver ISE (or chloride ISE for chloride titrations) and a reference electrode.

Figure 5 shows the titration of 50 mL of 0.1N NaCl with 0.1N $AgNO_3$. The potentiometric signal is from a chloride ISE and is plotted as pCl (- log [Cl^-]).

Figure 5



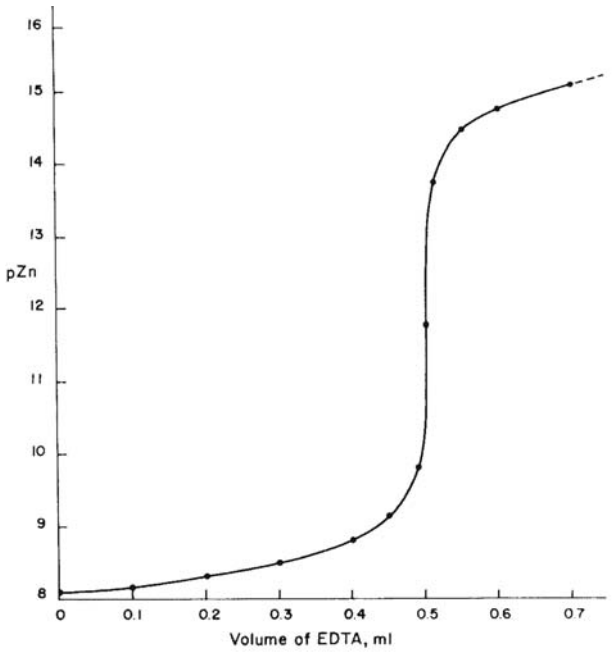
2.2.3 Complexometric Titrations

A complex is a species where a central metal ion is covalently bonded to one or more electron donating groups called ligands. In a complexometric titration, metal ions are titrated using a titrant that binds strongly to it. Often these titrants contain EDTA or CDTA, polydentate ligands that form very stable coordination compounds with metal ions. The complexation reaction must be fast in order to be useful for direct titration. Some metal ions react too slowly with EDTA for a direct titration.

An indicator electrode that responds to the metal ion can be used to monitor the titration progress. The titration curve will appear similar to a usual potentiometric titration. Complexation indicators change color at the endpoint as all metal ions are "consumed", or complexed, by the titrant.

The titration curve will appear similar to a potentiometric titration when using an indicator electrode that responds to the metal ion (see Figure 6).

Figure 6



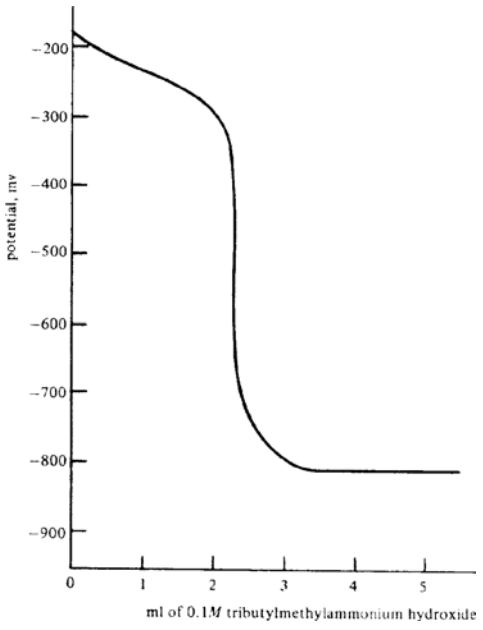
2.2.4 Ion Selective Titrations

The most popular ion selective titration is an acid-base titration. The hydrogen ion concentration is specifically measured and monitored during the titration process to locate the equivalence point. Using an ion selective electrode (ISE) as the indicator electrode, the potentiometric signal (in mV) is used to directly follow a specific ion's concentration (or activity).
Examples of ISE titrations include titrating fluoride with an aluminum titrant using a fluoride ISE, chloride with silver nitrate using a chloride ISE, sodium with a sodium ISE, etc. The equivalence point can be determined by plotting the mV value vs. the amount of titrant added.

2.2.5 Non-aqueous Solvent Acid-Base Titrations

Non-aqueous solvents must be used to titrate very weak acids and bases due to the inherent leveling effect water has on all acids and based dissolved in it. A wide variety of weak acids and bases can be titrated using non-aqueous solvents. Mixtures of acids or bases can often be individually analyzed in a single sequential titration.

Figure 7



Titration of Acids

Weak acids with pK_a's up to about 11 can be titrated in non-aqueous solvents. These include carboxylic acids, enols, phenols, imides, sulfonic acids, and inorganic acids. Water or lower alcohols are suitable for titrating medium to strong acids (pK_a less than 5). Titrating a weaker acid with a strong base titrant requires a solvent less acidic than water or ethanol/methanol. Solvents such as acetone, acetonitrile, t-butyl

alcohol, dimethylformamide, isopropanol and pyridine have been found to work well for acid-base titrations of strong, medium and weak acids/bases. Titrants include alcoholic potassium hydroxide and various sodium or potassium alkoxides in a 10:1 mixture of benzene/methanol. The best titrants are quaternary ammonium hydroxides (such as tetrabutylammonium hydroxide) due to good solubility of tetraalkylammonium salts of the titrated acids and the clean potentiometric titration curve obtained (see Figure 7).

Titration of Bases

Weak bases with pK_b's up to about 11, which do not ionize with water, can be titrated in non-aqueous solvents. These bases include aliphatic and aromatic amines, basic nitrogen heterocycles, alkali metal and amine salts of acids, and many other organic basic compounds. Titrating a weak base with a strong acid titrant requires a basic solvent that is as weak as possible. Water and alcohols allow the titration of medium strength bases such as aliphatic amines (pK_b = 4 to 5), but not the titration of weaker bases such as pyridine (pK_b = 8.8). Glacial acetic acid works well for weak bases and has been used extensively. Less basic solvents such as acetone, acetonitrile, and nitromethane extend the range of titrable compounds.

The endpoint for non-aqueous titrations are usually determined potentiometrically using a pH glass electrode, a modified calomel or double junction reference electrode with a low-flow rate reference junction. Good potentiometric titration curves are obtained in most solvents, except those with very low dielectric constants such as benzene, chloroform and others, when high electrical resistance of the solvent causes unstable potentials.

2.2.6 Precipitation Titrations

Precipitation titrations allow for faster analysis compared to the old gravimetric analysis, where a precipitate is formed, filtered, dried and weighed to analyze a compound. Typically silver halides, silver thiocyanate and a few mercury, lead, and zinc salts are titrated using this method. The chemical reactions must form an insoluble salt and precipitate out quickly in order to be analyzed by this method. When the reaction is not quick, a back titration can be used. A measured excess of the precipitating reagent (titrant) is added to force the reaction to occur, and then unreacted titrant is then titrated with a standard solution of another reagent.

2.2.7 Redox Titrations

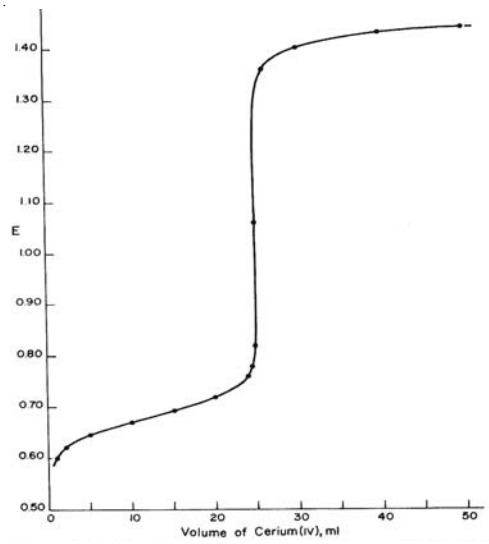
There are a number of oxidation-reduction reactions that can be used to determine unknown concentration by titration. If the reaction goes to completion, is fast and has an analytical signal available to follow it, a titration can be performed. The term “fast” means that each addition of titrant is reacted completely and the sensing electrode is able to detect the change in solution in less than one second.

Redox titrations are potentiometric titrations where the mV signal from a combination ORP (redox) electrode (usually with a platinum indicator electrode) is used to follow the reaction of oxidant/reductant. The electrode potential is determined by the Nernst equation and is controlled by the oxidant reductant ratio.

Visual indicators such as Ferrion are also available. The oxidized and reduced form of the indicator will have different colors and can be used to determine the end point.

Various reductants can be determined by titrants with oxidants such as potassium permanganate, potassium chromate or iodine. Commonly used reductants that are used as titrants include sodium thiosulfate, and ferrous ammonium sulfate.

Figure 8



As with Acid-Base titrations the potential changes dramatically at the equivalence point.

2.2.8 Karl Fischer Titrations

This method is based on a well-defined chemical reaction between water and the Karl Fischer reagent. The chemistry provides excellent specificity for water determination. The method can be used to determine free and bound water in a sample matrix. The Karl Fischer method is widely considered to produce the most rapid, accurate and reproducible results and has the largest detectable concentration range spanning 1 ppm to 100%.

The determination of water content is one of the most commonly practiced methods in laboratories around the world. Knowledge of water content is critical to understanding chemical and physical properties of materials and ascertaining product quality. Water content determination is conducted on many sample types including pharmaceuticals and cosmetics, foods and natural products, organic and inorganic compounds, chemicals, solvents and gases, petroleum and plastic products as well as paints and adhesives. The KF method is verifiable and can be fully documented. As a result, Karl Fischer titration is the standard method for analysis of water in a multitude of samples as specified by numerous organizations including the Association of Official Analytical Chemists, the United States and European Pharmacopoeia, ASTM, American Petroleum Institute, British Standards and DIN.

2.3 Titrations According to The Titration Sequence

2.3.1 Back Titrations

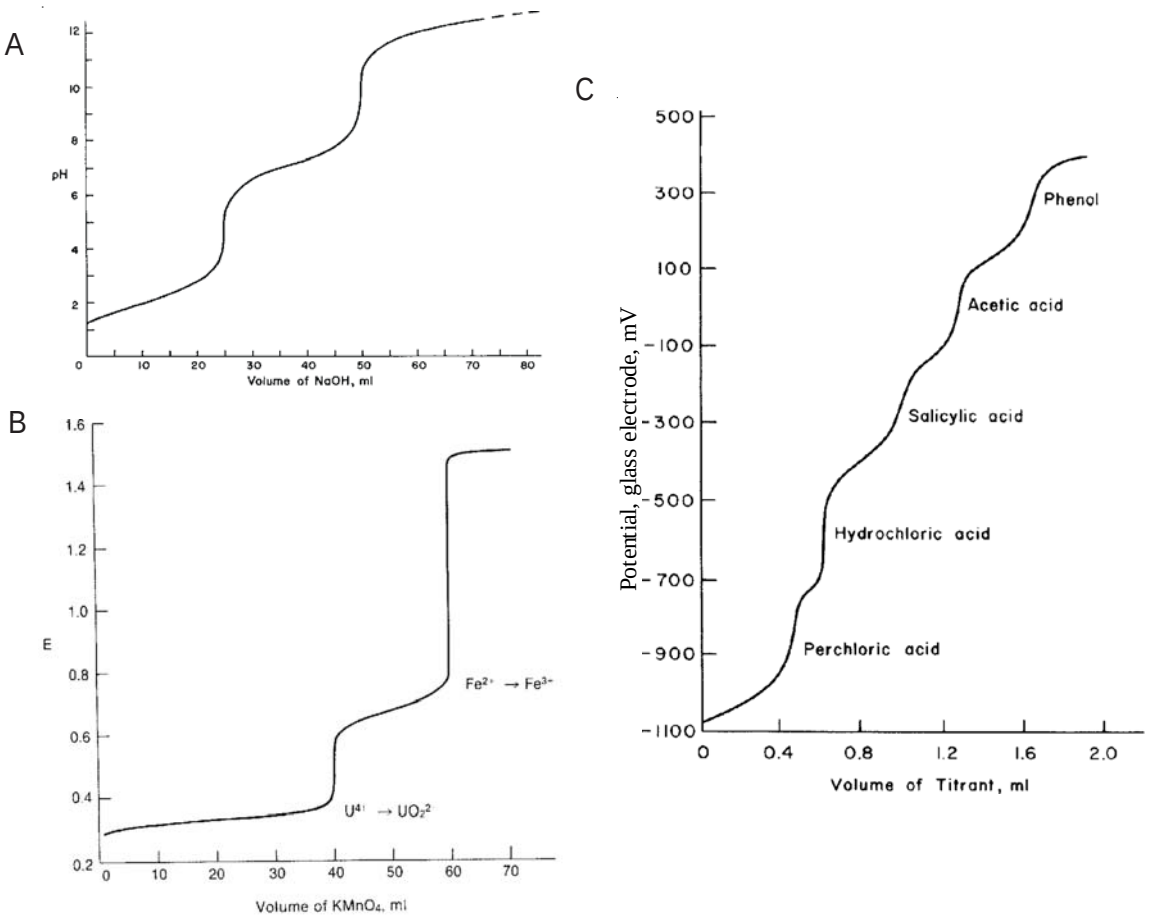
Back titrations are generally used when a reaction is too slow to be directly accomplished using a “direct” titration, where the reaction goes to completion within a few seconds. In a back titration, a large excess of a reagent is added to the sample solution, helping a slow reaction to go to completion. The unreacted, excess reagent is then titrated. The difference in the total volume of the first reagent added and amount determined from the second titration is the quantity of reagent required to complete the first reaction.

2.3.2 Multiple Endpoint Titrations

Under certain conditions, some titrations can exhibit more than one equivalence point and be titratable to the individual endpoints to determine the concentration of each individual component. Examples of these types of titrations include acid-base (where different strength acid or bases are in a mixture), redox (where each species has a different reduction potential), complexometric (where different species are separately titratable), and acid-base using polyprotic acids (the pK_a of the different protons varies enough to separate them).

Figure 9 shows three different types of multiple endpoint titrations. “A” shows the titration of a polyprotic acid. The different acid strengths of the first and second proton can be determined. “B” illustrates a mixture of two different metal redox species, where the different redox potentials allow the species to be separated. “C” is the titration of a solution containing strong, weak, and very weak acids.

Figure 9



3 INTRODUCTION TO TITRATION APPARATUS AND TYPICAL TITRATION PROCEDURE

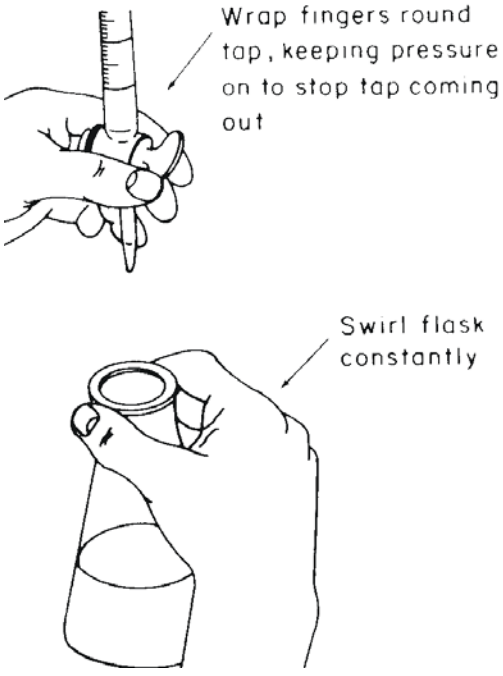
3.1 Manual Titration

Apparatus required for manual titration include:

- Volumetric Burette, for precisely controlled delivery of titrant to the reaction vessel
- An Erlenmeyer, or similar flask, that facilitates constant mixing or swirling required to ensure solution homogeneity
- Volumetric pipettes for the precise addition of samples and indicator solutions
- Titrant solutions of known concentration
- A visual or instrumental indicator for detecting the completion of the reaction

A typical manual titration consists of the following steps:

1. A volumetric pipette is typically used to add a known volume of sample to the flask
2. An indicator solution or instrument probe is added to the flask
3. A burette is used to measure the addition of titrant to the flask and dispense titrant in a controlled manner
4. Titrant is added via the burette until the method indication signals the reaction endpoint
5. The concentration of analyte is calculated based on the concentration and volume of titrant required to reach the endpoint



3.2 Automatic Titration

Automatic titrators are high-precision analytical instruments that deliver the titrant, monitor the physical change associated with the titration reaction, automatically stop at the endpoint and calculates the concentration of the analyte. Automatic titrators are best for repetitive titrations and high-accuracy analyses.

An automatic titrator must have an accurate liquid dispensing system. In high accuracy systems like the HI 900-series titrators, the liquid dispensing system consists of a stepper-motor driven piston syringe burette capable of accurately and precisely dispensing very small volumes of titrant, a valve system to switch between titrant intake and outlet and a dispensing tip. These three main subsystem components must be as accurate as possible, with very low gear backlash in the burette pump, minimal piston seal flexing, precision ground inner diameter of the glass syringe, a low dead volume valve, minimal evaporation/permeation, and chemically resistant tubing.

Apparatus required for automatic titration include:

- An automatic titrator, equipped with a burette
- A beaker
- An electronic stirring system, either a propeller stirrer or a magnetic stir bar and stir plate
- Volumetric pipettes for the precise addition of samples
- Standard titrant solutions of known concentration
- An electrode system that can be used to determine the endpoint of the titration

A typical automatic titration consists of the following steps:

1. Set up the automatic titrator according to the manufacturer's instructions
2. A volumetric pipette is typically used to add a known volume of sample to the beaker
3. Submerge the propeller stirrer or add the stir bar to the beaker, and turn on
4. Start the titration, the titrator will automatically stop at the endpoint and determine the concentration of the analyte

4 TITRATION RESULTS

4.1 Accuracy

The factors most critical to achieving accurate results with the HI 900 titration systems are the concentration of the sample, size of the sample and having an optimized set of method parameters.

4.2 Repeatability

Repeatability, or the agreement between replicate determinations, is expressed quantitatively as the relative standard deviation (RSD).

4.3 Sources of Error

One of the advantages of volumetric analysis is excellent accuracy and precision. The sources of error can be grouped into sampling, titrant and standards, chemical reactions, endpoint determination and calculations.

4.3.1 Sampling Errors

- Selection of a non-homogeneous or non-representative sample
- Sample changed or was contaminated during collection, storage or transfers
- Poor technique when transferring sample to beaker or flask
- Errors in the balance, calibrate and check balance regularly

4.3.2 Errors with Titrant and Standard

4.3.2.1 Preparation Errors

Incorrect preparation due to:

- Poor technique in weighing the salt or when transferring to volumetric glassware
- Low-purity of salts or water used to make titrant and standard
- Dirty or wet glassware
- Improper storage of titrant or standard which allows water gain, evaporation or deterioration
- Failure to standardize frequently to adjust for change in titrant
- Failure to flush titrator tubing with a volume of titrant before standardizing
- Volume errors from pipettes and volumetric flasks, grade A glassware is required
- Balance errors when weighing out salts, calibrate and check balance regularly

4.3.2.2 Dispensing Errors

Incorrect dispensing due to:

- Dead valve volume and leaking valve
- Inaccuracy in motor drive and gear lash/ backlash
- Poor burette/ piston seal
- Non-uniform diameter of burette glass cylinder
- Chemical incompatibility with tubing or bubble generation
- Density/ temperature changes in titrant

4.3.3 Chemical Reaction Errors

- Inappropriate solvent or sample resulting in side reactions
- Poor mixing of the titrant and solvent or sample in the titration vessel
- Reaction between titrant and sample is not rapid
- Reaction does not go to completion
- Reaction has side reactions

4.3.4 Endpoint Determination Errors

Most manual titrations use a visual indicator to indicate when the endpoint is reached and the titration should be stopped. Automatic titrators use instrumental methods to determine the end of a titration and the equivalence point. There are two predominant methods used to determine the equivalence point, first derivative and second derivative.

The inflection point of the titration curve (mV vs. Volume) is normally assumed to be the equivalence point. The first derivative is often used to determine the inflection point. The maximum value of the first derivative (dmV vs. dV) corresponds to the theoretical equivalence point. During a titration it is rare to have a data point exactly at the first derivative maximum, the maximum value is determined by interpolating the first derivative data points.

The second derivative (d^2 mV vs. dV^2) can also be used to determine the equivalence point, and can offer advantages over the first derivative method. Second derivatives have increased sensitivity to smaller inflection points and easier numerical evaluation of the actual equivalence point. The value where the second derivative is equal to zero is the equivalence point. The second derivative requires fewer points located near the equivalence point, where data is often not obtained or not as reliable.

Errors in determining the endpoint can result from:

- Incorrect signals from the sensor
- Sensor drift
- Sensor or instrument has slow response, keep sensors in good condition
- Inappropriate setting on the titrator

5 CALCULATIONS

The main variables used in calculating a result from a titration are the sample volume, the concentration of the titrant, and the volume of titrant required to reach the equivalence point. At the equivalence point, an equal number of equivalents of the analyte and titrant has been added.

5.1 Sample Calculation

By Mass

$$C\ sample = \frac{V\ titrant \times C\ titrant \times Ratio \times FW\ analyte}{m\ sample} \times 100$$

C sample	Sample Concentration (g/100g)
V titrant	Volume of titrant (L)
C titrant	Titrant Concentration (eq/L)
Ratio	Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
FW analyte	Formula Weight of the Analyte (g/mol)
m sample	Mass of sample (g)

By Volume

$$C\ sample = \frac{V\ titrant \times C\ titrant \times Ratio \times FW\ analyte}{V\ sample} \times 100$$

C sample	Sample Concentration (g/100mL)
V titrant	Volume of titrant (L)
C titrant	Titrant Concentration (eq/L)
Ratio	Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
FW analyte	Formula Weight of the Analyte (g/mol)
V sample	Volume of Sample (mL)

5.2 Standardize Titrant

Titrant standardization is the second most important calculation in titrations. A primary standard is titrated in order to determine the concentration of the titrant. This is essentially a typical titration calculated in “reverse”, where the concentration of the solution is known and the titrant is the unknown.

By Mass

$$C\ titrant = \frac{m\ standard \times Ratio}{FW\ standard \times V\ titrant}$$

C titrant	Titrant Concentration (N)
m standard	Mass of Standard (g)
Ratio	Equivalence ratio of titrant/standard (eq titrant/ mol standard)
FW standard	Formula Weight of the Standard (g/mol)
V titrant	Volume of Titrant (L)

By Volume

$$C\,titrant = \frac{V\,standard \times (1\,L/1000\,mL) \times C\,standard}{V\,titrant}$$

C titrant	Concentration of titrant (N)
V standard	Volume of Standard (mL)
C standard	Concentration of standard (eq/L)
V titrant	Volume of Titrant (L)

5.3 Blank Titration

In a blank titration a pre-titration is performed, often times on the solvent to be used for the sample titration, and the titrant volume required to reach the endpoint is noted. This blank value nullifies error due to titrant required to react with the components of the titration solution matrix. The basic titration equation can be used for a blank titration, with the single modification that the volume of titrant used in the blank titration should be subtracted from the regular titration titrant volume.

$$C\,sample = \frac{C\,titrant \times (V\,sample - V\,blank) \times Ratio \times FW\,analyte}{m\,sample} \times 100$$

C Sample	Sample Concentration (g/100g)
C titrant	Titrant Concentration (eq/L)
V sample	Volume of Titrant required for the sample (L)
V blank	Volume of Titrant required for the blank (L)
Ratio	Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
FW analyte	Formula Weight of the Analyte (g/mol)
m sample	Mass of sample (g)

5.4 Multiple Endpoint Titration

Some titrations have two or more endpoints, each corresponding to the equivalence point for a specific reaction. Multiple endpoint titrations are similar to a blank titration in that the volume of titrant required to reach the first endpoint is subtracted from the titrant volume used to reach the next sequential endpoint.

$$C\,sample\,1 = \frac{V\,titrant\,1 \times C\,titrant \times Ratio \times FW\,analyte\,1}{m\,sample} \times 100$$

$$C\,sample\,2 = \frac{(V\,titrant\,2 - V\,titrant\,1) \times C\,titrant \times Ratio \times FW\,analyte\,2}{m\,sample} \times 100$$

$$C\,sample\,3 = \frac{(V\,titrant\,3 - V\,titrant\,2) \times C\,titrant \times Ratio \times FW\,analyte\,3}{m\,sample} \times 100$$

C sample1	Sample 1 Concentration (g/100g)
C sample2	Sample 2 Concentration (g/100g)
C sample3	Sample 3 Concentration (g/100g)
V titrant 1	Volume of titrant required to reach the first end point (L)
V titrant 2	Volume of titrant required to reach the second end point (L)
V titrant 3	Volume of titrant required to reach the third end point (L)
C titrant	Concentration of titrant (N)
Ratio	Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
FW analyte 1	Formula Weight of the Analyte 1 (g/mol)
FW analyte 2	Formula Weight of the Analyte 2 (g/mol)
FW analyte 3	Formula Weight of the Analyte 3 (g/mol)
m sample	Weight of Sample (mL)

5.5 Back Titration

The equation used in back titration calculations is also similar to the equation for a blank titration. Instead of subtracting the initial amount of titrant needed to react with the blank, the amount of second titrant needed to react with the excess titrant added in the first titration is subtracted from the amount of the first titrant added. The difference between the two amounts is the amount of titrant necessary to reach the first equivalence point.

$$C\ sample = \frac{(C\ titrant\ 1 \times V\ titrant\ 1 - C\ titrant\ 2 \times V\ titrant\ 2) \times Ratio \times FW\ analyte}{V\ sample} \times 100$$

C sample	Sample Concentration (g/100mL)
C titrant 1	Concentration of titrant 1 (N)
V titrant 1	Volume of titrant 1 (L)
C titrant 2	Concentration of titrant 2 (N)
V titrant 2	Volume of titrant 2 (L)
Ratio	Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
FW analyte	Formula Weight of the analyte (g/mol)
V sample	Volume of sample (mL)

6 GLOSSARY

Acid
A chemical species that can donate one or more protons (hydrogen ions).

Acid-Base Titration
Stoichiometric neutralization titrations, based upon the reaction that occurs between an acid and base.

Activity
A physical property corresponding to the concentration of all ions in a solution. Electrodes respond to activity.

Amperometric Titration
Titrations where the current flow between two electrodes (often a metal electrode and a reference electrode) are used to monitor the titration progress.

Analyte
The chemical species being measured in a titration.

Argentometric Titration
Titrations that use silver (nitrate) as the titrant. These titrations are typically precipitation titrations.

Automatic Titrator
An instrument designed to automatically carry out a titration. It will add the appropriate amount of titrant, determine the endpoint and calculate the results.

Back Titration
A type of titration where an excess amount of titrant is added to a sample, forcing a sluggish reaction to go to completion. The excess reagent is then “back” titrated with a second titrant.

Base
A chemical species that can accept one or more protons (hydrogen ions).

Biamperometric Indication
Uses a double platinum pin electrode to measure the current flow through a titration solution.

Bivoltametric Indication
Uses a double platinum pin electrode to measure the voltage required to maintain a constant current flow through a titration solution while constant voltage is applied across the platinum elements of the electrode.

Burette
A graduated cylindrical piece of laboratory glassware that is used to dispense precise amounts of solution.

Complex Ion
A species where a central metal ion is covalently bonded to one or more electron donating groups called ligands.

Complexometric Titrations
Metal ions are titrated using a titrant that binds strongly to it. The titrants often contain Ethylenediaminetetraacetic Acid (EDTA) or Cyclohexylenedinitrilotetraacetic Acid (CDTA).

Endpoint

The point were a titration is stopped because a physical change in the solution has indicated a completed titration. Titration endpoints typically coincide with the equivalence point. A fixed value endpoint (pH or mV) can be used as well. The titration will stop at the desired point regardless if the titration is complete.

Equivalence point

The point where the quantity of titrant is stoichiometrically equal to the quantity of analyte.

Formal

The theoretical number of equivalents per liter of the solution. It is used in solutions where the exact concentration of a species may be affected by the other ions present, therefore the stated concentration may not be exactly correct.

Gravimetric Analysis

A quantitative determination of an analyte based on the mass of the solid.

Indicator Electrode

An electrode that responds to the species of interest. The electrode potential is proportional to the concentration or activity of that ion in the solution being measured.

Indicators

Chemical indicators are typically organic dyes that change form under different physical conditions, causing a color change that can be seen by an analyst. Typically used in manual titrations, chemical indicators have been replaced with electrometric indicators, which are used with automatic titrators.

Inflection Point

The point on a titration curve were the second derivative curve changes signs.

Ion Selective Electrode (ISE)

An electrode that responds to a specific ion. The electrode potential is proportional to the concentration or activity of that ion in the solution being measured.

Karl Fischer Titration

A titration that uses a chemical reaction that is specific for determining water.

Manual Titration

A titration that is carried out by hand. The analyst must add the appropriate amount of titrant, determine the endpoint and calculate the results.

Molar

The concentration of a solute in a solution.

Mole (mol)

A quantity of a chemical species. The molecular weight of a substance in grams is equal to the mass of one mole of the substance. One mole is equal to 6.022×10^{23} atoms or molecules.

Monochromator

A device that allows only a narrow range of wavelengths to pass though it by separating the light into different wavelengths.

Multiple Endpoint Titration

A titration that reacts multiple species in solution sequentially using the same titrant. The concentration of each analyte can be determined from their respective endpoints.

Nernst Equation

The fundamental equation relating cell voltage to the concentration of a solution.

Neutralization

A chemical reaction where an acid and a base react to form a neutral salt and water.

Non-aqueous

A solution that does not contain water.

Non-aqueous Titration

A titration that is preformed in non-aqueous solutions, typically used to titrate very weak acids and bases to eliminate the leveling effect water has on all acids and bases dissolved in it.

Normal

The concentration of a solution which accounts for any stoichiometric difference between the various species in a solution.

Oxidation / Reduction Potential (ORP)

The measurement describing whether a species wants to donate or accept electrons from other species in a redox reaction. If a solutions reduction potential is higher than the species it is reacting with, it will typically gain electrons or be reduced. If the potential is lower than the species it is reacting with, it will typically lose electrons or be oxidized.

Oxidant

The species that is accepting electrons in a redox reaction.

Pipette

Scientific apparatus that is used to deliver precise volumes of liquids.

Polyprotic Acid

Acids that are capable of donating more than one proton per acid molecule.

Potentiometric Titration

A titration in which the endpoint is determined by monitoring the voltage of the solution using an electrode.

Precipitation Titration

A titration in which the analyte reacts with the titrant to form an insoluble compound. The endpoint is typically detected with an ISE sensitive to either the analyte or titrant.

Reagent

The chemical added in a titration that causes the given reaction to occur.

Reduction-Oxidation Reaction (redox)

A chemical reaction in which the atoms involved in the reaction have their oxidation numbers changed. Reduction is the gain of electrons, which decreases the oxidation number. Oxidation is the loss of electrons, which increases the oxidation number.

Reductants

The electron donor in a redox reaction.

Reference Electrode

An electrode that supplies a constant electrode potential. It is used in combination with an “indicator” electrode, allowing for the “indicator” electrode potential to be measured.

Relative Standard Deviation (RSD)

A measure of the amount of relative variation in a set of data. It is calculated by dividing the standard deviation by the mean: RSD = (Standard Deviation of X) * 100 / (Mean of X)

Repeatability

The variation in sample measurements taken by a single person or instrument under the same conditions.

Spectrophotometric Titration

A titration in which the endpoint is marked by a change in the color and/or color intensity.

Stoichiometry

The quantitative relationship of the reactants and products in a chemical reaction.

Titrant

The chemical added in a titration that causes the given reaction to occur.

Titration

A quantitative, volumetric procedure used in analytical chemistry to determine the concentration of an analyte in solution. The concentration of the analyte is determined by slowly adding a titrant to the solution. As the titrant is added, a chemical reaction between the titrant and the analyte occurs.

Titration Curve

A graph containing the physical data obtained for a titration. The data plotted is often an independent variable (volume of titrant) vs. a dependent variable (pH of the solution). From the titration curve, the equivalence point or endpoint can be determined.

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